

Biomarkers of fitness and welfare in dairy cattle: healthy productivity

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Invited Review

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Abstract

Milk production intensification has led to several unwanted aspects, such as sustainability issues and environmental pollution. Among these, increased milk outputs that have been achieved over the last 70 years have led to several health and pathophysiological conditions in high yielding dairy animals, including metabolic diseases that were uncommon in the past. Increased occurrence of diverse metabolic diseases in cattle and other domestic animals is a key feature of domestication that not only affects the animals' health and productivity, but also may have important and adverse health impacts on human consumers through the elevated use of drugs and antibiotics. These aspects will influence economical and ethical aspects in the near future. Therefore, finding and establishing proper biomarkers for early detection of metabolic diseases is of great interest. In the present review, recent work on the discovery of fitness, stress and welfare biomarkers in dairy cows is presented, focusing in particular on possible biomarkers of energy balance and oxidative stress in plasma and milk, and biomarkers of production-related diseases and decreased fertility.

Introduction

This is a companion article to our recent review of biomarkers related to the stress response to environmental perturbations in dairy animals (Almeida *et al.*, 2019), and deals with biomarkers related to metabolic health and productivity, including fertility.

Most of the milk and dairy derivatives consumed in the developed world are produced in intensive production systems. Such systems are typically based on one breed: the Holstein Friesian dairy cow. The yields for this breed have been increasing steadily for the last 70 years, reaching up to average yields of 20 000 kg milk/per 305-d lactation. Such extraordinary increases were obtained mainly thanks to advances in genetics, nutrition and feeding, reproductive management, artificial insemination, health management, environmental control and milking parlour design and efficiency: Roche *et al.* (2017), Douphrate *et al.* (2013) and Duncan *et al.* (2013). Conversely, in tropical countries, different dairy species and breeds are raised, particularly water buffalo (*Bubalus bubalis*) and *Bos indicus* cattle. In addition, these production systems are not as intensified as those in temperate countries. Furthermore, small ruminants (sheep and goats) and dromedaries also contribute to the total amount of milk produced worldwide (Medhammar *et al.*, 2012). Despite being a small proportion of the world's dairy output, this sector has been growing steadily, particularly in industrialized countries (Pulina *et al.*, 2018). For more information, refer to the positioning paper by Hernández-Castellano *et al.* (2019).

The dairy sector is facing numerous challenges (Baumgard *et al.*, 2017; Boor *et al.*, 2017; Martin *et al.*, 2017; McGuffey, 2017; Polsky and von Keyserlingk, 2017; Tan *et al.*, 2017). On the one hand, it is vital to keep improving and optimizing production levels, particularly in the classical areas of animal production (nutrition, lactation physiology, reproductive biology and health management) as well as in relation to dairy farm facilities and milking parlour design and automation. On the other hand, such intensification processes have led to several undesirable aspects that are frequently associated with dairy production, such as perceived sustainability issues and environmental pollution. We shall not enter into that debate in this review, but we do recognize its importance. Another consequence of intensification is increased occurrence of several metabolic diseases that were not common in the past. Metabolic diseases in cattle and other domestic animals are a key feature of domestication

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that not only affects the animals' health and productivity, but also has negative consequences on human consumers through elevated use of drugs and antibiotics (Raboison *et al.*, 2016). Such metabolic diseases include, for instance, ruminal acidosis, mastitis, ketosis and laminitis, among others. Reducing the occurrence of these diseases will be a major research area in the near future. Another aspect of dairy production concerns animal welfare and how this is compromised by intensification. This is a topic of growing importance for consumers, who place particular emphasis on animal transport, reduced individual space in barns, separation of dam and calf and the increased occurrence of the aforementioned metabolic diseases. These will also be aspects of growing economical and ethical importance for the dairy sector in the coming years. Therefore, finding and establishing proper biomarkers of welfare will be of great interest in the near future.

The term 'animal fitness' refers to the ability of one individual, relative to others, to leave viable offspring. Over the last few decades, dairy cow breeding programmes have focused mainly on characters and traits related to increased milk yield with negative consequences for cow fitness (Essl, 1998). Further aspects about cow fitness can be found in our companion paper (Almeida *et al.*, 2019). Quantitative evaluations of fitness traits such as lameness and mastitis resistance, calving interval and lifespan show that, because of antagonistic relationships between production and these fitness traits, a trade-off may exist between the costs of lower milk yield and the benefits of better cow health status (Koolhaas and van Rennen, 2016). Therefore, identifying specific biomarkers related to fitness is of great interest to ensure optimal fitness in modern dairy cows

Based on these observations, the challenge for dairy researchers is to establish well-based, empiric and quantifiable biomarkers for metabolic diseases, welfare, fitness and wellbeing in dairy cows, and this challenge is the focus of the present review (Fig. 1).

Biomarkers of energy balance and oxidative stress

Due to the sudden demand of energy for milk production, the transition from late pregnancy to lactation represents an important metabolic challenge for modern dairy cows. During this period, energy intake does not meet energy requirements for body maintenance and milk production, which results in negative energy balance (NEB; Bell and Bauman, 1997; Drackley, 1999) and high adipose tissue mobilization. If adaptation to NEB fails, the risk of metabolic disorders such as ketosis, hypocalcaemia, fatty liver mastitis and others increases considerably. In addition, NEB is related to lower conception rates, early embryonic mortality, and silent oestrus in high yielding dairy cows.

Traditional ways for detecting or preventing NEB are based on blood metabolites (i.e. non-esterified fatty acids, NEFA), and body condition score (BCS; a subjective score of body fattening), all of which require complicated data collection (individual feed intake and body weight), or invasive and laborious blood collection, along with trained staff. Hence, there is a need for accurate, objective and preferably non-invasive biomarkers that indicate energy status in dairy cows postpartum. In this section, potential biomarkers of NEB in blood and milk of dairy cows will be discussed. These are by far the most important media for current analytical approaches, but it is worth pointing out that other tissues and/or fluids might also prove to have value in the future, for instance, a recent report has examined possible metabolic biomarkers in hair samples (Möller *et al.*, 2019).

Biomarkers in blood for negative energy balance diagnosis

Non-esterified fatty acids (NEFA) and β -hydroxybutyrate (BHBA) are probably the most-known blood parameters used to assess NEB in dairy cows. Blood NEFA concentrations reflect the extent of fat mobilization, while BHBA indicates fat oxidation in the liver. Therefore, both analytes have been extensively used in the field as indicators of NEB (McArt *et al.*, 2013; Ospina *et al.*, 2013). Elevated concentrations of NEFA and BHBA in blood have been shown to be associated with reduced milk yield (Duffield *et al.*, 2009; Ospina *et al.*, 2010; Chapinal *et al.*, 2012), and impaired periparturient immunity and increased risk of infectious diseases (Moyes *et al.*, 2009; Ospina *et al.*, 2010). NEFA concentrations higher than 0.3 and 0.6 mmol/l pre- and postpartum, respectively, are associated with an increased risk of displaced abomasum, clinical ketosis, retained placenta and metritis (Ospina *et al.*, 2010).

In addition to NEFA, specific fatty acids (FA) in blood could be used as potential and alternative biomarkers for NEB in transition cows. Imhasly *et al.* (2015) examined changes in the blood plasma lipidome in transition dairy cows and found that the levels of a number of triacylglycerides (TGs) were higher prepartum than postpartum: TG 48:3, TG 48:1, TG 49:2, TG 49:1, TG 50:4, TG 50:3, TG 50:2, TG 51:3, TG 51:2, TG 51:1, TG 52:4, TG 52:3, TG 53:3, TG 54:6 and TG 56:6. In addition, the levels of two fatty acid amides (i.e. linoleamide and anandamide) decreased only at calving (Imhasly *et al.*, 2015), which suggests enhanced energy requirement postpartum. In contrast, the levels of lyso-phosphatidylcholine (LPC) and phosphatidylcholine (PC), specifically: LPC 16:0, LPC 18:3, LPC 18:2, LPC 18:1, LPC 20:5, PC P-34:2, PC P-36:5, PC P-36:4 and PC 36:6, as well as the sphingomyelins 39:1 and 43:3 were increased postpartum (Imhasly *et al.*, 2015). However, in this study the relationship between these lipids and individual NEB was not analysed. Therefore, it is not clear whether NEB might alter the blood lipidome in cows.

In postpartum cows that are in NEB, increased inflammatory markers are found in plasma, such as tumour necrotizing factor alpha, the acute phase proteins haptoglobin, serum amyloid A and others (Bradford *et al.*, 2015). These inflammatory markers can also be used as indicators of the degree of NEB, since cows with severe NEB have a higher degree of systemic inflammation. This topic is reviewed thoroughly in Bradford *et al.* (2015).

Biomarkers in milk for negative energy balance diagnosis

The potential of milk biomarkers for NEB diagnosis is enormous, as sensors for these specific biomarkers could be implemented in milking parlours and milking robots to provide individual information about energy status in cows. BHBA, for instance, can be measured in milk, and has the potential to be measured frequently in individual cows in early lactation as an indicator of NEB (Duplessis *et al.*, 2019).

Fatty acids

Milk FA may be used as biomarkers of EB in dairy cows. Milk FA are derived from four major pathways (1) directly from the diet, (2) de novo synthesis in the mammary gland, (3) formation in the rumen by biohydrogenation or bacterial degradation, and (4) fat depots (Stoop *et al.*, 2009). Changes in energy status across lactation also imply changes in milk FA composition (Gross *et al.*, 2011). In cows under NEB, the de novo synthesis of fatty acids by

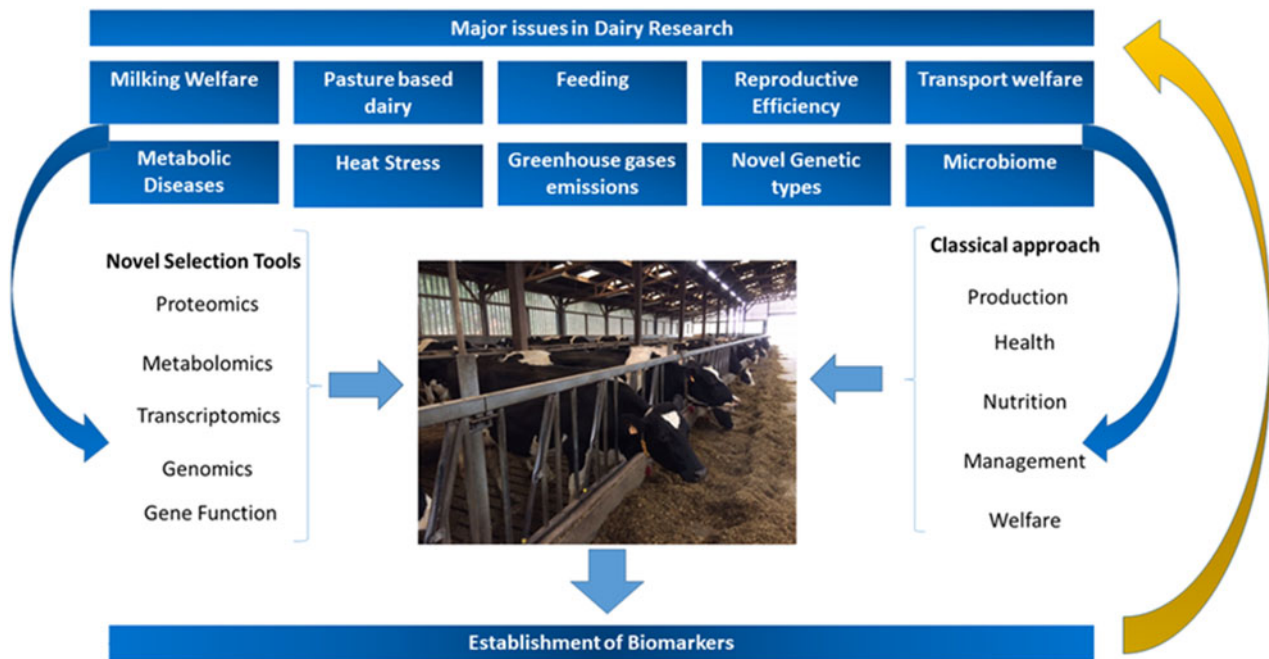


Fig. 1. Major challenges and areas of research in modern dairy production systems and how to address them, highlighting the importance of classical and novel selection tools as well as the establishment of biomarkers.

the mammary gland (i.e. C6:0 to C14:0) is reduced in favour of increased body fat mobilization (van Kneegsel *et al.*, 2005). Indeed, under severe negative EB, short-chain and medium-chain FA concentrations in milk are reduced, while long-chain FA concentrations are increased (Nogalski *et al.*, 2012). This can be explained by the fact that oleic acid (C18:1-cis9) is the predominant FA in adipocytes, and it is released primarily through lipolysis during NEB (Rukkamsuk *et al.*, 2000). Actually, Gross *et al.* (2011) found a correlation between NEB and the proportion of C18:1-cis9 in milk ($r^2 = 0.77$). Therefore, the proportion of this fatty acid could be used as a biomarker for EB diagnosis in dairy cows.

Glucose

Glucose is another possible biomarker for NEB diagnosis. Glucose is an essential metabolite for the mammary epithelial cells. Mammary epithelial cells do not synthesize glucose because they lack the enzyme glucose-6-phosphatase (Scott *et al.*, 1976). Therefore, glucose concentration in mammary epithelial cells depends on the glucose transferred from blood. Consequently, glucose concentrations in milk reflect its concentration in the mammary epithelial cell cytoplasm (Faulkner *et al.*, 1981; Zhao, 2014). Glucose-6-phosphate (G6P) is a central metabolite in the glycolytic pathway as it is an intermediate compound during lactose synthesis and participates in the first step for glycolysis and the pentose phosphate pathway (PPP; Zhao, 2014). On this basis, G6P has been also proposed as a biomarker for NEB diagnosis. Larsen and Moyes (2015) analysed 3200 milk samples from Holstein and Jersey cows for free glucose and G6P. During the first 21 weeks of lactation, free glucose concentrations increased whereas G6P concentrations in milk decreased. Accordingly, Zachut *et al.* (2016) reported that milk glucose concentrations were positively correlated to days in lactation. In contrast, the average concentration of G6P in the milk was the highest during the first week of lactation, and was negatively correlated to days in

lactation (Zachut *et al.*, 2016). Therefore, milk G6P/glucose ratio was suggested as a biomarker of the oxidative stress in the mammary epithelial cells (Zachut *et al.*, 2016). In addition, that study showed that G6P/glucose ratio was highly correlated to plasma NEFA concentrations ($r^2 = 0.81$, Zachut *et al.* 2016). More research is required to validate the use of free glucose and G6P as biomarkers of EB, which may potentially be used in the future for in-line surveillance systems on-farm.

Glycolytic enzymes

Based on the relation between milk G6P and EB, glucose-6-phosphate-dehydrogenase (G6PDH), the first enzyme in the pentose phosphate pathway, has been also suggested as a milk biomarker for NEB diagnosis in dairy cows. Only a few studies have reported G6PDH activity in milk from cows. Similar to milk G6P concentrations, Zachut *et al.* (2016) showed that milk G6PDH activity in cows was highest on the first and second week of lactation, the decreasing until the fifth week of lactation. Moreover, G6PDH activity was correlated with milk G6P ($r^2 = 0.68$), with G6P/glucose ratio ($r^2 = 0.53$) and negatively correlated to days in lactation ($r^2 = -0.69$), dry matter intake ($r^2 = -0.65$) and EB ($r^2 = -0.52$; Zachut *et al.* 2016). It was observed that milk G6PDH activity was found to be 2-fold higher in cows that were under NEB on the third week of lactation compared to those under positive energy balance (unpublished data). Based on these findings, G6PDH activity could be used as a biomarker in milk for NEB diagnosis in early lactation cows, however, more research is required to validate this biomarker.

Biomarkers of oxidative stress in blood and milk

Oxidative stress is affected during NEB as a consequence of the pro-inflammatory effects of fat mobilization (Sordillo *et al.*, 2009; Bradford *et al.*, 2015). The metabolic demands associated with late pregnancy, parturition and initiation of lactation are

thought to increase the production of reactive oxygen species (ROS) (Esposito *et al.*, 2014). The bulk of the oxidants are ROS, but reactive nitrogen species (RNS) contribute to the pools of oxidants and both are produced during physiological and pathological states in the organism. All macromolecules like lipids, proteins and DNA are targets for oxidative stress (Mavangira *et al.* Sordillo, 2018). There are a lot of useful biomarkers of oxidative stress connected to lipid and protein metabolism, the most known biomarker of protein oxidation being the measurement of carbonyl groups. When ROS attack the amino acid side chains of proline, arginine, lysine and threonine, carbonyl groups are generated. A more specific parameter for protein oxidation is 2,4-dinitrophenylhydrazine (DNPH) which allows the determination of total protein carbonyl groups (Mavangira *et al.* Sordillo, 2018). Hypochlorous acid-induced products are generated during inflammation (Shacter, 2000). The level of di-tyrosine reflects the oxidative damage of proteins and measurement is possible by chemiluminescence (Bordignon *et al.*, 2014). Under transitional stages cows are exposed to different oxidative processes which can compromise antioxidative status: parturition, NEB, ketosis risk, fertilization, early embryo development, concurrent pregnancy and lactation, inflammation in connection with the udder (subclinical mastitis) and uterus (subclinical endometritis), gut health etc. It has been observed that the ratio between the plasma level of advanced oxidation protein products and albumin (AOPP/albumin) is a sensitive indicator of oxidative stress (Celi *et al.*, 2011). Because of the lack of antioxidants in maize silage, feeding cows with maize silage increase AOPP concentration (Celi and Raadsma, 2010). Immune cells are particularly sensitive to oxidative stress due to the high content of polyunsaturated FA present in the cellular membrane, which are susceptible to peroxidation, increasing the production of ROS (Spears and Weiss, 2008; Esposito *et al.*, 2014, Celi and Gabai, 2015). Recently, Alharthi *et al.* (2018) reported a gradual increase in reactive oxygen metabolites (ROM) between -10 and 20 d relative to parturition, and a previous report demonstrated that cows losing more BCS, which is indicative of NEB, had lower superoxide dismutase (SOD) activity and higher ROM in the bloodstream (Bernabucci *et al.*, 2005). Also, a significant correlation between milk AOPP and somatic cells count has been observed (Guzzo *et al.*, 2015). Therefore, it seems that ROM and SOD activity in blood may be used as biomarkers of oxidative stress in dairy cows. The widely used biomarker of lipid peroxidation, malondialdehyde (MDA), a low-molecular-weight product created during the decomposition of polyunsaturated fatty acid (PUFA) may reflect the oxidative stress of the animal. In milk of PP cows, MDA concentration was found to be highest in early lactation and then exponentially decayed, and was inversely correlated with days in lactation (Zachut *et al.*, 2016). In agreement, milk anti-oxidative capacity (ORAC values) tended to be negatively and exponentially correlated with days in lactation ($r^2 = -0.29$) and EB ($r^2 = -0.30$), and to be positively and linearly correlated to milk G6P ($r^2 = 0.25$) (Zachut *et al.*, 2016). This suggests that milk MDA and ORAC can serve as biomarkers of oxidative stress in milk. However, MDA has been shown to be an inconsistent and variable marker (Celi, 2010). A more reliable marker of lipid oxidation may be ELISA based isoprostanes, as increased levels of 15-F2-IsoP were determined during coliform mastitis (Mavangira *et al.*, 2016) and related to inflammation (Mavangira and Sordillo, 2018).

A recent study demonstrated a positive correlation between reactive oxygen and nitrogen species (RONs) and oxidant status index (OSi, which is defined as the ratio between reactive oxygen

and nitrogen species) and total antioxidant potential ($r^2 = 0.75$), as well as a negative correlation between OSi and serum antioxidant potential (AOP; $r^2 = -0.58$). An increase in the ratio indicates a higher risk for oxidative stress due to an increase in pro-oxidant production or defensive antioxidant depletion (Ling *et al.*, 2018).

Environmental heat stress can increase oxidative stress in dairy cows. Bernabucci *et al.* (2002) reported that transition cows exposed to heat stress during summer had higher erythrocyte activity, glutathione peroxidase activity, intracellular thiols, and MDA compared to spring cows, indicating a condition of oxidative stress in the summer transitioning dairy cows. Plasma concentrations of the oxidative stress marker MDA were higher in transition dairy cows calving during summer heat stress compared to those calving in winter (Zachut *et al.*, 2017). In studies conducted in mid-lactation heat-stressed cows, a reduction in plasma antioxidant activity was found (Harmon *et al.*, 1997). Further research is required to establish the utility of oxidative stress as putative biomarkers of heat stress in cattle.

Consequences and applications

Biomarkers of production-related diseases

Dairy cows are one of the most intensively farmed animals worldwide. High-yielding dairy cows have been genetically selected for high milk production, which increases the susceptibility of these animals to develop certain diseases such as mastitis, hypocalcaemia, rumen acidosis, ketosis and laminitis. The establishment of biomarkers for early detection of these diseases is one of the most important aspects of current dairy research.

Mastitis, defined as the inflammation of the udder, is the most prevalent production-related disease in dairy herds worldwide. In dairy ruminants, mastitis is frequently caused by intra-mammary infections. Those infections often impact animal welfare and contribute to economic losses for farmers (Halasa *et al.*, 2007; Hernández-Castellano *et al.*, 2011). Currently, the most sensitive technique available for clinical and subclinical mastitis detection is SCC or somatic cell count (Schukken *et al.*, 2003), while the identification of pathogens requires bacteriological culture (Nyman *et al.*, 2014) or molecular methods, namely PCR. Cow-side or point of care diagnostic tests for bacterial identification are becoming available, but are not yet in widespread use (Jones *et al.*, 2019). In addition to the innate immune response, represented by increasing SCC, immunoglobulins (mainly IgG) are important components of the specific immune response transferred from blood to milk during mastitis (Wall *et al.*, 2016a, 2016b). The increase of IgG in milk appears to be pathogen-dependent and its use combined with SCC has been proposed for the prediction of the pathogen causing mastitis (Hernández-Castellano *et al.*, 2017a). In-line IgG measurements are currently under development for use at farm level (Lemberskiy-Kuzin *et al.*, 2019), but the technology is not yet available to farmers. Therefore, alternative markers such as lactate dehydrogenase (LDH) and differential somatic cell count (DSCC) have been proposed as markers for early mastitis detection and diagnosis (Chagunda *et al.*, 2006; Damm *et al.*, 2017; Wall *et al.*, 2018).

The use of omics technologies in the field of mastitis in dairy cows has provided knowledge about diverse components involved in the course of the disease and how those components may be affected by the mastitis causing pathogen. For instance, Thomas *et al.* (2016a, 2016b) and Mudaliar *et al.* (2016) used peptidomics,

metabolomics and quantitative proteomics to analyse milk from mastitis caused by *Streptococcus uberis*. These authors established several components such as casein derived peptides, peptides of glycosylation dependent cell adhesion molecule and serum amyloid A, antimicrobial peptides and different inflammation related metabolites. However, Kusebauch *et al.* (2018) described that gram-negative bacteria cause faster and more intense changes in the milk proteome compared to gram-positive bacteria. Based on this differential expression in milk, the authors proposed potential biomarkers to distinguish between mastitis caused by gram-negative and gram-positive bacteria. These biomarkers were α -2 macroglobulin, α -1 antitrypsin, haptoglobin, Serum Amyloid A3, cluster of differentiation CD14, calgranulin B, calgranulin C, cathepsin C, vanin-1, galectin-1, galectin-3 and interleukin 8. This field is growing all the time and it is likely that in the future it will extend to other species such as small ruminants (Katsafadou *et al.*, 2015; Hernández-Castellano *et al.*, 2016a, 2016b; Vasileiou *et al.*, 2019) and water buffalo (Patbandha *et al.*, 2015). Different biomarkers may be more applicable in non-bovine species, for instance, cathelicidin has been proposed for use in goats (Tedde *et al.*, 2019). The area has been recently reviewed by Almeida and Eckersall (2018) and Boschetti *et al.* (2019).

Metabolic diseases are usually linked to an increased demand for a specific nutrient that has become deficient under certain conditions. In the case of dairy cows, a special focus should be placed on the onset of lactation, when the sudden high demand of nutrients for milk production increases the metabolic load on the animal (Weaver *et al.*, 2017). Most common metabolic diseases in dairy cows are hypocalcaemia (HC), ketosis (KT) and ruminal acidosis (RA). Hypocalcaemia mainly occurs at the onset of lactation when the fast and high demand for calcium by the mammary gland for milk production decreases circulating calcium concentrations below 1.4 mmol/l (clinical HC) or below 2 mmol/l (subclinical HC) (Hernández-Castellano *et al.*, 2017b). Similar to HC, KT occurs in cattle when energy demands exceed energy intake, resulting in negative energy balance (Zarrin *et al.*, 2013). In order to prevent KT, sufficient energy has to be provided through the feed. However, when these high dietary requirements for milk production are reached by feeding diets with high amounts of rapidly fermentable carbohydrates (i.e. starch) and low fibre content, bacterial populations are altered in the rumen. Consequently, acids (i.e. lactate) and glucose accumulate, decreasing ruminal pH (pH < 4.8) leading to RA (the animal is not able to restore pH levels itself) or subacute ruminal acidosis (SARA, the animal restores normal pH levels within hours). This metabolic disease damages the ruminal and intestinal wall and decreases blood pH, leading to the metabolic acidosis.

The rapid development of new sensor technologies has allowed the creation of tools that provide individual and dynamic information about the animals (Caja *et al.*, 2016). Additionally, innovations in robotics have provided opportunities to develop powerful systems for the individualized feeding of dairy cows according to their specific and singular nutritional and physiological requirements. The constant flow of information can be used to monitor dairy cows and therefore prevent these metabolic diseases in dairy herds, as highlighted below.

It is quite evident that blood calcium is the best indicator for HC diagnosis. However, several additional markers can be used to determine the calcium status in the animal and prevent HC. Urine pH could be used as indicator of the acid-base balance in blood (Thilsing-Hansen *et al.*, 2002). Around parturition, it is convenient to create a physiological state of compensated systemic

acidosis in cows by feeding diets with a negative dietary cation-anion difference. This acidosis will be compensated in part by bone calcium resorption (Lemann *et al.*, 2003). Therefore, cows with urinary pH within 5.5 to 6.2 around parturition are considered to be in temporary acidosis, which is associated with reduced HC (Horst *et al.*, 1997). In the case of KT, BHB concentration in blood >1.4 mmol/l is the common marker used for KT diagnosis. However, other factors such as NEFA, glucose, glucagon or insulin are also affected during KT. Geishauser *et al.* (1998) and Koeck *et al.* (2014) showed how BHB measured in milk correlates with BHB measured in blood. Based on this principle, modern milking robots incorporate BHB measurements in milk, which provides constant information about the energy status of the animal, contributing to the prevention of KT. One of the most obvious markers for ruminal acidosis is the pH value of the ruminal fluid. Wireless pH probes located in ruminal boluses provide ruminal pH measurement in real time, but pH recording can differ depending on the compartment where the probe is placed (Neubauer *et al.*, 2018). Besides ruminal pH, other markers for detection of either RA or SARA can be measured in blood, urine, faeces or milk. In blood, D-lactate has been proposed as a marker for SARA (Larsen, 2017) as it is exclusively synthesized in the rumen by lactobacilli and bifidobacteria (Ewaschuk *et al.*, 2005) and it is poorly metabolized by mammals. Consequently, D-lactate accumulates in body fluids such as milk and it could be used as a biomarker for RA diagnosis (Ewaschuk *et al.*, 2005). In addition to D-lactate, Danscher *et al.* (2015) also described that RA or SARA do not affect milk protein content, but do reduce fat content in milk compared to control cows (4.14 and 5.08%, respectively). Therefore, animals with fat:protein ratio below 1 in milk are susceptible to suffer RA or SARA (Danscher *et al.*, 2015; Vlček *et al.*, 2016; Rojo-Gimeno *et al.*, 2018).

Biomarkers of reproductive state

An efficient management of fertility requires a tight collaboration between farmers and veterinarians, a consistent analysis of the farm records and accurate clinical data. Moreover, reduced fertility can be considered an indicator of poor health and welfare (Walsh *et al.*, 2011, Gabai *et al.* 2018). Therefore, the development and continuous validation of specific biomarkers for fertility is relevant in dairy research.

To decide a suitable reproductive management strategy, several factors need to be monitored. The importance of oestrus detection on the reproductive efficiency is widely recognized, but the assessment of the resumption of ovarian activity and uterine health during the puerperium also needs to be considered. Pregnancy diagnosis should be performed as soon as possible after artificial insemination (AI), and conception failure should be discriminated from embryonic loss.

Progesterone indicates the presence of an active corpus luteum. Therefore, it has been used as a biomarker of reproduction efficiency for decades (Veronesi *et al.*, 2002). As progesterone is transferred from blood to milk, milk progesterone is a suitable non-invasive biomarker in dairy animals for reproductive status (Xu *et al.*, 2005, 2013; Kappel *et al.*, 2007; Posthuma-Trumpie *et al.*, 2009; Oku *et al.*, 2011), although measurement of progesterone is usually too expensive to be extensively applied over periods of several weeks in a large number of animals. Moreover, manual sampling is not practical in commercial farms, where large herds need to be monitored, and the use of automated sampling systems is needed.

As described above, oestrus detection is essential to keep high reproductive efficiency in dairy herds. Considerable progress has been achieved to automatically detect oestrus on farm level. Most systems are activity-based and monitor the behavioural signs of mating, using detectors for standing heat and/or activity-meters (Saint-Dizier and Chastant-Maillard, 2012). These systems display high degrees of sensitivity and specificity if tested in experimental settings, but their efficiency can be affected by environmental conditions (e.g.: housing and flooring conditions) and animal health (e.g. lameness) (Saint-Dizier and Chastant-Maillard, 2012). The use of milk progesterone in combination with activity-based systems has led to increased oestrus detection efficiency. A fully automated system for milk progesterone measurement (Herd Navigator[®], Lattec, DK) has become available in Europe and Canada, which can be combined with DeLaval[®] milking robot or parlour (Mazeris, 2010) and allows the analysis and interpretation of frequently taken samples (Friggens and Chagunda, 2005; Friggens *et al.*, 2008).

The analysis of both plasma and milk progesterone profiles in combination with clinical findings is used for diagnosing atypical ovarian patterns and identifying potentially sub fertile cows (Lamming and Darwash, 1998). Delayed postpartum resumption of the ovarian activity and prolonged luteal phases commonly cause reduced fertility (Lamming and Darwash, 1998; Gautam *et al.*, 2010; Ranasinghe *et al.*, 2011) and reduced embryo survival (Santos *et al.*, 2009). Adequate endocrine regulation during the follicular phase is highly relevant for a good fertilization (Starbuck *et al.*, 2006). Accurate monitoring of both pre-ovulatory decline and post-insemination rise in milk progesterone can be used to identify animals with compromised fertility, as progesterone secretion is directly responsible for embryonic development since the very early stage of pregnancy (Green *et al.*, 2005; Stronge *et al.*, 2005; McNeill *et al.*, 2006). For instance, low milk progesterone concentrations around days 4–7 after insemination are associated with low fertility and increased risk of embryonic losses (McNeill *et al.*, 2006).

Progesterone concentrations can also be altered by hepatic metabolism (Rhinehart *et al.*, 2009). In dairy cows, the cytochrome CYP2C (converting progesterone to 21-hydroxyprogesterone) and the aldo-keto reductase AKR1C (converting progesterone to 20 α -hydroxyprogesterone) are the most active enzymes in the liver (Lemley and Wilson, 2010). Future studies should test progesterone metabolites as potential biomarkers of fertility. It is worth noting that food intake plays an important role in regulating progesterone metabolism by altering liver blood flow and hepatic enzymes (Sangsritavong *et al.*, 2002; Lemley *et al.*, 2011; Hart *et al.*, 2014).

Progesterone profiles can be mathematically modelled (Friggens and Chagunda, 2005; Blavy *et al.*, 2016) to define 'typical' and 'atypical' progesterone profiles. The retrospective analysis of in-line progesterone records matched by accurate clinical information offers a unique possibility of developing biological models useful for management purposes. Some of the potential applications are the identification of abnormal oestrous cycle responsible for poor fertility (Bruinjé *et al.*, 2017a) or milk progesterone profiles that may be helpful in predicting the AI outcome (Bruinjé *et al.*, 2017b). In the future, information obtained with activity-based devices and progesterone profiles may be combined with novel indicators measured in milk, which show slight but significant variation related to the reproductive cycle (Toledo-Alvarado *et al.*, 2018).

As described above, progesterone concentrations cannot be considered as a sensitive biomarker for performing pregnancy

diagnosis or detecting embryonic losses, in particular when embryonic death occurs before CL regression (Szenci *et al.*, 2000). The measurement of pregnancy specific metabolites is a synergistic tool for diagnosing pregnancy and embryonic death. Pregnancy-associated glycoproteins (PAGs) constitute a large family of glycoproteins specifically expressed in the trophoctoderm of the placenta in ungulate species. Pregnancy-associated glycoproteins can be found in the maternal blood from approximately 3 weeks of pregnancy (Wallace *et al.*, 2015). Plasma PAG-1 concentrations in cows seem to be a good biomarker for pregnancy diagnosis and embryonic loss from day 28 after AI, if the time interval between calving and insemination is of at least 60 d (Haugejorden *et al.*, 2006; Friederick and Holtz, 2010; Celi *et al.*, 2011; Barbato *et al.*, 2013). Milk PAG concentrations are 20–30 times lower than in plasma (Friederick and Holtz, 2010). Therefore, most of the available assays are not suitable for measuring PAG in milk before day 60 of pregnancy (Friederick and Holtz, 2010; LeBlanc, 2013; Lawson *et al.*, 2014). However, an immunoradiometric assay that is able to measure milk PAG (picogram levels) has been developed by Melo Sousa *et al.* (2015) and it may be used in low fertility herds for pregnancy diagnosis.

Reduced uterine health during the first 45 d postpartum decreases fertility in dairy cows. In addition, risk for suffering clinical or subclinical endometritis is increased under these conditions (Kasimanickam *et al.*, 2004; Sheldon *et al.*, 2009; Walsh *et al.*, 2011). Reported prevalence of endometritis in dairy cows is very variable (5–68%). Some of the factors that affect this prevalence variability are the timing of examination after calving and the diagnostic methods (de Boer *et al.*, 2014). In order to reduce such variability, the development of an *in vivo* cow test for uterine inflammation (specific electronic noses, for instance) would be useful. Electronic noses consist of an array of electronic sensors for chemical detection of volatile molecules. In addition, electronic noses are cheaper, faster, portable, and easier to manipulate than gas chromatography techniques. However, low specificity is one of the most limiting factors for electronic noses (Kou *et al.*, 2017). Although imperfect, electronic noses are a reasonable tool to improve odour assessment of vaginal discharges (Sannmann *et al.*, 2013). This system displays higher intra-assay repeatability compared to the human nose, although this device is not able to fully discriminate between pathogens causing endometritis (Burfeind *et al.*, 2014).

Conclusions and future prospects

This paper summarizes the current knowledge of biomarkers for some important aspects affecting animal production and welfare in dairy herds and sets the focus of the future research that needs to be done to improve performance and welfare in dairy animals.

In the future, the emergence of new technologies (omics approaches and systems biology) will probably contribute to identify biomarkers for specific health and welfare problems at a much earlier stage. Faster analytical procedures with enhanced analytical sensitivity are also required. For instance, for mastitis detection a huge simplification of the milk proteome complexity can be achieved by exploiting the selectivity derived by the peculiar surface topography of surface active maghemite nanoparticles, which allow the rapid determination of hidden putative biomarkers by a cutting edge diagnostic strategy (Magro *et al.*, 2018).

Some biomarkers, although recognized as very specific, still need to be integrated in automated systems, platforms and

technologies so they can be used by farmers and veterinarians. For instance, although PAGs can be considered specific biomarkers for pregnancy diagnosis and foetal welfare, such biomarkers have not yet been implemented at the farm level. Biomarkers can be mathematically modelled to create biological models that contribute to management decisions. Therefore, in the near future artificial intelligence technology could take advantage of retrospective examination of the available databases. For instance, progesterone and BHB concentrations can be measured in milk and, therefore, can be used to obtain large datasets.

Effective strategies for improving performance, health and welfare in dairy animals will require collaboration across a broad range of specialisms and must embrace farmers, consultants, veterinarians, and bio-informaticians. Many methods are available under the term precision livestock farming, which is defined as real-time monitoring technologies aimed at managing the smallest manageable production unit. This implies a novel machine-based approach about the most significant diseases in intensive dairy farming (lameness, mastitis, ketosis; Halachmi and Guarino, 2016) as well as quantifying pain and stress, NEB, heart rate, odour etc. (Halachmi *et al.* 2019). Animal monitoring will inform farmers about disturbances at early stages of specific diseases, improving animal performance, health and welfare.

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References

- Alharthi A, Zhou Z, Lopreiato V, Trevisi E and Loor JJ (2018) Body condition score prior to parturition is associated with plasma and adipose tissue biomarkers of lipid metabolism and inflammation in Holstein cows. *Journal of Animal Science and Biotechnology* **9**, 12.
- Almeida AM and Eckersall PD (2018) Proteomics and mammary gland research in dairy species. In de Almeida AM, Eckersall D and Miller I (eds), *Proteomics in Domestic Animals: From Farm to Systems Biology*. Cham, Switzerland: Springer International Publishing, pp. 255–280.
- Almeida AM, Zachut M, Hernández-Castellano LE, Šperanda M, Gabai G and Mobasher A (2019) Biomarkers of fitness and welfare in dairy animals: healthy living. *Journal of Dairy Research* **86**, 379–387.
- Barbato O, Merlo M, Celi P, Sousa NM, Guarneri L, Beckers JF and Gabai G (2013) Relationship between plasma progesterone and pregnancy-associated glycoprotein concentrations during early pregnancy in dairy cows. *The Veterinary Journal* **195**, 385–387.
- Baumgard LH, Collier RJ and Bauman DE (2017) A 100-year review: regulation of nutrient partitioning to support lactation. *Journal of Dairy Science* **100**, 10353–10366.
- Bell AW and Bauman DE (1997) Adaptations of glucose metabolism during pregnancy and lactation. *Journal of Mammary Gland Biology and Neoplasia* **2**, 265–278.
- Bernabucci U, Ronchi B, Lacetera N and Nardone A (2002) Markers of oxidative status in plasma and erythrocytes of transition dairy cows during hot season. *Journal of Dairy Science* **85**, 2173–2179.
- Bernabucci U, Ronchi B, Lacetera N and Nardone A (2005) Influence of body condition score on relationships between metabolic status and oxidative stress in periparturient dairy cows. *Journal of Dairy Science* **88**, 2017–2026.
- Blavy P, Derks M, Martin O, Hoglund JK and Friggens NC (2016) Overview of progesterone profiles in dairy cows. *Theriogenology* **86**, 1061–1071.
- Boor KJ, Wiedmann M, Murphy S and Alcaine S (2017) A 100-year review: microbiology and safety of milk handling. *Journal of Dairy Science* **100**, 9933–9951.
- Bordignon M, Da Dalt L, Marinelli L and Gabai G (2014) Advanced oxidation protein products are generated by bovine neutrophils and inhibit free radical production in vitro. *Veterinary Journal* **199**, 162–168.
- Boschetti E, Hernández-Castellano LE and Righetti PG (2019) Progress in farm animal proteomics: the contribution of combinatorial peptide ligand libraries. *Journal of Proteomics* **197**, 1–13.
- Bradford BJ, Yuan K, Farney JK, Mamedova LK and Carpenter AJ (2015) Invited review: inflammation during the transition to lactation: new adventures with an old flame. *Journal of Dairy Science* **98**, 6631–6650.
- Bruinjé TC, Colazo MG, Gobikrushnth M and Ambrose DJ (2017a) Relationships among early postpartum luteal activity, parity, and insemination outcomes based on in-line milk progesterone profiles in Canadian Holstein cows. *Theriogenology* **100**, 32–41.
- Bruinjé TC, Gobikrushnth M, Colazo MG and Ambrose DJ (2017b) Dynamics of pre- and post-insemination progesterone profiles and insemination outcomes determined by an in-line milk analysis system in primiparous and multiparous Canadian Holstein cows. *Theriogenology* **102**, 147–153.
- Burfeind O, Bruins M, Bos A, Sannmann I, Voigtsberger R and Heuwieser W (2014) Diagnosis of acute puerperal metritis by electronic nose device analysis of vaginal discharge in dairy cows. *Theriogenology* **82**, 64–70.
- Caja G, Castro-Costa A and Knight CH (2016) Engineering to support well-being of dairy animals. *Journal of Dairy Research* **83**, 136–147.
- Celi P (2010) The role of oxidative stress in small ruminants' health and production. *Brazilian Journal of Animal Science* **39**, 348–363.
- Celi P and Gabai G (2015) Oxidant/antioxidant balance in animal nutrition and health: the role of protein oxidation. *Frontiers in Veterinary Science* **2**, 1–13.
- Celi P, Merlo M, Da Dalt L, Stefani A, Barbato O and Gabai G (2011) Relationship between late embryonic mortality and the increase in plasma advanced oxidised protein products (AOPP) in dairy cows. Reproduction, *Fertility and Development* **23**, 527–533.
- Celli P and Raadsma HW (2010) Effects of Yerba Mate (*Ilex paraguariensis*) supplementation on the productive performance of dairy cows during mid-lactation. *Animal Production Science* **50**, 339–344.
- Chagunda MG, Friggens NC, Rasmussen MD and Larsen T (2006) A model for detection of individual cow mastitis based on an indicator measured in milk. *Journal of Dairy Science* **89**, 2980–2998.
- Chapinal N, LeBlanc SJ, Carson ME, Leslie KE, Godden S, Capel M, Santos JEP, Overton MV and Duffield TF (2012) Herd-level association of serum metabolites in the transition period with disease, milk production, and early lactation reproductive performance. *Journal of Dairy Science* **95**, 5676–5682.
- Damm M, Holm C, Blaabjerg M, Bro MN and Schwarz D (2017) Differential somatic cell count – a novel method for routine mastitis screening in the frame of Dairy Herd Improvement testing programs. *Journal of Dairy Science* **100**, 4926–4940.
- Danscher AM, Li SC, Andersen PH, Khafipour E, Kristensen NB and Plaizier JC (2015) Indicators of induced subacute ruminal acidosis (SARA) in Danish Holstein cows. *Acta Veterinaria Scandinavica* **57**, 39.
- de Boer MW, LeBlanc SJ, Dubuc J, Meier S, Heuwieser W, Arlt S, Gilber RO and McDougall S (2014) Systematic review of diagnostic tests for reproductive-tract infection and inflammation in dairy cows. *Journal of Dairy Science* **97**, 3983–3999.
- Douphrate DI, Hagevoort GR, Nonnenmann MW, Lunner Kolstrup C, Reynolds SJ, Jakob M and Kinsel M (2013) The dairy industry: a brief description of production practices, trends, and farm characteristics around the world. *Journal of Agromedicine* **18**, 187–197.
- Drackley JK (1999) ADSA Foundation Scholar Award. Biology of dairy cows during the transition period: the final frontier? *Journal of Dairy Science* **82**, 2259–2273.
- Duffield TF, Lissemore KD, McBride BW and Leslie KE (2009) Impact of hyperketonemia in early lactation dairy cows on health and production. *Journal of Dairy Science* **92**, 571–580.
- Duncan AJ, Teufel N, Mekonnen K, Singh VK, Bitew A and Gebremedhin B (2013) Dairy intensification in developing countries: effects of market quality on farm-level feeding and breeding practices. *Animal: An International Journal of Animal Bioscience* **7**, 2054–2062.
- Duplessis M, Santschi DE, Plante S, Bergeron C, Lefebvre DM, Durocher J and Cue RI (2019) Milk β -hydroxybutyrate concentration measured by

- Fourier-transform infrared and flow-injection analyses from samples taken at different times relative to milking. *Journal of Dairy Research* **86**, 208–210.
- Espósito G, Irons PC, Webb EC and Chapwanya A** (2014) Interactions between negative energy balance, metabolic diseases, uterine health and immune response in transition dairy cows. *Animal Reproduction Science* **144**, 60–71.
- Essl A** (1998) Longevity in dairy cattle breeding: a review. *Livestock Production Science* **57**, 79–89.
- Ewaschuk JB, Naylor JM and Zello GA** (2005) D-lactate in human and ruminant metabolism. *Journal of Nutrition* **135**, 1619–1625.
- Faulkner A, Chaiyabutr N, Peaker M, Carrick DT and Kuhn NJ** (1981) Metabolic significance of milk glucose. *Journal of Dairy Research* **48**, 51–56
- Friederick M and Holtz W** (2010) Establishment of an ELISA for measuring bovine pregnancy-associated glycoprotein in serum or milk and its application for early pregnancy detection. *Reproduction in Domestic Animals* **45**, 142–146.
- Friggens NC and Chagunda MGG** (2005) Prediction of the reproductive status of cattle on the basis of milk progesterone measures: model description. *Theriogenology* **64**, 155–190.
- Friggens NC, Bjerring M, Ridder C, Højsgaard S and Larsen T** (2008) Improved detection of reproductive status in dairy cows using milk progesterone measurements. *Reproduction in Domestic Animals* **43**(suppl. 2), 113–121.
- Gabai G, Amadori M, Knight CH and Werling D** (2018) The immune system is part of a whole-organism regulatory network. *Research in Veterinary Science* **116**, 1–3.
- Gautam G, Nakao T, Yamada K and Yoshida C** (2010) Defining resumption of ovarian activity postpartum and its impact on subsequent reproductive performances in Holstein cows. *Theriogenology* **73**, 180–189.
- Geishauer T, Leslie K, Kelton D and Duffield T** (1998) Evaluation of five cowside tests for use with milk to detect subclinical ketosis in dairy cows. *Journal of Dairy Science* **81**, 438–443.
- Green MP, Hunter MG and Mann GE** (2005) Relationships between maternal hormone secretion and embryo development on day 5 of pregnancy in dairy cows. *Animal Reproduction Science* **88**, 179–189.
- Gross J, van Dorland HA, Bruckmaier RM and Schwarz FJ** (2011) Milk fatty acid profile related to energy balance in dairy cows. *Journal of Dairy Research* **78**, 479–488.
- Guzzo N, Balloni L, Mantovani R, Da Dalt I and Gabai G** (2015) Oxidized protein biomarkers in the blood and milk of cows supplemented with flaxseed during the dry period. In Knight CH (ed.), *2nd DairyCare Conference March 3–4*. Cordoba Spain-DairyCare COST Action FA1308, p. 60.
- Halachmi I and Guarino M** (2016) Editorial: precision livestock farming: a ‘per animal’ approach using advanced monitoring technologies. *Animal: An International Journal of Animal Bioscience* **10**, 1482–1483.
- Halachmi I, Guarino M, Bewley J and Pastell M** (2019) Smart animal agriculture: application of real-time sensors to improve animal well-being and production. *Annual Review of Animal Biosciences* **7**, 403–425.
- Halasa T, Huijps K, Osteras O and Hogeveen H** (2007) Economic effects of bovine mastitis and mastitis management: a review. *Veterinary Quarterly* **29**, 18–31.
- Harmon RJ, Lu M, Trammel DS and Smith BA** (1997) Influence of heat stress and calving on antioxidant activity in bovine blood. *Journal of Dairy Science* **80**(suppl. 1), 264
- Hart CG, Camacho LE, Swanson KC, Vonnahme KA and Lemley CO** (2014) Hepatic steroid metabolizing enzyme activity during early, mid, and late bovine pregnancy. *Domestic Animal Endocrinology* **49**, 31–38.
- Haugejorden G, Waage S, Dahl E, Karlberg K, Beckers JF and Ropstad E** (2006) Pregnancy associated glycoproteins (PAG) in postpartum cows, ewes, goats and their offspring. *Theriogenology* **66**, 1976–1984.
- Hernández-Castellano LE, Torres A, Alavoine A, Ruiz-Diaz MD, Arguello A, Capote J and Castro N** (2011) Effect of milking frequency on milk immunoglobulin concentration (IgG, IgM and IgA) and chitotriosidase activity in Majorera goats. *Small Ruminant Research* **98**, 70–72.
- Hernández-Castellano LE, Almeida AM, Renaut J, Arguello A and Castro N** (2016a) A proteomics study of colostrum and milk from the two major small ruminant dairy breeds from the Canary Islands: a bovine milk comparison perspective. *Journal of Dairy Research* **83**, 366–374.
- Hernández-Castellano LE, Ferreira AM, Nanni P, Grossmann J, Argüello A, Capote J, Cai G, Lippolis J, Castro N and de Almeida AM** (2016b) The goat (*Capra hircus*) mammary gland secretory tissue proteome as influenced by weight loss: a study using label free proteomics. *Journal of Proteomics* **145**, 60–69.
- Hernández-Castellano LE, Wall SK, Stephan R, Corti S and Bruckmaier R** (2017a) Milk somatic cell count, lactate dehydrogenase activity, and immunoglobulin G concentration associated with mastitis caused by different pathogens: a field study. *Schweizer Archiv für Tierheilkunde* **159**, 283–290.
- Hernández-Castellano LE, Hernández LL, Weaver S and Bruckmaier RM** (2017b) Increased serum serotonin improves parturient calcium homeostasis in dairy cows. *Journal of Dairy Science* **100**, 1580–1587.
- Hernández-Castellano LE, Nally JE, Lindahl J, Wanapat M, Alhidary IA, Fanguero D, Grace D, Ratto M, Bambou JC and de Almeida AM** (2019) Dairy science and health in the tropics: challenges and opportunities for the next decades. *Tropical Animal Health and Production* **51**, 1009–1017.
- Horst RL, Goff JP, Reinhardt TA and Buxton DR** (1997) Strategies for preventing milk fever in dairy cattle. *Journal of Dairy Science* **80**, 1269–1280.
- Imhasly S, Bieli C, Naegeli H, Nyström L, Ruetten M and Gerspach C** (2015) Blood plasma lipidome profile of dairy cows during the transition period. *BMC Veterinary Research* **11**, 252
- Jones G, Bork O, Ferguson SA and Bates A** (2019) Comparison of an on-farm point-of-care diagnostic with conventional culture in analysing bovine mastitis samples. *Journal of Dairy Research* **86**, 222–225.
- Kappel ND, Proll F and Gauglitz G** (2007) Development of a TIRF-based biosensor for sensitive detection of progesterone in bovine milk. *Biosensors and Bioelectronics* **22**, 2295–2300.
- Kasimanickam R, Duffield TF, Foster RA, Gartley CJ, Leslie KE, Walton JS and Johnson WH** (2004) Endometrial cytology and ultrasonography for the detection of subclinical endometritis in postpartum dairy cows. *Theriogenology* **62**, 9–23.
- Katsafadou AI, Tsangaris GT, Billinis C and Fthenakis GC** (2015) Use of proteomics in the study of microbial diseases of small ruminants. *Veterinary Microbiology* **181**, 27–33.
- Koeck A, Jamrozik J, Schenkel FS, Moore RK, Lefebvre DM, Kelton DF and Miglior F** (2014) Genetic analysis of milk beta-hydroxybutyrate and its association with fat-to-protein ratio, body condition score, clinical ketosis, and displaced abomasum in early first lactation of Canadian Holsteins. *Journal of Dairy Science* **97**, 7286–7292.
- Koolhaas JM and van Reenen CG** (2016) Interaction between coping style/personality, stress, and welfare: relevance for domestic farm animals. *Journal of Animal Science* **94**, 2284–2296.
- Kusebauch U, Hernández-Castellano LE, Bislev SL, Moritz RL, Rontved CM and Bendixen E** (2018) Selected reaction monitoring mass spectrometry of mastitis milk reveals pathogen-specific regulation of bovine host response proteins. *Journal of Dairy Science* **101**, 6532–6541.
- Kou L, Zhang D and Liu D** (2017) A novel medical e-nose signal analysis system. *Sensors* **17**, 402.
- Lamming GE and Darwash AO** (1998) The use of milk progesterone profiles to characterise components of subfertility in milked dairy cows. *Animal Reproduction Science* **52**, 175–190.
- Larsen T** (2017) Fluorometric determination of D-lactate in biological fluids. *Analytical Biochemistry* **539**, 152–157.
- Larsen T and Moyes KM** (2015) Are free glucose and glucose-6-phosphate in milk indicators of specific physiological states in the cow? *Animal: An International Journal of Animal Bioscience* **9**, 86–93.
- Lawson BC, Shahzad AH, Dolecheck KA, Martel EL, Velek KA, Ray DL, Lawrence JC and Silva WJ** (2014) A pregnancy detection assay using milk samples: evaluations and considerations. *Journal of Dairy Science* **97**, 6316–6325.
- LeBlanc S** (2013) Field evaluation of a pregnancy confirmation test using milk samples in dairy cows. *Journal of Dairy Science* **96**, 2345–2348.
- Lemann J Jr, Bushinsky DA and Hamm LL** (2003) Bone buffering of acid and base in humans. *American Journal of Physiology-Renal Physiology* **285**, F811–F832
- Lemberskiy-Kuzin L, Lavie S, Katz G, Merin U and Leitner G** (2019) Determination of immunoglobulins levels in colostrum by using an on-line milk analyzer. *Canadian Journal of Animal Science* **99**, 631–633.

- Lemley CO and Wilson ME (2010) Effect of cytochrome P450 and aldo-keto reductase inhibitors on progesterone inactivation in primary bovine hepatic cell cultures. *Journal of Dairy Science* **93**, 4613–4624.
- Lemley CO, Vonnahme KA, Tager LR, Krause KM and Wilson ME (2011) Diet-induced alterations in hepatic progesterone (P4) catabolic enzyme activity and P4 clearance rate in lactating dairy cows. *Journal of Endocrinology* **205**, 233–241.
- Ling T, Hernández-Jover M, Sordillo LM and Abuelo A (2018) Maternal late-gestation metabolic stress is associated with changes in immune and metabolic responses of dairy calves. *Journal of Dairy Science* **101**, 6568–6580.
- Magro M, Zaccarin M, Miotto G, Da Dalt L, Baratella D, Fariselli P, Gabai G and Vianello F (2018) Analysis of hard protein corona composition on selective ironoxide nanoparticles by MALDI-TOF mass spectrometry: identification and amplification of a hidden mastitis biomarker in milk proteome. *Analytical and Bioanalytical Chemistry* **410**, 2949–2959.
- Martin NP, Russelle MP, Powell JM, Sniffen CJ, Smith SI, Tricarico JM and Grant RJ (2017) Invited review: sustainable forage and grain crop production for the US dairy industry. *Journal of Dairy Science* **100**, 9479–9494.
- Mavangira V and Sordillo LM (2018) Role of lipid mediators in the regulation of oxidative stress and inflammatory responses in dairy cattle. *Research in Veterinary Science* **116**, 4–14.
- Mavangira V, Mangual MJ, Gandy JC and Sordillo LM (2016) 15-F_{2t}-isoprostane concentrations and oxidant status in lactating dairy cattle with acute coliform mastitis. *Journal of Veterinary Internal Medicine* **30**, 339–347.
- Mazeris F (2010) DeLaval Herd Navigator® proactive herd management. *The First North American Conference on Precision Dairy Management, March 2010*, Toronto, Can. Available at <http://www.precisiondairy.com/proceedings/slmazeris.pdf>.
- McArt JAA, Nydam DV, Oetzel GR, Overton TR and Ospina PA (2013) Elevated non-esterified fatty acids and β-hydroxybutyrate and their association with transition dairy cow performance. *The Veterinary Journal* **198**, 560–570.
- McGuffey RK (2017) A 100-year review: metabolic modifiers in dairy cattle nutrition. *Journal of Dairy Science* **100**, 10113–10142.
- McNeill RE, Diskin MG, Sreenan JM and Morris DG (2006) Associations between milk progesterone concentration on different days and with embryo survival during the early luteal phase in dairy cows. *Theriogenology* **65**, 1435–1441.
- Medhammar E, Wijesinha-Bettoni R, Stadlmayr B, Nilsson E, Charrondiere UR and Burlingame B (2012) Composition of milk from minor dairy animals and buffalo breeds: a biodiversity perspective. *Journal of the Science of Food and Agriculture* **92**, 445–474.
- Sousa N M, Tchimbou AF and Beckers JF (2015) Development of a new immunoradiometric assay for pregnancy-associated glycoproteins (IRMA-PAG) allowing pregnancy follow-up in cattle by using milk samples. Final OptiMIR Scientific and Expert Meeting: From milk analysis to advisory tools (Palais des Congrès, Namur, Belgium, 16–17 April 2015). *Biotechnology, Agronomy, Society and Environment* **19**, 120.
- Möller R, Dannenberger D, Nürnberg G, Strucken E-M and Brockmann GA (2019) Relationship between the fatty acid profile of hair and energy availability of lactating primiparous cows. *Journal of Dairy Research* **86**, 77–84.
- Moyes KM, Larsen T, Friggens NC, Drackley JR and Ingvarstsen KL (2009) Identification of potential markers in blood for the development of subclinical and clinical mastitis in dairy cattle at parturition and during early lactation. *Journal of Dairy Science* **92**, 5419–5428.
- Mudaliar M, Tassi R, Thomas FC, McNeilly TN, Weidt SK, McLaughlin M, Wilson D, Burchmore R, Herzyk P, Eckersall PD and Zadoks RN (2016) Mastitomics, the integrated omics of bovine milk in an experimental model of *Streptococcus uberis* mastitis: 2. Label-free relative quantitative proteomics. *Molecular Biosystems* **12**, 2748–2761.
- Neubauer V, Humer E, Kroger I, Braid T, Wagner M and Zebeli Q (2018) Differences between pH of indwelling sensors and the pH of fluid and solid phase in the rumen of dairy cows fed varying concentrate levels. *Journal of Animal Physiology and Animal Nutrition* **102**, 343–349.
- Nogalski Z, Wroski M, Sobczuk-Szul M, Mochol M and Pogorzelska P (2012) The effect of body energy reserve mobilization on the fatty acid profile of milk in high-yielding cows. *Asian-Australasian Journal of Animal Science* **25**, 1712–1720.
- Nyman AK, Persson Waller K, Bennedsgaard TW, Larsen T and Emanuelson U (2014) Associations of udder-health indicators with cow factors and with intramammary infection in dairy cows. *Journal of Dairy Science* **97**, 5459–5473.
- Oku Y, Osawa T, Hirata T-I, Kon N, Akasaka S, Senosy WS, Takahashi T and Izaiki Y (2011) Validation of a direct time-resolved fluoroimmunoassay for progesterone in milk from dairy and beef cows. *The Veterinary Journal* **190**, 244–248.
- Ospina PA, Nydam DV, Stokol T and Overton TR (2010) Association between the proportion of sampled transition cows with increased nonesterified fatty acids and β-hydroxybutyrate and disease incidence, pregnancy rate, and milk production at the herd level. *Journal of Dairy Science* **93**, 3595–3601.
- Ospina PA, McArt JA, Overton TR, Stokol T and Nydam DV (2013) Using nonesterified fatty acids and β-hydroxybutyrate concentrations during the transition period for herd-level monitoring of increased risk of disease and decreased reproductive and milking performance. *Veterinary Clinics of North America: Food Animal Practice* **29**, 387–412.
- Patbandha TK, Ravikala K, Maharana BR, Marandi S, Ahlawat AR and Gajbhiya PU (2015) Effect of season and stage of lactation on milk components of Jaffrabadi Buffaloes. *The Bioscan* **10**, 635–638.
- Polsky LM and von Keyserlingk AG (2017) Invited review: effects of heat stress on dairy cattle welfare. *Journal of Dairy Science* **100**, 8645–8657.
- Posthuma-Trumple GA, van Amerongen A, Korf J and van Berkel WJH (2009) Perspectives for on-site monitoring of progesterone. *Trends in Biotechnology* **27**, 652–660.
- Pulina G, Milán MJ, Lavín MP, Theodoridis A, Morin E, Capote J, Thomas DL, Francesconi AHD and Caja G (2018) Current production trends, farm structures, and economics of the dairy sheep and goat sectors. *Journal of Dairy Science* **101**, 6715–6729.
- Raboisson D, Barbier M and Maigné E (2016) How metabolic diseases impact the use of antimicrobials: a formal demonstration in the field of veterinary medicine. *PLoS One* **11**, e0164200.
- Ranasinghe RMSBK, Nakao T, Yamada K, Koike K, Hayashi A and Dematawewa CMB (2011) Characteristics of prolonged luteal phase identified by milk progesterone concentrations and its effects on reproductive performances in Holstein cows. *Journal of Dairy Science* **94**, 116–127.
- Rhinehart JD, Starbuck-Clemmer MJ, Flores JA, Milvae RA, Yao J, Poole DH and Inskoop EK (2009) Low peripheral progesterone and late embryonic/early fetal loss in suckled beef and lactating dairy cows. *Theriogenology* **71**, 480–490.
- Roche JR, Berry DP, Bryant AM, Burke CR, Butler ST, Dillon PG, Donaghy DJ, Horan B, Macdonald KA and Macmillan KL (2017) A 100-year review: a century of change in temperate grazing dairy systems. *Journal of Dairy Science* **100**, 10189–10233.
- Rojo-Gimeno C, Fievez V and Wauters E (2018) The economic value of information provided by milk biomarkers under different scenarios: case-study of an ex-ante analysis of fat-to-protein ratio and fatty acid profile to detect subacute ruminal acidosis in dairy cows. *Livestock Science* **211**, 30–41.
- Rukkhwamsuk T, Geelen MJ, Kruip TA and Wensing T (2000) Interrelation of fatty acid composition in adipose tissue, serum, and liver of dairy cows during the development of fatty liver postpartum. *Journal of Dairy Science* **83**, 52–59.
- Saint-Dizier M and Chastant-Maillard S (2012) Towards an automated detection of oestrus in dairy cattle. *Reproduction in Domestic Animals* **47**, 1056–1061.
- Sangsrivong S, Combs DK, Sartori R, Armentano LE and Wiltbank MC (2002) High feed intake increases liver blood flow and metabolism of progesterone and estradiol-17beta in dairy cattle. *Journal of Dairy Science* **85**, 2831–2842.
- Sannmann I, Burfeind O, Suthar V, Bos A, Bruins M and Heuwieser W (2013) Technical note: evaluation of odor from vaginal discharge of cows in the first 10 days after calving by olfactory cognition and an electronic device. *Journal of Dairy Science* **96**, 5773–5779.

- Santos JEP, Rutigliano HM and Sa Filho MF (2009) Risk factors for resumption of postpartum estrous cycles and embryonic survival in lactating dairy cows. *Animal Reproduction Science* **110**, 207–221.
- Schukken YH, Wilson DJ, Welcome F, Garrison-Tikofsky L and Gonzalez RN (2003) Monitoring udder health and milk quality using somatic cell counts. *Veterinary Research* **34**, 579–596.
- Scott RA, Beuman DE and Clark JH (1976) Cellular gluconeogenesis by lactating bovine mammary tissue. *Journal of Dairy Science* **59**, 50–56.
- Shacter E (2000) Quantification and significance of protein oxidation in biological samples. *Drug Metabolism Reviews* **32**, 307–326.
- Sheldon IM, Price SB, Cronin J, Gilbert RO and Gadsby JE (2009) Mechanisms of infertility associated with clinical and subclinical endometritis in high producing dairy cattle. *Reproduction in Domestic Animals* **44** (suppl. 3), 1–9
- Sordillo LM, Contreras GA and Aitken SL (2009) Metabolic factors affecting the inflammatory response of periparturient dairy cows. *Animal Health Research Reviews* **10**, 53–63.
- Spears JW and Weiss WP (2008) Role of antioxidants and trace elements in health and immunity of transition dairy cows. *The Veterinary Journal* **176**, 70–76.
- Starbuck GR, Gutierrez CG, Peters AR and Mann GE (2006) Timing of follicular phase events and the postovulatory progesterone rise following synchronization of oestrus in cows. *The Veterinary Journal* **172**, 103–108.
- Stoop WM, Bovenhuis H, Heck JML and van Arendonk JAM (2009) Effect of lactation stage and energy status on milk fat composition of Holstein-Friesian cows. *Journal of Dairy Science* **92**, 1469–1478.
- Stronge AJH, Sreenan JM, Diskin MG, Mee JF, Kenny DA and Morris DG (2005) Post-insemination milk progesterone concentration and embryo survival in dairy cows. *Theriogenology* **64**, 1212–1224.
- Szenci O, Humblot P, Beckers JF, Sasser G, Sulon J, Baltussen R, Varga J, Bajcsy CSA and Taverne MAM (2000) Plasma profiles of progesterone and conceptus proteins in cows with spontaneous embryonic/fetal mortality as diagnosed by ultrasonography. *The Veterinary Journal* **159**, 287–290.
- Tan C, Bian C, Yang D, Li N, Wu ZF and Hu XX (2017) Application of genomic selection in farm animal breeding. *Yi Chuan* **39**, 1033–1045.
- Tedde V, Bronzo V, Puggioni GMG, Pollera C, Casula A, Curone G, Moroni P, Uzzau S and Addis MF (2019) Milk cathelicidin and somatic cell counts in dairy goats along the course of lactation. *Journal of Dairy Research* **86**, 217–221.
- Thilting-Hansen T, Jorgensen RJ and Ostergaard S (2002) Milk fever control principles: a review. *Acta Veterinaria Scandinavica* **43**, 1–19.
- Thomas FC, Mudaliar M, Tassi R, McNeilly TN, Burchmore R, Burgess K, Herzyk P, Zadoks RN and Eckersall PD (2016a) Mastitomics, the integrated omics of bovine milk in an experimental model of *Streptococcus uberis* mastitis: 3. Untargeted metabolomics. *Molecular Biosystems* **12**, 2762–2769.
- Thomas FC, Mullen W, Tassi R, Ramirez-Torres A, Mudaliar M, McNeilly TN, Zadoks RN, Burchmore R and David Eckersall P (2016b) Mastitomics, the integrated omics of bovine milk in an experimental model of *Streptococcus uberis* mastitis: 1. High abundance proteins, acute phase proteins and peptidomics. *Molecular Biosystems* **12**, 2735–2747.
- Toledo-Alvarado H, Vazquez AI, de los Campos G, Tempelman RJ, Gabai G, Cecchinato A and Bittante G (2018) Changes in milk characteristics and fatty acid profile during the estrous cycle in dairy cows. *Journal of Dairy Science* **101**, 9135–9153.
- van Knegsel ATM, van den Brand H, Dijkstra J, Tamminga S and Kemp B (2005) Effect of dietary energy source on energy balance, production, metabolic disorders and reproduction in lactating dairy cattle. *Reproduction Nutrition Development* **45**, 665–688.
- Vasileiou NGC, Chatzopoulos DC, Sarrou S, Fragkou IA, Katsafadou AI, Mavrogianni VS, Petinaki E and Fthenakis GC (2019) Role of staphylococci in mastitis in sheep. *Journal of Dairy Research* **86**, 254–266.
- Veronesi MC, Gabai G, Battocchio M, Mollo A, Soldano F, Bono G and Cairoli F (2002) Ultrasonographic appearance of tissue is a better indicator of CL function than CL diameter measurement in dairy cows. *Theriogenology* **58**, 61–68.
- Vlček M, Žitný J and Kasarda R (2016) Changes of fat-to-protein ratio from start to the midlactation and the impact on milk yield. *Journal of Central European Agriculture* **17**, 1194–1203.
- Wall SK, Hernández-Castellano LE, Ahmadpour A, Bruckmaier RM and Wellnitz O (2016a) Differential glucocorticoid-induced closure of the blood-milk barrier during lipopolysaccharide- and lipoteichoic acid-induced mastitis in dairy cows. *Journal of Dairy Science* **99**, 7544–7553.
- Wall SK, Wellnitz O, Hernández-Castellano LE, Ahmadpour A and Bruckmaier RM (2016b) Superphysiological oxytocin increases the transfer of immunoglobulins and other blood components to milk during lipopolysaccharide- and lipoteichoic acid-induced mastitis in dairy cows. *Journal of Dairy Science* **99**, 9165–9173.
- Wall SK, Wellnitz O, Bruckmaier RM and Schwarz D (2018) Differential somatic cell count in milk before, during, and after lipopolysaccharide- and lipoteichoic-acid-induced mastitis in dairy cows. *Journal of Dairy Science* **101**, 5362–5373.
- Wallace RM, Pohler KG, Smith MF and Green JA (2015) Placental PAGs: gene origins, expression patterns, and use as marker of pregnancy. *Reproduction* **149**, R115–R126.
- Walsh SW, Williams EJ and Evans ACO (2011) A review of the causes of poor fertility in high milk producing dairy cows. *Animal Reproduction Science* **123**, 127–138.
- Weaver SR, Prichard AS, Maerz NL, Prichard AP, Endres EL, Hernández-Castellano LE, Akins MS, Bruckmaier RM and Hernández LL (2017) Elevating serotonin pre-partum alters the Holstein dairy cow hepatic adaptation to lactation. *PLoS One* **12**, e0184939.
- Xu YF, Velasco-Garcia M and Mottram TT (2005) Quantitative analysis of the response of an electrochemical biosensor for progesterone in milk. *Biosensors and Bioelectronics* **20**, 2061–2070.
- Xu X, Liang F, Shi J, Zhao X, Liu Z, Wu L, Song Y, Zhang H and Wang Z (2013) Determination of hormones in milk by hollow fiber-based stirring extraction bar liquid-liquid microextraction gas chromatography mass spectrometry. *Analytica Chimica Acta* **790**, 39–46.
- Zachut M, Kra G, Portnik Y, Shapiro F and Silanikove N (2016) Milk glucose-6-phosphate dehydrogenase activity and glucose-6-phosphate are associated with oxidative stress and serve as indicators of energy balance in dairy cows. *Royal Society of Chemistry Advances* **6**, 65412–65417.
- Zachut M, Kra G, Livshitz L, Portnik Y, Yakoby S, Friedlander G and Levin Y (2017) Seasonal heat stress affects adipose tissue proteome toward enrichment of the Nrf2-mediated oxidative stress response in late-pregnant dairy cows. *Journal of Proteomics* **158**, 52–61.
- Zarrin M, De Matteis L, Vernay MC, Wellnitz O, van Dorland HA and Bruckmaier RM (2013) Long-term elevation of beta-hydroxybutyrate in dairy cows through infusion: effects on feed intake, milk production, and metabolism. *Journal of Dairy Science* **96**, 2960–2972.
- Zhao F-Q (2014) Biology of glucose transport in the mammary gland. *Journal of Mammary Gland Biology and Neoplasia* **19**, 3–17.