



Micro- and mesozooplankton respiratory metabolism: comparison of simultaneous incubation experiments and ETS activity.

By Miguel Alcaraz^{1*}, Theodor T. Packard¹, Enric Saiz¹, Albert Calbet¹, Isabel Trepat¹ and Alf Skovgaard²

Group of Zooplankton Ecology. Institut de Ciències del Mar, CSIC. P. Marítim de la Barceloneta 37-49, 08003 Barcelona (Spain). ² University of Copenhagen, Department of Biology. Øster Farimagsgade 2D, DK-1353 Copenhagen K, Denmark

* e-mail: miquel@icm.csic.es

1. Introduction

The contribution of the different plankton components to the Ocean respiratory carbon losses is of prime importance for the improvement of models dealing with global climate changes and plankton production.

However, the obtention of plankton respiratory losses for the separate groups by classical incubation methods is extremely time- and labour consuming.



Enzymatic methods like the activity of the respiratory electron transport system (ETS) overcome most of the experimental inconveniences, but gives potential, not actual respiration rates.

In order to use ETS as valid estimator of respiration rates, it is necessary to determine wheter there exist robust relationships between ETS activity and directly measured O_2 consumption.

2. Aims

1.- The obtention of robust relationships between directly measured O₂ consumption rates and ETS activity for different zooplankton components.

2.- To ascertain whether the relationships between respiration rates and ETS for different zooplankton size-fractions are also different and can be used as a proxy for respiration rates.

3. Methods

The study was based in the comparison of O_2 consumption rates obtained by classical incubation methods with those obtained from ETS activity in the incubated organisms.

Experimental organisms

The study included three groups of experimental zooplankton



Fig. 5.- Relationships betwee RR and ETS for the different grups and for all the data pooled. For > 200 µm zooplankton and pooled data, a line with an asterisc means the corresponding equation after the high value point has been removed.

Table II.- Table II.- Average ETS/RR ratios; standard deviation (SD), number of data (N) and authors: 1) King and Packard (1975); 2) Alcaraz and Packard (1989); 3) Bamstedt (1980). The numbers in red are potential respiration rates (ETS) lower than those measured.

Froups	ETS/RR	SD	Ν	Authors
lixed copepods	0.46	0.09	6	1
tenophores	0.69	0.19	15	1
ledusae	1.19	1.13	36	1
lixed zooplankton	0.65	0.25	7	1
lixed zooplankton	0.39	0.11	6	2
cartia tonsa	1.25	0.42	5	3
cartia tonsa	1.75	1.45	5	3
cartia tonsa*	2.27	0.23	5	3
lixed zooplankton	2.46	1.35	18	This study
lixed microzooplankton	1.75	0.66	17	This study
Dithona Nauplii II-IV	2.07	0.59	10	This study
atal (nacled data)	2 00	1 75	1 E	

Table I.- Regression eaquations, determination coefficients (r²), and number of data (n) for the different zooplankton groups and for all grouped.

Group	Equation	R^2	n	
200 µm mixed ooplankton	RR = 0.28 ETS + 1.05	0.93	18	
200 µm mixed cooplankton *	RR = 0.36 ETS + 0.29	0.88	17	
200, > 50 µm mixed ooplankton	RR = 0.441 ETS + 0.65	0.91	17	
auplius II-IV Oithona davisae	RR = 0.45 ETS + 0.44	0.92	10	
ooled data	RR = 0.31 ETS + 1.46	0.87	45	
ooled data *	RR = 0.41 ETS + 0.51	0.88	44	

me as the above with the largest value point removed

organisms (Fig. 1):

 >200 µm mixed zooplankton (mesoplankton) from 3 stations in the NW Mediterranean in March 1999 (Alcaraz et al. (2007). Obtained by 100 - 0 m depth vertical net hauls made with a 200 µm-mesh WP-2.



• <200 >50 µm mixed zooplankton (metazoan microzooplankton) from a station 1 nM offshore Barcelona. Samples obtained from november 2003 to january 2004. 40 – 0 m depth vertical hauls made with a 50 µm-mesh net...

• **Copepod nauplii**. (nauplii II-IV) obtained from a laboratory culture of the cyclopoid Oithona davisae maintained by the Zooplankton Ecology Group in the Institute of Marine Sciences.

Fig. 1.- The three experimental zooplankton groups: A: Mixed mesozooplankton; B: Mixed microzooplankton; C Oithona davisae nauplii.

Direct measurements of respiration rates:

O₂ consumption rates were measured by 24 h-duration incubation experiments at "in situ" temperature as described in Alcaraz et al. (1998). In most of the experiments, semi-continuous analysis of dissolved O_2 concentration were obtained with pulsed polarographic O_2 electrodes (ENDECO®, Fig. 2). Time - O_2 concentration relationships were fitted by linear regression (Fig. 3). In few cases, the incubation was made in BOD bottles and the O_2 concentration measured by Winkler titration (Fig. 4).

ETS activity:

After the incubations, the experimental organisms were transferred by filtration into GF/A glass fibre filters, immediately frozen in liquid N₂ and maintained at -80 °C until the analysis of the ETS activity. These were performed according to Packard et al. (1996).

* Winter values

ETS/RR ratios (Table II) were similar to those obtained by Bamstedt (1980) for Acartia tonsa. The values ranged from 1.75 to 2.46, with an average value for pooled data of 2.09. These values contrast with previous ones in which ETS gave potential respiration rates lower than those directly measured (seeTable II). The reason could be the improvement of the analysis of ETS activity by the two substrates method (Packard et al. 1996).

The relationship between **calculated** and **measured** respiration rates, using the **regression equation** for the pooled data, and the average ETS/RR ratio explain the same percentage of the variance, 88 % in both cases (Fig. 6).



respiration rates for the threegroups of zooplankton (pooled data). A: Equations for pooled data as in Table I. B: ETS/RR ratios (Table II).

This relatively good agreement obtained between observed and calculated respiration rates (Fig. 6), both using the **regression** equation for pooled data (A) and the average ETS/RR ratio (B), could be due to the relatively homogeneous environmental conditions (mainly temperature and food) experienced by all the zooplankton groups before catching them and during the incubation experiments.

5. Conclusions

•The ETS-RR regression equations and ETS/RR ratios for the different zooplankton





Fig. 2.- Incubation flasks showing the electrode and the pressure control



components appear to be relatively constant.

• At least when the experimental conditions (mainly temperature and food) are similar, both regression equations and ETS/RR ratios are robust predictors of actual respiration rates (RR). The variance explained is in both cases up to 88 %.

• The factors affecting the variability of the relationships between ETS-derived and directly measured RR must be ascertained in order to efficiently use ETS activity as a proxy of respiration rates.



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