



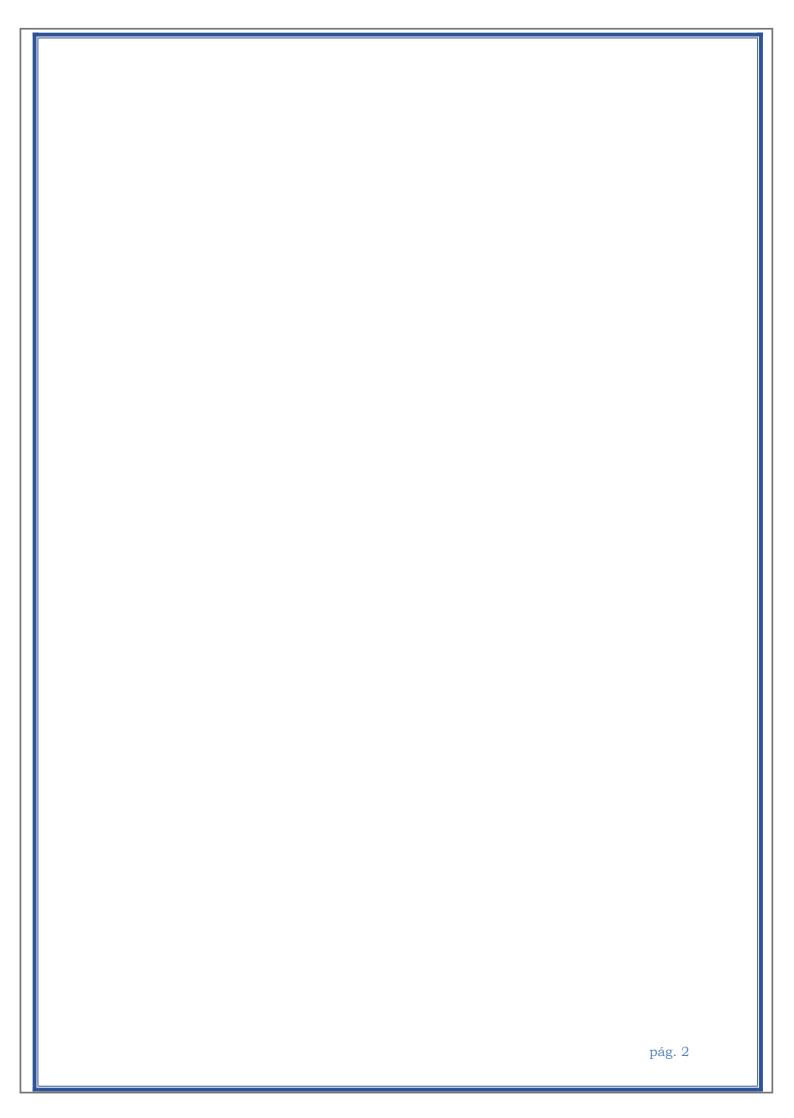


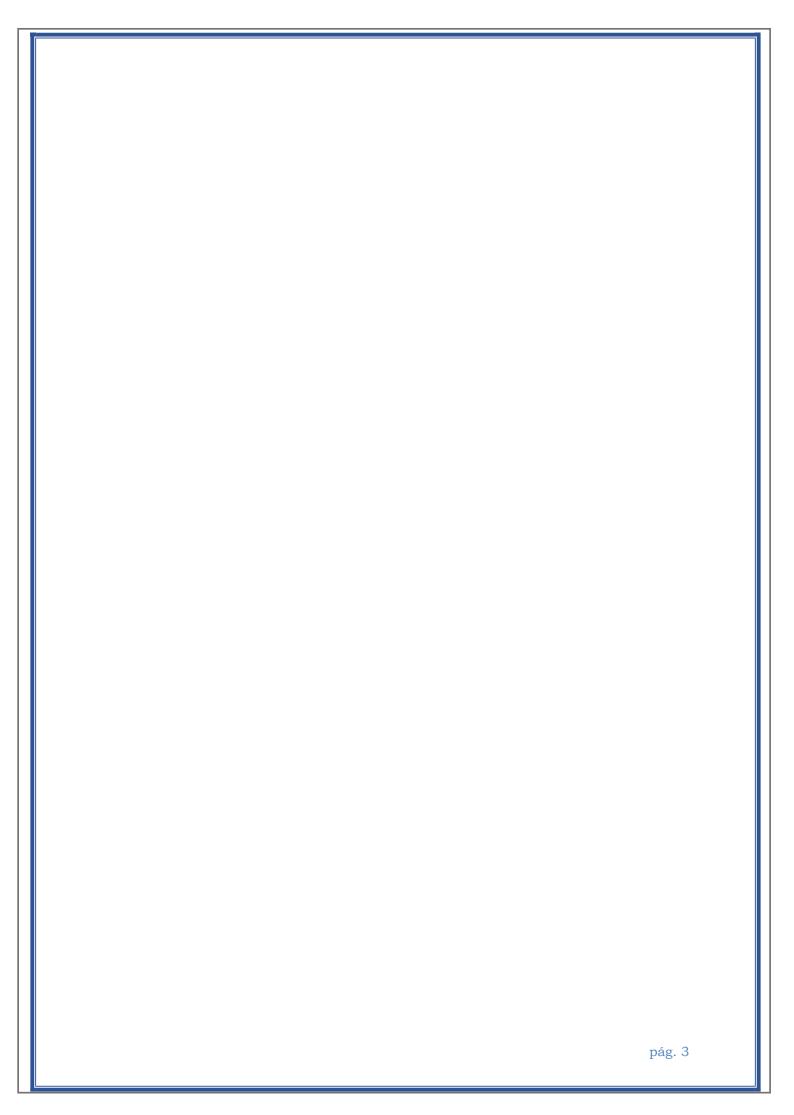
PROGRAMA DE DOCTORADO EN INVESTIGACIÓN EN BIOMEDICINA

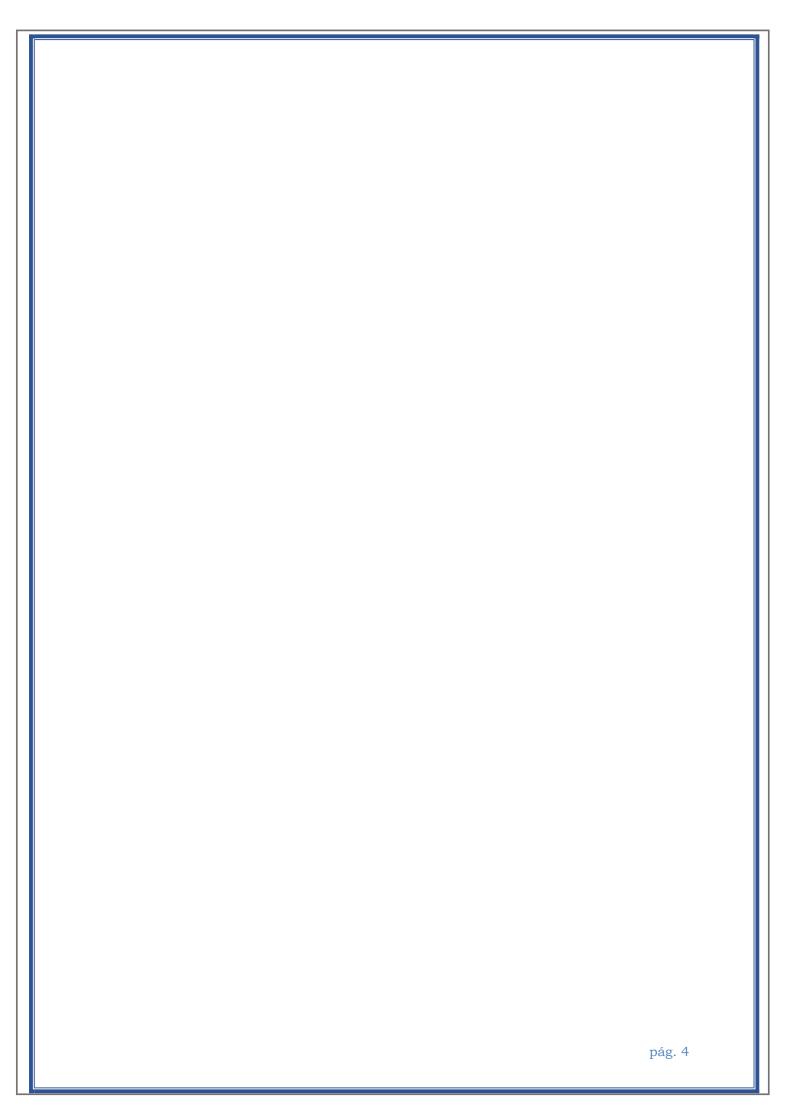
Uso del Plasma Rico en Plaquetas en Osteoartritis y fracturas en la especie canina: Aspectos funcionales y de seguridad



Sergio López Barbeta Las Palmas de Gran Canaria 2019







UNIVERSIDAD DE LAS PALMAS DE GRAN CANARIA Escuela de Doctorado

Programa de Doctorado Investigación en Biomedicina

JOSÉ ALBERTO MONTOYA ALONSO, Catedrático de Medicina Animal y COORDINADOR DEL PROGRAMA DE DOCTORADO DE INVESTIGACIÓN EN BIOMEDICINA DE LA UNIVERSIDAD DE LAS PALMAS DE GRAN CANARIA

INFORMA:

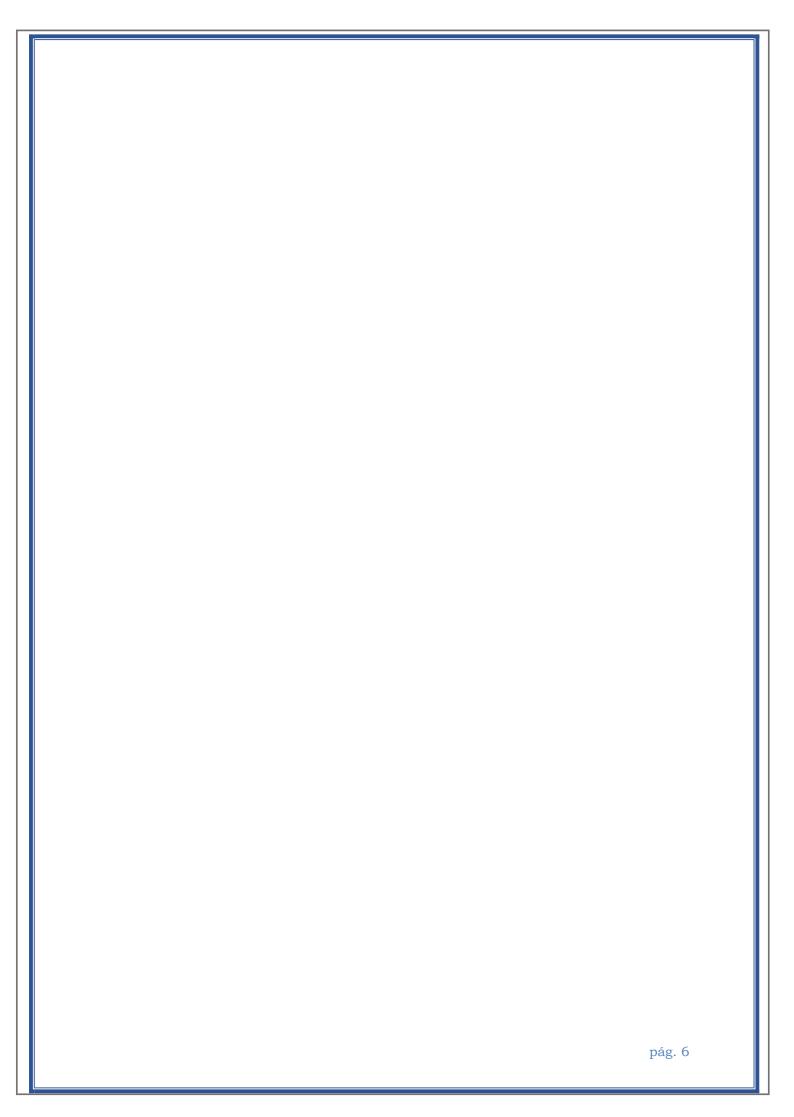
Que la Comisión Académica del Programa de Doctorado de Investigación en Biomedicina, en su sesión de fecha 9 de julio de 2019 tomó el acuerdo de dar el consentimiento para su tramitación, a la tesis doctoral titulada "Uso del Plasma Rico en Plaquetas en osteoartritis fracturas en la especie canina: Aspectos funcionales y de seguridad" presentada por el doctorando D. Sergio López barbeta y dirigida por el Dr. José Manuel Vilar Guereño.

Que la citada tesis doctoral reúne todos los requisitos exigidos por la normativa de este programa de doctorado y de esta universidad, para ser tramitada como tesis doctoral.

Y para que así conste, y a efectos de lo previsto en el Art^o 11 del Reglamento de Estudios de Doctorado (BOULPGC 7/10/2016) de la Universidad de Las Palmas de Gran Canaria, firmo el presente informe en Las Palmas de Gran Canaria, a nueve de julio de dos mil diecinueve.

MONTOYA ALONSO JOSE ALBERTO -

J. Alberto Montoya Alonso







Programa de Doctorado en Investigación en Biomedicina

TESIS DOCTORAL

Uso del Plasma Rico en Plaquetas en Osteoartritis y Fracturas en la Especie Canina: Aspectos funcionales y de seguridad

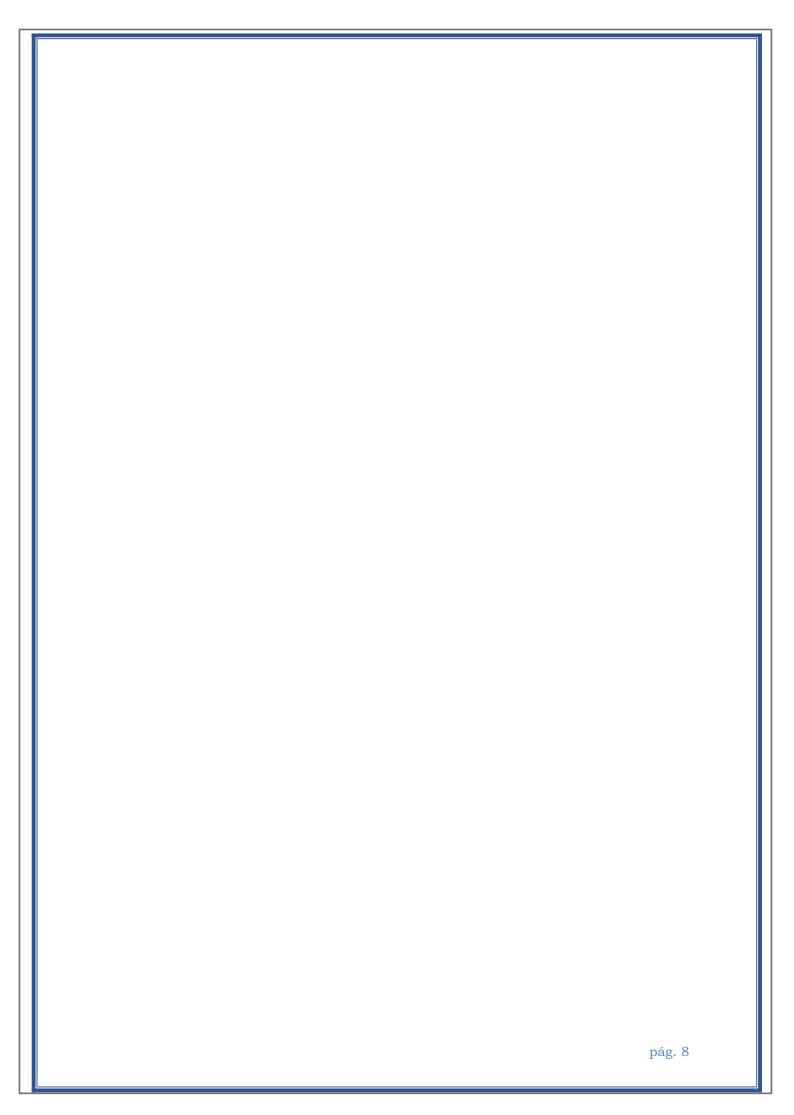
Doctorando

Sergio López Barbeta

Director

José Manuel Vilar Guereño

Las Palmas de Gran Canaria, 8 de julio de 2019







José Manuel Vilar Guereño, con DNI Doctor en veterinaria, profesor titular de cirugía veterinaria de la Facultad de Veterinaria de la Universidad de Las Palmas de Gran Canaria.

INFORMA:

Que D. Sergio López Barbeta Graduado en

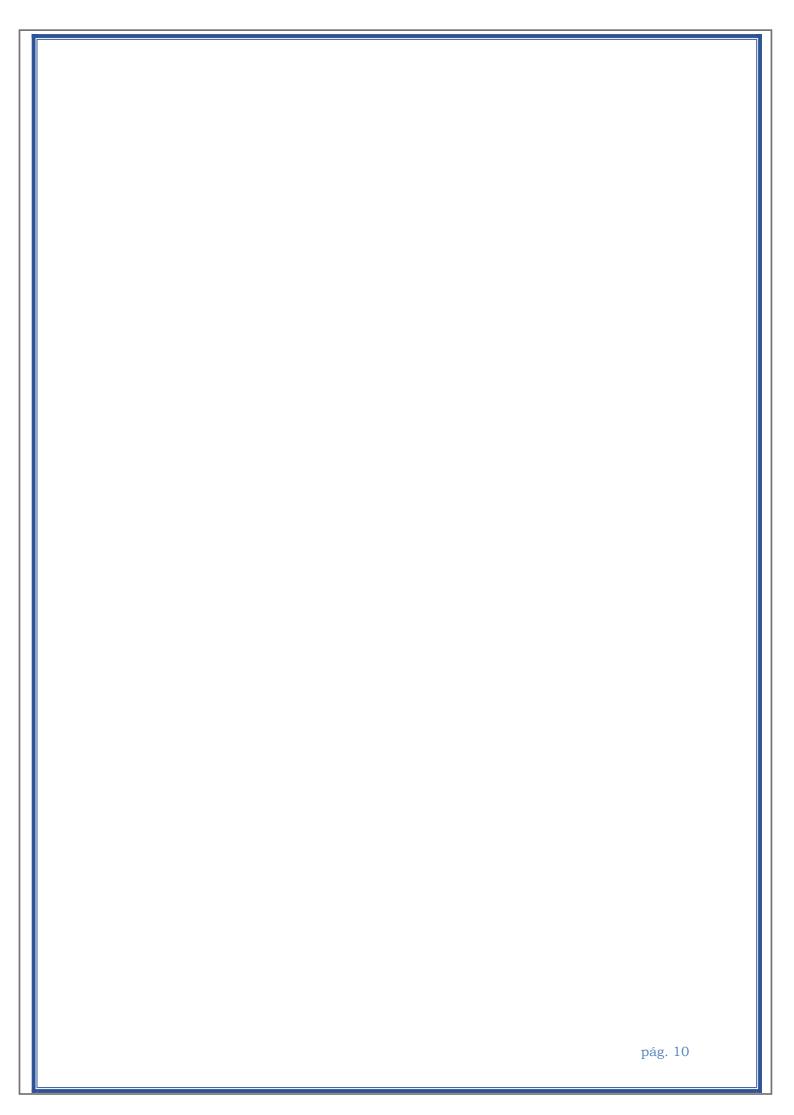
Fisioterapia, ha realizado, bajo mi dirección y asesoramiento, el presente trabajo de tesis doctoral titulado: "Uso del Plasma Rico en Plaquetas en osteoartritis y fracturas en la especie canina: Aspectos funcionales y de seguridad" que considero reúne las condiciones reglamentarias y de calidad científica necesarias, para su presentación y defensa, para optar al título de Doctor por la Universidad de Las Palmas de Gran Canaria.

Lo que firmo, a los efectos oportunos, en Arucas (Las Palmas) a nueve de julio de dos mildiecinueve

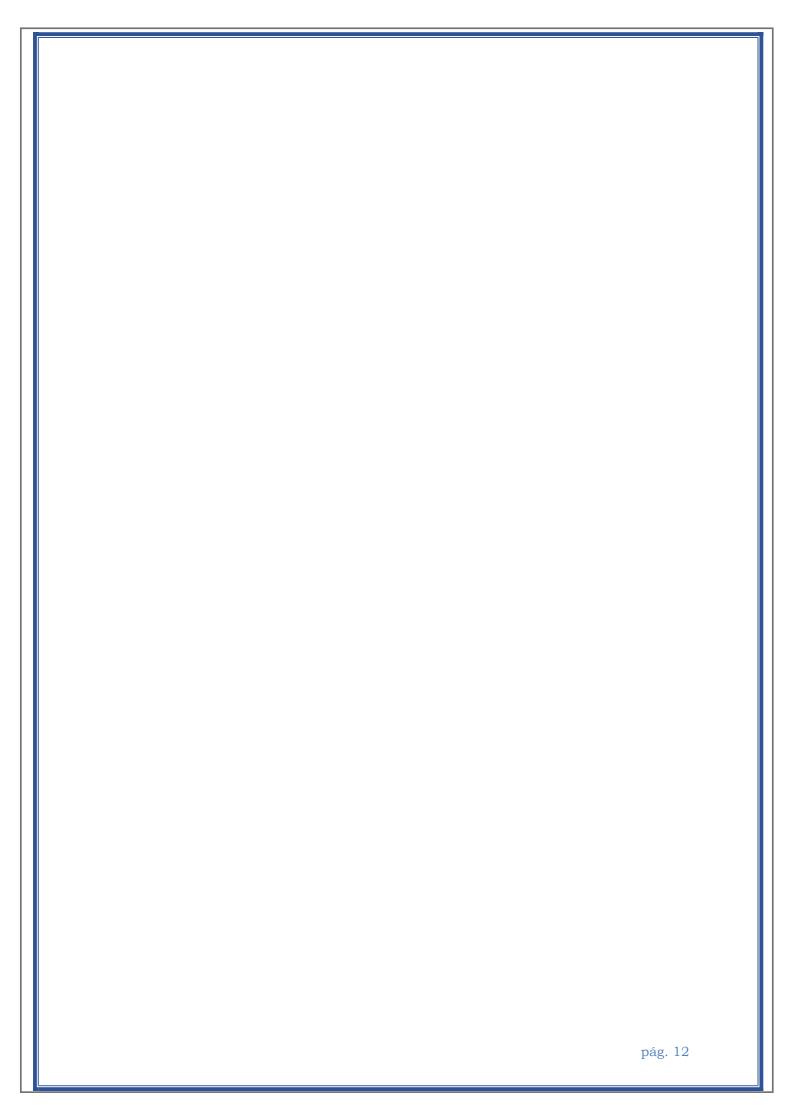
José Manuel Vilar Guereño

Profesor titular de cirugía veterinaria

35413 - ARUCAS - LAS PALMAS - ESPAÑA



A mis familiares, en especial a mis padres y abuelos, por darme la educación y valores necesarios para ser un buen hombre. A mi mujer, por ser el faro que me guía en medio de la tormenta.







La vida, tal y como la entendemos; se puede resumir en diferentes etapas que conforman un recorrido personal y transmutable, personal porque sólo cada uno de nosotros puede experimentar lo que siente, vive y conoce, transmutable porque cada experiencia vivida (buena y mala) nos trasforma, nos amolda y nos hace avanzar en el largo camino de la vida.

El doctorado, es una de esas experiencias férreas; que marcan un paso en la vida de toda persona que se inicia en el campo del conocimiento, y que busca, de una manera palpable el avance propio y porque no de toda la humanidad, aportando un granito de arena en lo que tiene que ser un movimiento colectivo.

En mi caso, siento que una etapa de mi vida se cierra, pero que otra se abre; que he llegado a poder vislumbrar una luz de conocimiento sin que esta cegara mis ojos, con la firme convicción de seguir subiendo peldaño a peldaño, la sinuosa escalera que me aleja de las sombras de la ignorancia.

Quisiera pues, agradecer de una manera íntima y personal a todas las figuras que han sido y son importantes en esta maravillosa etapa de mi vida; en primer lugar, agradecer a José Manuel Vilar, mi tutor, maestro y compañero que la diosa fortuna tuvo el

beneplácito de poner en mi camino y que sin él nada de esto hubiera sido posible.

A mis padres, Enrique y Tere, por confiar su tiempo, dinero e incluso ilusiones frustradas en que no dejara de avanzar y luchar por conseguir mis sueños.

Por último, a mi mujer Itzel, quién sostiene como un pilar la bóveda de nuestra vida, soportando juntos las enormes cargas que ésta reparte diariamente sobre nosotros, con paciencia, amor y sabiduría a partes iguales, siendo la pieza maestra sin la cual nada tendría sentido.





ÍNDICE





ÍNDICE

1. Introducción.Definición	. Pág. 25-32
2. Articulaciones afectadas de OA y patología	Pág. 33-40
2.1. Articulación coxofemoral	Pág. 35-36
2.2. Articulaciones del codo	Pág. 36-38
2.3. Articulación de la rodilla	Pág. 38-40
3. Diagnóstico de la OA	Pág. 41-64
3.1. Técnicas de imagen	Pág. 43
3.1.1. Radiología	Pág. 43
3.1.2. Artroscopia	Pág. 43-44
3.2. Técnicas de evaluación funcional	Pág. 44
3.2.1. Método de análisis subjetivo	Pág. 44-47
3.2.2. Análisis objetivo (biomecánico)	. Pág. 47-53
. a. Cinemático	
b. Electrogoniometría	
c. Cinematografía de alta velocidad y videogra	ıfia
d. Sensores inerciales o unidades de medición	n inercial (IMU)
3.2.3. Análisis cinético	Pág. 53-64
a. Plataforma de Fuerza	
b. Plataforma de Presión	
c. Estudio del COPN	
Art #1: Center of pressure limb path differences for the detection of la	ameness in dogs:
a preliminary study	Pág. 67-76
4. Alternativas terapéuticas para la OA	Pág. 77
4.1. Tratamientos convencionales	Pág. 77-78
4.2. Terapias regenerativas	Pág. 78
4.2.1 Células madre mesenquimales	Pág. 78-80



Art #2 Adipose derived-mesenchymal-stem-cells-Are-they-a-good-therap	peutic-strategy-
for-osteoarthritis?	Pág. 81-98
4.2.2 Plasma rico en Plaquetas (PRP)	Pág. 99
4.2.2.1 Indicaciones del aparato locomotor	Pág. 99
Art #3 Assessment of the Efficacy of Platelet-Rich Plasma in the Treatm	ent of
Traumatic Canine Fractures	Pág. 101-114
4.2.2.2. Seguridad del PRP, efectos secundarios	Pág. 115-116
Art #4Can-plasma-rich-in-growth-factors-be-safe-for-parental-use-A-sa	afety-study-in-
the-canine-mode.	Pág. 117-130
5. CONCLUSIONES	Pág. 131-134
6. RESUMEN	Pág. 135-140
7. SUMMARY	Pág. 141-146
8. REFERENCIAS BIBLIOGRÁFICAS	Pág. 147-166







INTRODUCCIÓN



1. INTRODUCCION Y OBJETIVOS

La osteoartritis (OA) es una patología cuya morbilidad en la especie canina aumenta constantemente, sobre todo debido al incremento de la esperanza de vida en esta especie.

La OA causa dolor de intensidad variable lo que se traduce en alteraciones en la locomoción que se traducen en cojera. La evaluación de dicha disfunción suele valorarse tradicionalmente mediante la inspección visual y, por lo tanto, subjetiva.

La radiología ha sido, dentro de los sistemas de diagnóstico por imagen, la técnica más usada. Aunque efectiva, por desgracia no existe una gran correlación entre los hallazgos lesionales por radiología y los signos clínicos. Otras técnicas más actuales como la Tomografía Computarizada y la Resonancia Magnética aportan muchos más datos, sobre todo por sus características tomográficas, pero la correlación sigue sin ser alta.

Para valorar la funcionalidad, se han creado una serie de escalas numéricas visuales que tratan de cuantificar aspectos funcionales, comportamentales e incluso radiológicos. Validadas muchas de ellas, son susceptibles de sufrir variaciones en la valoración dependiendo del observador que las realice, e incluso variaciones intraobservador, por lo que la subjetividad está presente.

En este sentido, cobran especial relevancia las diferentes técnicas biomecánicas para analizar las alteraciones locomotrices. No solo para establecer de forma

objetiva la presencia o no de cojera, sino incluso a la hora de cuantificar la eficacia de un tratamiento en un animal con cojera y poder compararlo con otros en términos de potencia y duración del efecto.

Dentro de las técnicas cinéticas, las fuerzas de reacción del suelo (GRF) están consideradas como el "Gold Standard" en la evaluación del movimiento. Estos datos se han proporcionado clásicamente mediante el uso de la plataforma de fuerza.

En tiempos más recientes, la plataforma de presión, gracias a la presencia de miles de sensores, es capaz de proporcionar muchos más datos cinéticos, entre los que queremos destacar el centro de presiones (COP). El estudio de la trayectoria del COP en los miembros de perros afectados decojera unilateral por OA constituirá por lo tanto nuestro primer objetivo, y el primer artículo aportado en la presente tesis Doctoral.

Dentro de las estrategias desde el punto de vista médico para tratar la cojera, no solo debida a AO sino también por otras patologías; se han venido utilizando de forma clásica los AINES; no carentes de efectos secundarios. Actualmente, se investigan alternativas terapéuticas lo más inocuas y eficaces posibles. En este sentido, las terapias regenerativas-reparativas están adquiriendo un auge importante en los últimos años. De ellas, las células madre mesenquimales se han postulado como una terapia eficaz en los casos de OA, lo que nos ha llevado a publicar nuestro segundo artículo, en este caso de revisión, en el que se discuten estos aspectos. Otro tratamiento muy actual dentro de las terapias regenerativas-reparativas, es el elaborado a partir de

Plasma rico en Plaquetas (PRP) y sus derivados como el Plasma rico en Factores de Crecimiento (PRGF); Ya utilizado de forma habitual en el tratamiento de OA, su eficacia para acelerar la consolidación de fracturas en la especie humana no ha sido apenas contrastada científicamente, y en el caso de la especie canina, no hay publicaciones al respecto, lo que nos lleva a plantearnos el segundo objetivo, que consiste en valorar su eficacia en fracturas en perros.

Por último, un aspecto que preocupa desde el punto de vista de la presentación de efectos secundarios en animales tratados con PRP, es sobre todo el supuesto efecto ergogénico atribuido a este producto. Dicha cuestión nos lleva a plantearnos como tercer y último objetivo un estudio en el que valorar si a dosis terapéuticas, el PRP presenta dicho efecto.

Definición de osteoartritis.

La osteoartritis (OA) ha pasado a ser una de las enfermedades con mayor prevalencia, y por tanto ha aumentado enormemente su casuística tanto en medicina humana como en veterinaria (Rychel, 2010; Malek y cols, 2012). Este hecho obedece a dos factores principales: uno, el hecho de que la longevidad está aumentando de forma significativa en los últimos años; dos, porque por desgracia la obesidad también lo hace (Lawrence y cols, 2008).

Ya en el campo de la veterinaria, y en concreto en la especie canina, se ha determinado su incidencia en un 20 % en animales adultos, llegando al 80% en geriátricos (Cuervo y cols, 2016).

En los estadios iniciales, el cartílago articular comienza a aparecer rugoso, deja de ser brillante y según va avanzando la enfermedad aparecen fisuras cada vez más profundas llegando incluso a formarse ulceras (Chen y cols, 2013)

Este proceso en última instancia crea fricción entre los huesos; lo cual desencadena un proceso inflamatorio tanto agudo como crónico, presentando los tejidos en ultima instancia un marcado engrosamiento. Todos estos factores determinan una pérdida de funcionalidad (Bland, 2015; Cuervo y cols, 2015).

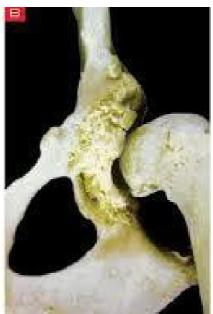
El inicio del proceso puede ser multifactorial ya que puede originarse por un traumatismo, herida, sobrecarga, desgaste, o bien asociado a un proceso de envejecimiento. Todos estos hechos, cambiaran la composición, estructura y propiedades de los tejidos que forman dicha articulación, limitando el apropiado funcionamiento del cartílago articular (Cuervo y cols, 2015).

El diagnóstico de esta patología se suele realizar mediante parámetros de tipo clínico y por radiología. Desgraciadamente, existe una falta de concordancia entre los hallazgos clínicos y radiológicos, lo que exige el uso de metodologías más objetivas (Dougados, 2005; Martel-Pelletier y cols, 2012).









ARTICULACIONES MÁS
FRECUENTEMENTE AFECTADAS
DE OA Y PATOLOGÍA MÁS COMÚN





2. ARTICULACIONES MÁS FRECUENTEMENTE AFECTADAS DE OA Y PATOLOGÍA MÁS COMÚN

En el ámbito de la clínica de pequeños animales, sobre todo en perros, las articulaciones más afectadas por OA son el codo, la cadera y la rodilla (Malek y cols, 2012). Su origen puede ser múltiple, como defectos del desarrollo o incluso traumatológicos.

2.1. articulación coxofemoral

Displasia cadera

La displasia de cadera (DC) constituye la enfermedad más frecuente en dicha articulación en la especie canina, sobre todo en perros de razas grandes y gigantes (Manera y cols, 2019). Existen aspectos muy importantes en esta patología a considerar, ya que, al ser un problema hereditario, y del desarrollo, los animales es posible que no manifiesten sintomatología hasta que son adultos; esto hace que las medidas para impedir la extensión de la enfermedad excluyendo a los animales descartados como reproductores lleguen demasiado tarde (Rodríguez, 2003).

Esta patología además se caracteriza por una falta de congruencia entre las superficies articulares; por lo que la consecuencia más directa, es que las fuerzas se distribuyan de un modo irregular en la articulación. En definitiva, la articulación se vuelve inestable lo que hace que se instaure un proceso osteoartrítico (Cardinet, 1997).





Radiografías correspondientes a un caso de displasia de grado D (a) y grado E (b). Se puede observar el aplanamiento del acetábulo, cabeza del fémur triangular en vez de esférica, esclerosis en el borde acetabular, y fragmentos óseos (osteofitos).

2.2. articulación del codo

Displasia de codo

La displasia de codo (DC) es un síndrome que engloba diferentes patologías en dicha articulación que en general, supone una incongruencia entre las superficies articulares que la forman, lo cual conlleva una inestabilidad que en último termino provocará la aparición de la osteoartritis como enfermedad articular degenerativa, sobre todo si no se aplica un tratamiento efectivo (Manera y cols, 2019). Las patologías más conocidas que provocan displasia de codo son la fragmentación del proceso coronoides, la osteocondritis disecante y por último el proceso ancóneo no unido (Tobias y Johnston, 2011).

La DC es de carácter hereditario, y multifactorial. Lo cierto es que puede desencadenarse por anomalías en el desarrollo de los centros de osificación,



anomalías en alguno de los huesos largos que forman el codo o una combinación de estos (Durante, 1998; Kirberger y Barr, 1998).



Bassethound con displasia de codo. En esta patología es típica la abducción del miembro afectado.

https://www.basset.net/boards/general-basset-hound-discussion/45337-basset-elbow-dysplasia-surgery-3.html

Esta patología es típica de razas grandes y gigantes. Es conveniente resaltar que afecta 3 veces más a machos que a hembras (Durante y Brusa, 1998; Kirberger, 2003; Komsta y cols, 2008).





Osteoartritis avanzada con neoformación ósea (flechas) así como esclerosis bajo las superficies articulares (cruz). https://www.acvs.org/small-animal/canine-elbow-dysplasia

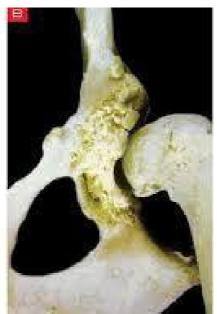
2.3. articulación de la rodilla

Dentro de esta articulación, debemos decir que la rotura del CCL es el principal motivo de cojera de miembro pelviano, y una de las más comunes de las que pueden afectar a la rodilla en la especie canina (Whitehair y cols, 1993; Johnson y cols, 1994; Duval y cols, 1999). Este ligamento proporciona estabilidad en la articulación femorotibial; por lo tanto, su rotura determina inestabilidad, que, como se ha explicado anteriormente, produce en último término la generación de un proceso osteoartrítico (Elkins y cols, 1991; Innes y Barr 1998).









DIAGNÓSTICO DE LA OA

3. DIAGNÓSTICO DE LA OA

3.1. Técnicas de imagen

3.1.1. Radiología

Las técnicas de imagen radiológicas son pruebas de gran utilidad, pues además de realizar un diagnóstico preciso de la dolencias anteriormente citadas, nos permite descubrir en qué fase de la misma se encuentran, ya que; podemos identificar diferentes signos de la osteoartritis (Vilar y cols, 2013; Bland, 2015) y aunque es cierto, que se ha demostrado que la relación entre los signos clínicos y los signos radiológicos no son siempre correlativos, si es muy recomendable en este tipo de patologías realizarlas de manera regular.

Las pruebas de imagen como la RM y el TC nos permiten visualizar de manera más detallada las diferentes estructuras articulares sin que existan superposición de capas de tejido muscular, fascia o tejido ligamentoso, lo cual es una ventaja a la hora de evaluar los cambios en las estructuras óseas o irregularidades articulares (Fossum, 1999).

3.1.2. Artroscopia

La artroscopia es una de las técnicas diagnósticas más utilizadas en displasia de codo; no en vano, aunque sigue siendo una técnica invasiva, es menos agresiva que las artrotomias y proporcionan una visión de alta calidad en las estructuras intraarticulares

(Adrian y cols, 2017; Kim y Joo, 2018) quienes, sin embargo, en lesiones de la articulación femorotibial recomiendan el uso de la ecografía en combinación con la artroscopia para acceder a las zonas articulares de gran interés clínico.

3.2. Técnicas de evaluación funcional

Como hemos podido observar anteriormente, el dolor de intensidad variable es uno de los signos más habituales de la OA. Funcionalmente hablando, este dolor va a provocar una limitación que se traduce en una alteración del paso (cojera) de menor o mayor grado. Como se ha expresado anteriormente, la correlación entre los signos clínicos y radiológicos no son de gran valor por lo que la evaluación de la funcionalidad del miembro o los miembros afectos es de vital importancia (Gordon y cols, 2003).

Si tenemos en cuenta lo que hemos argumentado anteriormente, los métodos para evaluar el estado funcional de los animales se dividen en:

3.2.1. Métodos de análisis funcional subjetivos

La manera tradicional de valorar el dolor en los animales domésticos por parte de los profesionales veterinarios, ha sido teniendo en cuenta de forma subjetiva alteraciones comportamentales, así como la valoración del movimiento en si o incluso mediante la palpación o movilización pasiva. Estos factores (y otros más) han servido a muchos investigadores para elaborar escalas de valoración numéricas, en un intento por cuantificar la discapacidad funcional del aparato locomotor.

En primer lugar, resaltamos el cuestionario denominado Bristol Osteoarthritis in Dogs (BrOAD) (Innes and Barr, 1998) o el Texas A&M Client (Hudson y cols, 2004). Son escalas análogas validadas, cuya cuantificación depende del criterio del propietario del perro. También es muy conocido el denominado Canine Brief Pain Inventory (CBPI) ya que tiene en cuenta una gran cantidad de parámetros (Cimino Brown y cols, 2007.). En línea con esta escala se encuentra también la Helsinki Chronic Pain Index (HCPI), añade parámetros comportamentales (Hielm- Björkman y cols, 2003; Hielm- Björkman y cols, 2009; Walton y cols, 2013).

El HCPI se ha utilizado de forma continua para la evaluación del dolor crónico en perros con osteoartritis de rodilla, codo y cadera (Hielm-Björkman y cols, 2003; Hielm-Björkman y cols, 2009; Wernham y cols, 2011; Hielm-Björkman y cols, 2012; Heikkilä y cols, 2014).

En España, podemos destacar por su utilización, la escala denominada Bioarth®, ya que está basada en aspectos clínicos, radiológicos y aspectos comportamentales que deben ser cuantificados; la valoración obtenida puede aplicarse para medir por ejemplo la efectividad de un tratamiento instaurado en casos de cojera, ya sea de cadera, codo o rodilla (Vilar y cols, 2016).



FUNCTIONAL EVALUATION SCALE OF THE HIP

Pet Name

Owner name

1. CHANGES STANDING STILL:	
(0) Standing normally/ (1) Leaning on one side/ (2) Resting the tip of its paw	· Internal
(3) Non weight bearing on that limb	
2. CHANGES STANDING UP:	
(0) Standing up normally/(1) Adopting different positions when standing up/	
(2) Difficulty to rise/ (3) Does not stand up	
3. LAMENESS AT THE BEGINNING OF EXERCISE:	
(0) No lameness present/ (1) The lameness disappears when the dog moves (up to 10 minutes)	
(2) The lameness does not disappear	
4. LAMENESS AFTER WARM-UP (10 MINUTES):	
(0) No lameness present/ (1) Mild lameness/ (2) Severe lameness	
(3) Continuous non-weight-bearing lameness	
5. LAMENESS DURING THE WALK:	_ [
(0) It can walk without difficulties/ (1) It often stops while going for a walk	
(2) It can take just very short walks (less than 5 minutes)/	
(3) It does not want to go for a walk	
6. LAMENESS WHILE RUNNING AND PLAYING:	
(0) It can run and play without difficulties/	
(1) It runs and plays with difficulties or it gets tired easily	
(2) It runs with lots of difficulties under a stimulus	
(3) It neither runs nor plays under any stimulus	
7. GOING UP THE STEPS:	
(0) It goes up without difficulties/ (1) It goes up 16 steps (a flight) with difficulty	
(2) It goes up 1 or 2 steps either a kerb, with difficulty	
(3) It does not go up the steps	
8. SMALL JUMPS 40-50 CM:	_
(0) It gets on the sofa or on the car without difficulties	
(1) It gets on the sofa or on the car with difficulty	
(2) It neither gets on the sofa or on the car	
TOTAL SCORE OF FUNCTIONAL LIMITATION	

FUNCTIONAL EVALUATION SCALE OF THE HIP

et Name	
owner name	
B) RANGE OF MOVEMENT	
9. MANUAL MOBILIZATION PRODUCES:	
(0) No pain and no crepitation/ (1) There is pain on the last stages	
(2) There is pain and/or crepitation during the process	
(3) It cannot be carried out or there is severe pain and crepitation	
10. ROM IN FLEXION:	
(0) Total flexion 50-60°/ (1) Mild limitation $<\!80^{\circ}\!/$ (2) Severe limitation $>\!80^{\circ}\!$	
11. ROM IN EXTENSION:	
(0) Total extension 160-170°/ (1) Mild limitation>150°/ (2) Severe limitation <150°	
TOTAL SCORE OF THE RANGE OF MOVEMENT (sum of scores 9-11)	
C) MUSCULAR ATROPHY:	
(0) There is no muscular atrophy/ (1) Mild atrophy/ (2) Severe atrophy	
TOTAL SCORE (sum of A+B+C)	

Escala Bioarth® para la evaluación de la articulación coxofemoral.

Todas las escalas anteriormente citadas tienen en cuenta en mayor o menor medida factores subjetivos, lo que hace que pueda existir de forma potencial una gran variabilidad intraobservador y/o interobservador que puede alterar de forma significativa la valoración final (Horstam y cols, 2004).

3.2.2. Análisis objetivo (Biomecánica)

Teniendo en cuenta las afirmaciones anteriores, una valoración objetiva sobre el estado alterado del paso (cojera) del animal, así como su diagnóstico y

progresión mientras está siendo tratado son parámetros muy actuales desde el punto de vista científico (Mölsä y cols, 2010).

La metodología de evaluación biomecánica nos ofrece una medición de los procesos que generan la locomoción.

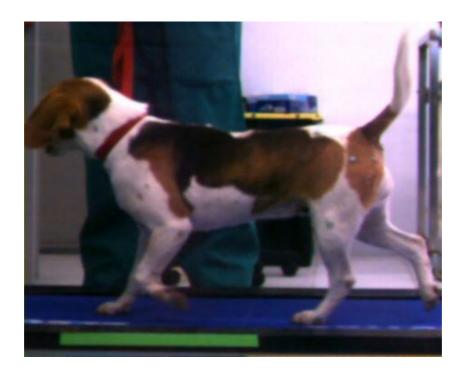
Estos métodos están divididos en métodos cinemáticos y cinéticos.

Sin embargo, debemos hacer hincapié en el uso del tapiz rodante o Treadmill antes de la utilización otras tecnologías, pues supone un complemento muy interesante desde el punto de vista de la estandarización de las condiciones del estudio como velocidad, inclinación, etc. Evidentemente, los animales deben pasar previamente por un periodo de acostumbramiento o habituación. (Khumsap y cols, 2004; Cruz y cols, 2017).

Además, nos permite obtener un registro de pasos consecutivos, muy útil en la valoración final (Drevemo y cols, 1980).

Recientemente, un estudio (Vilar y cols, 2016) ha utilizado el Treadmill para establecer las diferentes características dinámicas, en concreto las características cinéticas y cinemáticas de varias razas de perro con una morfología completamente diferente.

El método consistió en grabar a los animales con una cámara de alta velocidad al paso, para obtener los datos cinemáticos. Para obtener los datos de tipo cinético, se insertó una plataforma de fuerza sobre el tapiz rodante.



Treadmill utilizado para evaluar el movimiento de un perro de raza Beagle. Existe una plataforma de fuerza insertada debajo de la cinta rodante, y para la toma de datos cinemáticos se han aplicado unos reflectantes en los centros de rotación articular.



http://trends.medicalexpo.com/project-45657.html

En esta imagen podemos observar que el Treadmill, también es utilizado en las fases de rehabilitación animal, incluso con los miembros en inmersión.

http://trends.medicalexpo.com/project-45657.html

A pesar de que es una técnica realmente útil y efectiva, presenta una serie de inconvenientes y limitaciones, principalmente en la posible alteración del movimiento natural que el animal realizar sobre un suelo normal.

Cinemáticos

La cinemática se encarga de evaluar el movimiento, pero sin tener en cuenta las fuerzas que lo provocan. Los parámetros obtenidos pueden ser lineares (amplitud de paso, altura del miembro durante la fase de vuelo...), angulares (ángulos de flexión, extensión, rango articular de movimiento, velocidades y aceleraciones angulares...) y temporales para el cálculo de la duración de las diferentes fases de la marcha (apoyo, vuelo, etc....) así como frecuencias, etc. (Manera, 2019).

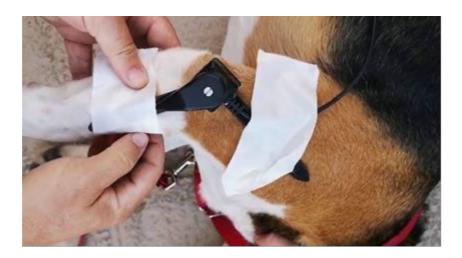
Electrogoniometría.

Los electrogoniómetros son instrumentos que se sitúan sobre los centros de rotación articular para obtener los datos angulares de forma simultánea al movimiento del animal mientras se está desplazando. Su utilización en pequeños animales es bastante escasa (Thomas y cols, 2006; Jaeger y cols, 2007).

El electrogoniómetro genera una corriente eléctrica proporcional a la rotación que se esté produciendo, de modo que podremos obtener diferentes parámetros angulares (ángulo, velocidad angular, aceleración angular...).

A pesar de ser un instrumento preciso y de gran interés por sus datos fiables, existen dos factores fundamentales que limitan su precisión: uno es el posible desplazamiento respecto al eje de rotación que se puede producir por el propio movimiento, y el segundo es que hay articulaciones que realizan movimientos

en sentido tridimensional, no pudiéndose detectar por este instrumento (Ratzlaff, 1989).



Colocación de un electrogoniómetro sobre la articulación del codo

Cinematografía de alta velocidad y videografía.

La cinematografía ha quedado superada hoy en día por técnicas más actualizadas como la videografía de alta velocidad; ésta es probablemente el método más usado para la investigación biomecánica en pequeños y grandes animales (Keegan y cols, 2000; Kramer y cols, 2004; Vilar y cols, 2010).

Los principales datos cinemáticos pueden ser obtenidos con esta metodología (ángulos, tiempos de las distintas fases del paso, ángulos articulares en 2D y 3D, etc.).

En dichas determinaciones se utilizan unos marcadores reflectantes fijados a la piel sobre los centros de rotación de las articulaciones. Los datos

anteriormente citados se pueden extraer de forma manual, o existen metodologías actuales que permiten hacerlo de forma automatizada. (Langlois y cols, 1978; Leach y Cymbaluk, 1986; Leach y Dyson, 1988; Kramer y Keegan, 2007). La principal limitación es que, al estar los dispositivos fijados en la piel, el movimiento puede alterar la posición de los marcadores reflectantes (Van Weeren y cols, 1990).

Sensores inerciales o unidades de medición inercial (IMU).

Estos dispositivos obtienen datos cinemáticos como la aceleración lineal (acelerómetros) o velocidad angular (giroscopios). Son de tamaño muy reducido. Los datos registrados se emiten de forma inalámbrica en tiempo real (Martínez-Méndez y Huertas, 2013).

En la especie equina se pueden encontrar un alto número de trabajos de investigación con esta instrumentación, ya que se ha estandarizado su uso para la detección de cojeras (Keegan y cols, 2002; Watanabe y cols, 2011).

Debido a que pueden registrarse simultáneamente los datos de varios de estos dispositivos, su fijación en varias regiones corporales además de los miembros ha contribuido de forma determinante al conocimiento de los mecanismos de redistribución de fuerza compensatorios en los casos de cojera (Maliye y cols, 2013; Rhodin y cols, 2017).

3.2.3. Análisis cinético

La cinética es aquella parte de la biomecánica que se encarga de analizar el movimiento, pero en este caso se ocupa específicamente de las fuerzas que lo

están produciendo. De todas ellas, concretamente las fuerzas de reacción del suelo (GRF por sus siglas en ingles), adquieren una especial importancia por serlas más utilizadas. Además, a partir de dichas fuerzas se podrá analizar el centro de presiones (COP). Su relación se analizará a continuación:

- El pico de fuerza vertical (PVF, por sus siglas en inglés) es la GRF más utilizada en los análisis cinéticos y se define como el valor más alto de fuerza (en Newton o quilogramos) que se aplica contra el suelo cuando el miembro está apoyando durante la progresión del movimiento; el impulso vertical (VI, por sus siglas en inglés) es el producto de la fuerza por el tiempo de apoyo. El IV se evidencia en la gráfica que se genera durante el apoyo por ser el área bajo la curva. La objetividad de este parámetro en el diagnóstico de cojeras no está todavía clarificada (Vilar y cols, 2013).

-El centro de masas (COM) o centro de gravedad (CG) se define como la suma de las trayectorias de todos los segmentos de fuerza del cuerpo (cráneo/caudal y latero/medial) (Winter y cols, 1991).

-**El COP**, que supondría la posición en el plano de apoyo del CG, sea del cuerpo o de una parte del organismo como por ejemplo un miembro (Winter y cols, 1996); De este modo, el COP representa de forma explícita la fuerza que se está ejerciendo en un área concreta (Baratto y cols, 2002).

Plataforma de fuerza

Las plataformas de fuerza son unos dispositivos que a través de medidores dinamométricos o piezoeléctricos insertados (normalmente 4) en una estructura de forma cuadrada son capaces de medir la fuerza durante la fase de apoyo. Dichos medidores generaran un impulso eléctrico de forma proporcional a la fuerza ejercida durante la pisada.



Plataforma dinamométrica de fuerza con 4 sensores de la marca Pasco®

Como se había descrito anteriormente, la visualización subjetiva del movimiento tiene como gran limitación la poca discriminación que hace entre cojeras leves o incluso moderadas, e incluso que estas no puedan ser detectadas; esto hace que en estos casos la evaluación del movimiento con las plataformas s de fuerza adquiera una especial importancia (Evans y colsm, 2005). Para poder llevarlo a cabo, los parámetros más utilizados van a ser los citados anteriormente, es decir, el PVF y el VI, considerándose hoy en día el "Gold Standard" (Walton y cols, 2014) para la evaluación del movimiento.

Debido a que las GRF pueden ser modificadas por la velocidad a la que el sujeto se desplaza, los análisis para ser validos deben ser realizados en un

rango muy estrecho de velocidad e incluso de aceleración (Riggs y cols, 1993; Budsberg y cols, 1999).

Otro factor que va a influir en las GRF es el peso, por lo que los estudios científicos, a la hora de obtener valores validados no solo debe hacer acopio de animales con pesos (e incluso conformaciones) muy similares, sino que se hace necesario realizar procesos de normalización; el más utilizado en este campo es el de expresar los valores en relación con el peso corporal (%) (Vilar y cols, 2013).

Normalmente, y de forma genérica se debe decir que en los cuadrúpedos siempre, los miembros torácicos dan valores de PVF más altos que los pelvianos, ya que estos animales el CG está desplazado cranealmente.

En el campo clínico, este dispositivo se viene utilizando de forma regular para evaluar la respuesta a los tratamientos instaurados en animales con osteoartritis derivadas de patologías como la displasia de cadera, codo, e incluso la CCLR (Nelson y cols, 2013; Skinner y cols, 2013; Vilar y cols, 2013; Wurcherer y cols, 2013).

Las plataformas de fuerza presentan como limitación principal que los datos se pueden obtener solamente de un miembro aislado, lo que hace que los exámenes sean largos y por lo tanto dificultosos cuando este estudio se realiza con animales; evidentemente, el hecho de que solo mida la fuerza generada durante la fase de apoyo supone otra gran limitación, ya que en la fase de vuelo también se generan fuerzas. Este dispositivo, debido a su tamaño, tampoco obtiene datos de apoyos consecutivos. (Manera y cols, 2019).

Plataforma de presión

La plataforma de presión es un dispositivo de estudio tanto en el campo clínico como en el investigador que se ha ido añadiendo progresivamente en los últimos años al elenco de dispositivos para obtener GRF. Su principal característica es que dispone de miles de sensores distribuidos por el área de estudio de modo que los datos son específicos de áreas determinadas. Por otro lado, y de forma análoga a las plataformas de fuerza, se puede regular la velocidad de adquisición de datos (Oosterlinck y cols, 2010).

Estos dispositivos no son más costosos que los anteriores, pero adquiere como ventaja importante, el hecho de que pueden ser de grandes dimensiones (hasta 2 metros de largo) por lo que se pueden registrar los datos de varios apoyos.

Otra gran ventaja es que además de los clásicos parámetros (PVF y VI) se pueden obtener otros que cada vez adquieren mayor relevancia como como la distribución del peso en el cuerpo, la presión media y máxima de apoyo, que al estudiarse de forma unificada conforman el análisis podobarométrico (Oosterlinck y cols, 2011; Carr y cols, 2015; Bockstahler y cols, 2016; Schnabl-Feichter y cols, 2017).

Como se podrá ver a continuación, este dispositivo podrá evaluar la evolución del COP durante la marcha (Gomes-Costa y cols, 2015), cuyo análisis en animales que presentan cojera constituye uno de nuestros objetivos principales en la presente tesis.



Las plataformas de presión de gran longitud (hasta 2 metros) permiten el registro de varios pasos consecutivos. Plataforma tekscan.

https://twitter.com/vetbiomechanics/status/666602613715173376

Como viene siendo habitual, muchos de estos dispositivos han venido utilizándose de forma precedente en medicina humana; entre los estudios consultados quisiéramos destacar aquellos en donde las plataformas de presión han servido para determinar cómo patologías como la esclerosis múltiple pueden afectar al mantenimiento postural (Abrantes y Santos, 2012). Pero otro tipo de patologías crónicas como la diabetes también influyen en la postura y por lo tanto en la distribución de la presión durante el apoyo (Anjos y cols, 2010).

Ya en el campo de la veterinaria, es en el mundo del caballo donde estas plataformas se empezaron a utilizar, destacando la profundización en los eventos cinéticos concretos que se producen durante la fase de apoyo del casco (Van Heel y cols, 2005), así como los efectos del recorte del casco en la distribución de presiones en la suela (Moleman y cols, 2006), o la simetría del casco en caballos y ponis sanos (Oosterlinck y cols, 2010a;2010b).

En estos últimos años existe una tendencia hacia el estudio de cojeras, pero en este caso nos basamos en los cambios posturales con el animal inmóvil. La consecuencia (y ventaja) más evidente es que este tipo de determinaciones se pueden realizar en espacios pequeños, ya que no es necesario que el animal realice movimientos de desplazamiento (Gomes-Costa, 2015).

En pequeños animales las plataformas de presión se han empezado a utilizar de forma muy reciente, sobre todo en estudios descriptivos del paso en animales sin patologías locomotrices (Marghitu y cols, 2003; Souza y cols, 2013), en algunos estudios posteriores ya utilizan esta tecnología para estudios en perros con CCLR (Souza y cols, 2014), osteoartritis de cadera (Upchurch y cols, 2016) o en análisis del movimiento en perros a los que se le habían implantado prótesis en dicha articulaciones (Tomas y cols, 2014).

Como decíamos anteriormente, las plataformas de presión permiten obtener datos de diferentes miembros simultáneamente y en pasos consecutivos, lo que ha facilitado el análisis de las cojeras mediante el uso de fórmulas matemáticas encaminadas a obtener los niveles de simetría entre miembros (Carr y cols, 2015).

El desarrollo de estos instrumentos hace que se aumente de forma acelerada su potencial, que se consigue fundamentalmente con la producción de dispositivos de tamaños, pero, sobre todo, de resolución diferentes, lo que permite actualmente obtener datos de animales con áreas de apoyo muy pequeñas, como es el caso de los gatos (Schnabl-Feichter y cols, 2017).

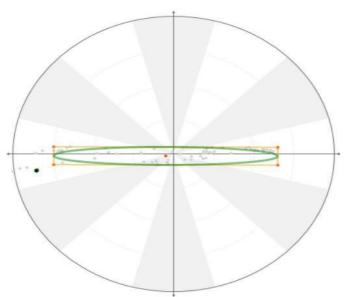
Entre las limitaciones que poseen las plataformas de presión cabe destacar la lenta respuesta de los sensores; por otro lado, un estudio realizado en caballos ha revelado que cuando se comparan datos de GRF obtenidos en los mismos individuos de forma simultánea con una plataforma de fuerza y una de presión, esta última arroja resultados más bajos (Oosterlinck y cols, 2010).

Como habíamos adelantado anteriormente, aunque basados en principios físicos diferentes, podíamos asumir que la posición del COP coincidía con la proyección vertical del CG sobre el plano donde los miembros están apoyados (Baratto y cols, 2002). En un animal en la estación, el COP va a describir una trayectoria que se corresponde a los movimientos de desestabilización y recuperación de la postura. Su análisis en los planos X e Y se denomina estabilograma. Esta trayectoria en el plano de apoyo va a ocupar un área determinada, mas pequeña cuanta más estabilidad esté presente; al grafico generado por la trayectoria del COP se le denomina estatocinesiograma.

¿Entonces, cómo se altera la trayectoria del COP cuando existe cojera? De forma experimental, y ya en el campo de la veterinaria, se ha demostrado que en la estación se manifiestan unas alteraciones posturales debido a los esfuerzos por aliviar el dolor transfiriendo la presión hacia el lado sano. Evidentemente su sistema nervioso corregirá de forma totalmente refleja esas alteraciones posturales, creando en definitiva, alteraciones en las características del COP (trayectoria, área del estatocinesiograma, velocidad del desplazamiento del COP, etc.) generándose un ciclo continuo de perturbaciones-correcciones; estas alteraciones, no solo se han evidenciado en la especie

humana sino que, como apuntábamos anteriormente, se ha verificado en canidos y équidos (Buchner y cols, 2001; Oosterlinck y cols, 2010; Manera y cols, 2017). Estos eventos producidos por las desviaciones en el comportamiento del COP, tal y como se evidencia en las referencias anteriormente citadas, han servido desde el punto de vista clínico para la detección de las cojeras en estas especies.





Estatocinesiograma de un perro cojo de la derecha. Aparte de la gran superficie (61,92 mm²), se ve la elipse desplazada hacia el lado izquierdo

Evidentemente, el COP no solo describe una trayectoria debida a los eventos de alteración-corrección realizados por el SNC tanto en condiciones fisiológicas como patológicas; sino que cuando el individuo está en movimiento, el COP corporal durante toda la fase de apoyo de los miembros, este va a

describir una cierta trayectoria. La trayectoria, o, mejor dicho, las alteraciones de su trayectoria en los miembros ya han servido en la especie humana para detectar las modificaciones en este parámetro en situaciones de cojera (Riskowski y cols, 2013) Parkinson (Kim y cols, 2017) así como en hemiparesia (Robain y cols, 2006)

En las situaciones patológicas anteriores, las principales características derivadas del COP durante el desplazamiento son:

- 1) **El desplazamiento craneocaudal del COP**; es decir, la medición en un sistema de coordenadas de la distancia entre los puntos inicial y final de la trayectoria (Robain y cols, 2006).
- 2) El desplazamiento lateromedial que se va a producir en el COP; Este índice, denominado en ingles center of pressure excursion index (CPEI) a efectos de normalización, refleja este desplazamiento en relación con la anchura del miembro y se multiplica por 100 para obtener este dato en términos de porcentaje (Yoon y cols, 2010; Riskowski y cols, 2013).





TRATAMIENTO DE LA OA Y

ARTÍCULOS

TRATAMIENTO OA Y ARTÍCULOS



López S, Vilar JM, Rubio M, Sopena J, Damià E, Santana A, Carrillo JM.

Center of pressure limb path differences for the detection of

lameness in dogs: a preliminary study. BMC Vet Res. 2019 May

8;15(1):138. doi: 10.1186/s12917-019-1881-1.

JCR Impact factor (2017): 1.958

Cuartil: Q1

Grupo: Veterinary sciences

Posición: 20/140



López et al. BMC Veterinary Research (2019) 15:138 https://doi.org/10.1186/s12917-019-1881-1

BMC Veterinary Research

RESEARCH ARTICLE

Open Access

Center of pressure limb path differences for the detection of lameness in dogs: a preliminary study



Sergio López¹, José M. Vila¹¹⁴*o, Mónica Rubio², Joaquin J. Sopena², Elena Damiá², Déborah Chicharro², Angelo Santana³ and José M. Carrillo²

Abstract

Background: The limb center of pressure (COP) path measures and quantifies the load distribution within a limb in a still or moving subject. Under this premise, the aim of this study was to test whether data derived from this parameter could detect the differences between sound and lame limbs in unilaterally lame dogs with elbow dysplasia.

To accomplish this purpose, ten unilaterally lame dogs of similar conformation were walked over a pressure platform. Next, the COP path, in relation to the position of sound and lame limbs, was measured in a coordinate system over a standard paw template obtained by pedobarography during the whole support phase. To compare variables, force platform data (peak vertical force and vertical impulse) from the same animals were obtained. Sound and lame limb statokinesiograms were also obtained while the animals stood still.

Results: The statistical analysis clearly showed that COP in lame limbs start cranially and were shorter than sound limbs. In addition, the value of the COP excursion index was lower in lame limbs. Finally, the area of statokinesiograms was greater in lame limbs.

Conclusion: This methodology based in limb COP characteristics serves to discriminate between sound and lame limbs in dogs with elbow dysplasia.

Keywords: Balance, Center of pressure, COP, Dog, Statokinesiogram

Background

Various methods to analyze the locomotor status within the veterinary field have been developed in order to generate useful parameters from both kinematic and/ or kinetic perspectives. These methodologies should be able to provide accurate and reliable data and, if possible, form a set of parameters that will allow for the normal/abnormal static/dynamic events from a wide perspective. This invariably requires the use of more sophisticated systems [1].

These data should ultimately serve to detect lameness, and, among them, the center of pressure (COP) position

may be considered the net output variable of interaction between all of the forces and torques that occur in the body (bCOP) or limb (ICOP) and its inertial properties. The COP position over time is named the COP path. This parameter quantifies the dynamic load distribution under the foot [2]. The ICOP path characteristics obtained in moving subjects provide insights into foot dynamics during the support phase of gait in human and, potentially, in animal species [3–6]. In this sense, it has been able to reliably detect biomechanical modifications due to neurological deficits, such as Parkinson's [7], Hemiparesis [8] or even pain [3], in humans.

The main ICOP pathway characteristics that have been reported as useful are: 1) craniocaudal COP excursion (measured as an initial and final COP relative coordinates) [8]; 2) lateromedial displacement of the ICOP by means of the center of pressure excursion index (CPEI),

Canaria, Arucas, Las Palmas, Spain
Full list of author information is available at the end of the article



© The Author(s). 2019 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons Rublic Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated

^{*} Correspondence: jose.vilar@ulpgces ¹Instituto Universitario de Investigaciones Biomédicas y Sanitarias,

^{&#}x27;Instituto Universitario de Investigaciones Biomedicas y Sanitarias, Universidad de las Palmas de Gran Canaria, Arucas, Las Palmas, Spain "Departamento de Patología Animal, Universidad de las Palmas de Gran Canaria, Arucas, Las Palmas, Spain

López et al. BMC Veterinary Research (2019) 15:138

Page 2 of 6

which represents the ICOP path lateromedial excursion relative to limb width and multiplied by 100 to obtain this data in terms of percentage [3, 9].

The COP path can be also obtained in a standing position and records its resultant area during a determinate period of time. This parameter is named statokinesiogram, and its value shows body or limb balance [10].

In the veterinary field, previously published studies only examine the bCOP path [11–13]; more recently, the bCOP path's efficacy for the detection of lameness in ponies at walk has been settled [14]. In dogs, bCOP modifications in unilaterally lame animals with elbow dysplasia (ED) have also been reported [15].

Regarding ED, this is a complex syndrome, where different factors could lead to a growth incongruence between the radius and ulna. Over time, ED causes joint damage, pain, and lameness [16, 17].

The hypothesis of this study was to prove that certain ICOP path characteristics are different in lame and sound limbs in dogs at walk and while standing still. For this reason, the aim of this study was to set a number of ICOP pathways –derived data that could serve to detect lameness in dogs with unilateral ED.

Methods

Animals

This study utilized 10 client-owned, adult dogs with similar conformation (2 rottweiler, 3 labrador retriever, 1 golden retriever, 2 german shepherd, 2 belgian shepherd). The body weight of the enrolled dogs ranged from 30 to 41,8 kg, and the ages were from 3 to 9 years.

Inclusion criteria comprised of the presence of weight-bearing unilateral forelimb lameness due to OA secondary to elbow dysplasia. The lameness of every dogs reached a score of 3–4 in a scale of 0–5 [18].

Furthermore, no medication could have been administered 1 month prior to the analysis.

To confirm or rule out OA, three standard radiographic views of both elbow joints (a lateral extension, lateral flexion, and a 15° oblique craniomedial caudolateral) [19] were taken under sedation with dexmedetomidine $10\pm20\,\mu\text{g/kg}$ (Dexdomitor, zoetis, Spain). Standard radiographs of stifle and hip joints were also taken in order to exclude other reasons for the observed clinical signs.

A complete clinical evaluation (physical examination, including vital signs and neurologic and orthopedic exams) assured that general health was otherwise normal.

Pressure platform study

A Pressure platform (EPS/R1, Loran Engineering, Bologne, Italy) was used for this study. This device contains a total of 2096 pressure sensors of 1 cm2

distributed in an area of $48\times48\,\text{cm}.$ The range of pressure was set from 30 to $400\,\text{kPa}.$

The procedure for the dynamic and static pressure platform analysis has been previously published [15, 20]; briefly, dogs were leash guided by their owners over the pressure platform at a walk (velocity 1.2 ± 0.2 m/s; acceleration ± 0.2 m/s2). Velocity and acceleration were measured with a motion sensor (PS-2103A, Pasco*, California, USA) placed within the dogs trajectory. Three trials were recorded at a sampling frequency of 100 Hz from each dog. A trial was considered valid when the studied limb fully supported over the pressure platform and when the dog walked next to the owner without pulling on the leash and without head turns. The pressure platform was interfaced with a dedicated computer using Biomech® (Loran Engineering, Bologna, Italy) software. Once the images were isolated, the paws' length was normalized to a fixed value of 9 cm, and width was then proportionally modified. Measurements were taken with a reference to an X-Y coordinate system.

Statokinesiograms were obtained while the dogs were placed in a quiet stance with their thoracic limbs over the pressure platform, perpendicular to the ground. The dog's owner remained in front of the animal to attract the dog's attention at a close distance. Three trials of 20-s recordings were obtained from each animal. A trial was considered valid when the animal remained with immobile limbs, tail and head during the whole 20 s recording procedure.

The following were the obtained measurements (Fig. 1):

- Caudal margin (Cm): defined as the distance between the most caudal limit of the paw print and the most caudal limit of the ICOP path.
- ICOP pathway length (e): the length of the line that joins the recorded points of the ICOP trajectory. Measured in cm.
- Craniocaudal index (CrCI): determines the COP length (b) related to the paw length (a). This is obtained with the following formula: % = (b / a) × 100. Expressed as a percentage.
- Center of the pressure excursion index (CPEI): determines the lateromedial excursion of the COP (c) related to the paw width (d). The formula was the following: % = (c / d) × 100. Expressed as a percentage.

Higher values of all the above parameters are associated with better limb support [3, 8, 9].

 statokinesiograms: defined as the area determined by an ellipse that contains 90% of the recorded points of the COP trajectory [10]. Measured in mm2, a lower value means more stability [15, 21]. López et al. BMC Veterinary Research (2019) 15:138

Page 3 of 6

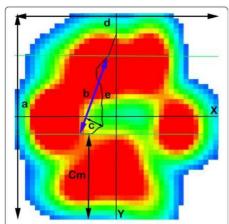


Fig. 1 Paw podobarographic print with coordinate system and measurements made. X: X coordinate; Y: Coordinate; a: paw length; b: COP length; c: ICOP width; d: paw width; e: COP path length; Cm: caudal margin

Force platform analysis

A force platform (Pasco, California, USA) was placed adjacent to the pressure platform in such a way that recordings from animals were performed in the same session. DataStudio software (Pasco, California, USA) was used to obtain PVF (N) values from three valid trials. Mean values were normalized to body weight (%BW).

Statistical analysis

For the data analysis, a linear mixed effects model was considered: for each response variable (COP Length, CPEI, etc), the status of the limb (lame/sound) is a fixed effects factor, while the dog is a random effects factor.

The model is as follows:

$$\begin{array}{ll} y_{ij}k = \mu_{i} + b_{j} + \epsilon_{ij}k, i = 1,...,2 \ j = 1,...,10, \ k \\ = 1,...,3 \end{array}$$

$$b_i \approx N(0,\sigma_b \;) \quad \epsilon_ijk \approx N(0,\sigma)$$

where:

- 1. y_i is the k-th measure (k = 1,2,3) on the limb i (i = sound/lame) of the dog j (j = 1...10)
- µ_i is the (fixed) effect of limb status i. This
 parameter represents the mean value of the variable
 in the sound (lame) limb.
- 3. b_j is the (random) effect of dog j. Values of b_j are supposed to be normally distributed with mean 0

- and standard deviation σ_b , so σ_b is the variability in the response of the dogs.
- 4. ϵ_{-} ijk is the residual in the measure ijk. This variable is assumed to be normally distributed with the mean 0 and standard deviation σ .

Statistical analysis was performed with 'R' statistical language and environment, version 3.3.2. (https://www.R-project.org/). For assessing the validity of the model, a Shapiro-Wilk test is applied to test the normality of the residuals, and a Levene test is used to test homoscedasticity.

Results

Mean weight (\pm SD) was 37.08 \pm 3.76 kg, and age was 5.80 \pm 1.99 years. The mean (\pm SD) values and 95% CI of all obtained parameters are shown in Table 1. All data were normally distributed and homoscedastic ($p \ge 0.25$ and $p \ge 0.12$, respectively).

Significant differences between LL and CL were found in all cases (<0.0001); concretely, a higher value of Cm and a lower COP Length, COP Path Length, and CrCI values in LL were observed when compared with CL. In the same manner, CPEI in LL were also lower than CL (Fig. 2, Additional file 1).

In agreement with the data shown above, PVF and VI values also showed significant differences between LL and CL ($p \le 0.0001$) (Table 1). PVF and VI data were also normally distributed and homoscedastic ($p \ge 0.64$ and $p \ge 0.51$, respectively).

Table 1 Mean ± SD, 95% confidence interval and difference between LL and CLs for CM, Cop Path Length, CrCl, PVF, VI and stately leading to the company of means significant difference.

	LL	CL	Difference
Cm (%)	44.85 ± 5.12	31.13 ± 7.61	a13.72 ± 0.94
	40.86, 48.84	27.14, 35.12	11.83, 15.61
COP Path Length (%)	42.00 ± 4.94	55.68 ± 9.92	a 13.69 \pm 0.97
	37.05, 46.95	50.73, 60.63	11.74, 15.63
CrCl (%)	31.07 ± 4.49	44.01 ± 6.75	a12.94 ± 1.23
	28.08, 34.06	41.02, 47.01	10.47, 15.42
CPEI (%)	4.57 ± 1.65	9.30 ± 1.78	^a 4.73 ± 0.35
	3.65, 5.49	8.38, 10.22	4.02, 5.44
PVF (%)	32.72 ± 4.66	71.12 ± 3.57	^a 38.40 ± 0.78
	31.13, 34.31	69.53, 72.71	36.84, 39.96
VI (%)	13.49 ± 1.32	22.93 ± 1.58	a 9.44 \pm 0.24
	12.80, 14.18	22.24, 13.62	8.96, 9.92
Statokinesiogram (mm²)	16.18 ± 6.10	5.70 ± 3.43	a10.48 ± 0.75
	13.16, 19.19	2.68, 8.71	8.98, 11.98

Cm Caudal margin, CrCl Craniocaudal index, CPEl Center of pressure excursion index

López et al. BMC Veterinary Research (2019) 15:138 Page 4 of 6

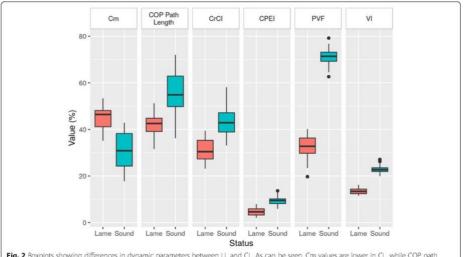


Fig. 2 Boxplots showing differences in dynamic parameters between LL and CL. As can be seen, Cm values are lower in CL, while COP path Length, CrCl and CPEI indexes are higher when compared with LL. This also occurs in PVF and VI values

Finally, the area from the statokinesiograms showed a higher value in LL (Fig. 3, Additional file 2). Additionally, a craniomedial COP slope was observed in both LL and CL when COP length was measured (Fig. 1, blue arrow).

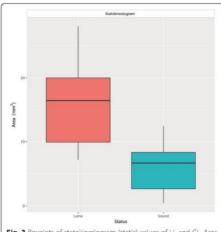


Fig. 3 Boxplots of statokinesiogram (static) values of LL and CL. Area of LL is higher than LL i.e., more instable

Discussion

Our results provide a novel insight into the adaptive changes in ICOP characteristics in unilaterally lame dogs

To the best of our knowledge, no other previous studies exist regarding the clinical implications of dynamic and static ICOP path characteristics in lame dogs.

Limb weight load amount could be influenced by the gait speed or cadence and, consequently, could alter COP path patterns [22]. Acknowledging this possibility, we performed the study in a narrow range of velocity and acceleration and tried to enroll similarly sized animals in order to minimize severe cadence discrepancies.

Once the data were obtained, we assumed that measurements on caudocranial and mediolateral COP displacement would provide four basic differences between LL and CL regarding:

1) The extent of net forward lCOP path progression. Based in our results, ICOP path in LL is shortened and cranialized compared with CL. This is in concordance with other authors' findings [8]. As made evident by the data, a larger Cm directly implies a shorter COP path length. This is invariably due to a shortened swing phase by a lack of limb extension, meaning the limb lands more vertically at the start of the braking phase [23]. This event prevents the metacarpal pad to exert a correct load absorption, expanding with the

López et al. BMC Veterinary Research (2019) 15:138 Page 5 of 6

- increase of weight-bearing when the limb lands [24, 25]. The impact shock could be, in the last instance, potentially transferred to muscles higher up the limb [5].
- 2) Net mediolateral lCOP deviation. As reported in previous research [26], a higher CPEI in CL is determined by an increased pad deformation, given that pad expansion is a direct response to weight loading. This effect has also been observed in human feet [9] and equine hooves [27].
- 3) Statokinesiograms. A greater area determines more instability [15]. This finding, although previously in reference to the body, remains true for limbs as well, since the area was greater in LL.
- 4) The lCOP direction of progression in both sound and lame limbs. As stated above, ICOP path described a certain angle (slope) as it pursued craniomedially with respect to the longitudinal axe of the paw. A possible explanation for this finding may be that the ICOP path follows the direction of the body's center of mass and not the craniocaudal paw axe, which corresponds to other reports in humans [28].

Another interesting finding was that the ICOP caudocranial displacement is constant during the support phase, but velocity is not (Additional file 1), which coincides with reports in human research regarding sound limbs [8]. In the present study, this characteristic was evident not only in CL but also in LL.

In humans, longitudinal COP displacement corresponds to 83% of foot length and 18% of foot width [28]; their equivalent values in CL of our study with dogs were about 44% (CrCI) and 9% (CPEI), respectively, which is approximately half. Two facets could explain these differences: 1- that humans have plantigrad support, which starts in the calcaneus bone, whereas in dogs the support is digitigrade; 2- human bipedalism determines full load transfer to the support limb when walking, whereas dogs walk with two (or even three) limbs simultaneously sharing the load support.

The following are some limitations in our study:

- 1. The ICOP path patterns in sound limbs cannot be extrapolated to limbs from sound dogs. As in lame dogs, sound limb patterns are showing compensatory movements. For the same reason, data from unilaterally lame limbs should not be extrapolated to bilateral lameness.
- 2. Compensatory weight redistribution in lame dogs not only implies to the contralateral limb, as has been well established in dogs and horses [29, 30]; thus, it would be useful to obtain hind limb ICOP path values in a subsequent study. Moreover, it

- should be determined if any correlations exist between the ICOP path values with the lameness degree or lameness origin. Unfortunately, the relatively large dog sizes impede the simultaneous analysis of more than two limbs, and a larger platform pressure mat would be essential.
- 3. Parameters, such as Cm and CPEI, need to be qualitative and not quantitatively considered, given that cut-points were not defined in our study, although significant differences were found in our study between CL and LL. To establish an accurate limit value for soundness or lameness, a higher number of patients with the same characteristics (weight, conformation, or even breed) are necessary, as reported by others authors in similar human studies [4].
- 4. Finally, the number of ICOP characteristics assessed could represent a "signature" diagnosis of ED, where the kinetic parameters to detect it have been previously proven [23]. This also means that COP patterns in other musculoskeletal and neurodegenerative disorders could be quite different, which needs further investigation.

Conclusion

This study showed that the ICOP path in LL is shorter, cranialized, and with smaller mediolateral excursion when compared with SL in dogs with unilateral ED. In addition, the ICOP path follows a craniomedial direction and not the paw longitudinal axe in both LL and CL. Its progression velocity is not constant.

Additional files

Additional file 1: Video S1. Limb and body statokinesiograms from a dog with a left limb lameness. As can be seen, the area of ellipse (18.28 mm² Vs 8, 33mm²) in the left (red) LL is greater than the right (blue) CL In the center (green) the body statokinesiogram can also be seen. (MP4

Additional file 2: Video S2. Simultaneous videosequence of suppo phase in a CL (left) and LL (right). The ICOP (black point) path in LL starts more cranially and therefore shortened. (MP4 650 kb)

Abbreviations
bCOP: Body Center of Pressure; CL: Sound limb; Cm: Caudal margin;
COP: Center of pressure; CPE: Center of pressure excursion index; CrCl: Craniocaudal index; ED: Elbow dysplasia; ICOP: Limb Center of Pressure.; LL: Lame limb; PVF: Peak vertical force; VI: Vertical impulse

Acknowledgments

The authors thank Amanda Hand for translation and editing. We would also like to thank the dog's owners for their collaboration. Thanks also to the Câtedra García Cugat for its technical support.

Fundina

No funds were received for this research.

López et al. BMC Veterinary Research

(2019) 15:138

Page 6 of 6

All data supporting our findings are included in the manuscript, if readers need additional information and/or data sets, they will be provided by the corresponding author upon reasonable request.

Authors' contributions

JMV, JS, and JMC conceived and designed the experiments; MR, ED and DC performed the clinical and radiological analyses, JMV and SL performed the pressure platform analysis; AS analyzed the data; all authors read and approved the final manuscript.

Authors' information

Ethics approval and consent to participate
The research protocol was revised and authorized by the Ethical Committee of Animal Welfare at the Instituto Universitario de Investigaciones Biomédicas y Sanitarias of the Universidad de Las Palmas de Gran Canaria (UIBS 14/2017) in compliance with the Directive 2010/63/EU of the European Union. Dog owners were informed of the study and signed a consent to participate in the study, including all performed procedures

Consent for publication

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

'Instituto Universitario de Investigaciones Biomédicas y Sanitarias, Universidad de las Palmas de Gran Canaria, Arucas, Las Palmas, Spain ²Departamento Medicina y Cirugía Animal, Cátedra García Cugat, Universidad Cardenal Herrera-CEU, CEU Universities, Valencia, Spain. ²Departamento de Matemáticas, Universidad de las Palmas de Gran Canaría, Las Palmas, Spain. ⁴Departamento de Patología Animal, Universidad de las Palmas de Gran Canaria, Arucas, Las Palmas, Spain

Received: 18 July 2018 Accepted: 24 April 2019 Published online: 08 May 2019

- References

 1. Kotti M, Duffell LD, Faisal AA, McGregor AH. Detecting knee osteoarthritis and its discriminating parameters using random forests. Med Eng Phys. 2017:43:19-29
- 2017/43:19–29.
 Alexander U, Campell KR. Dynamic assessment of foot mechanics as an adjunct to orthotic prescription. In: Wolf SL, editor. The biomechanics of the foot and ankle. Ist ed. Philadelphia: FA Davis, 1990. p. 148–52.
 Riskowski JL, Doffour AB, Hapedom TJ, Hillstorn HJ, Casey VA, Hannan MT.
 Associations of foot posture and function to lower extremity pain: results
- from a population-based foot study. Arthritis Care Res. 2013;65:1804-12.
- Grundy M, Tosh PA, McLeish RD, Smidt L. An investigation of the centers of pressure under the foot while walking. J Bone Joint Surg (B). 1975;57:98–103. Nauwelaerts S, Hobbs SJ, Back W. A horse's locomotor signature: COP path
- determined by the individual limb. PLoS One. 2017;12(2):e0167477.
 LeBlanc C, Tobakke B, Szkotnick B. Harlander-Matauschek a. locomotor behavior of chickens anticipating incline walking. Front Vet Sci. 2017;4:233.
 Kim DY, Hwang SH, Kim MK, Song JH, Lee SW, Kim IK. Development of
- Parkinson patient generated data collection platform using FHIR and IoT devices. Stud Health Technol Inform. 2017;245:141–5.

 Robain G, Valentini F, Renard-Deniel S, Chennevelle JM, Piera JB. A baropodometric parameter to analyze the gait of hemiparetic patients: th path of center of pressure. Ann Readapt Med Phys. 2006;49:609–13. Yoon HK, Park KB, Roh JY, Park HW, Chi HJ, Kim HW. Extraarticular subtala
- arthrodesis for pes planovalous; an interim result of 50 feet in patients with spastic diplegia. Clin Orthop Surg. 2010;2:13-21.

- Scoppa F, Capra R, Gallamini M, Shiffer R. Clinical stabilometry standardization: basic definitions—acquisition interval—sampling frequency. Gait Posture, 2013;37:290-2.
- Nauwelaerts S, Malone SR, Clayton HM. Development of postural balance in foals. Net J. 2013;198(Suppl 1):e70-4.
 Clayton HM, Nauwelaerts S. Effect of blindfolding on Centre of pressure
- variables in healthy horses during quiet standing. Vet J. 2014;199:365–9.

 Clayton HM, Buchholz R, Nauwelaerts S. Relationship between morphological and stabilographic variables in standing horses. Vet J. 2013; 198(Suppl 1):e65-9.
- Pitti L, Oosterlinck M, Diaz-Bertrana ML, Carrillo JM, Rubio M, Sopena J, Santana A, Vilar JM. Assessment of static posturography and pedobarography for the detection of unilateral forelimb lameness in ponies. BMC Vet Res. 2018;14:151.
- Manera ME, Carrillo JM, Batista M, Rubio M, Sopena J, Santana A, Vilar JM. Static Posturography: a new perspective in the assessment of lameness in a
- canine model. PLoS One. 2017;12(1):e0170692.
 Ternwichitr J, Leegwater PA, Hazewinkel HA. Fragmented coronoid process in the dog: a heritable disease. Vet J. 2010;185:123–9.
- Cook CR, Cook JL. Diagnostic imaging of canine elbow dysplasia: a review.
- Vet Surg. 2009;38:144–53. Scott H, Witte P. Investigation of lameness in dogs 1. Forelimb. Practice. 2011;33:20-7.
- Villamonte-Chevalier v BH, Broeckx B, Dingemanse W, Soler M, Van Ryssen B, Gielen I. Assessment of medial coronoid disease in 180 canine lame elbow joints: a sensitivity and specificity comparison of radiographic, computed tomographic and arthroscopic findings. BMC Vet Res. 2015;11243.
- Carrillo JM, Manera ME, Rubio M, Sopena J, Santana A, Vilar JM.

 Posturography and dynamic pedobarography in lame dogs with elbow dysplasia and cranial cruciate ligament rupture. BMC Vet Res. 2018;14:108.
- oppulsas and camer rousite inganient ropidie, owe ver res. 2016;4-106. https://doi.org/10.1186/s1297-018-1435-y. Asseman F, Caron O, Gremieux J. Is there a transfer of postural ability from specific to unspecific postures in elite gymnasts? Neurosci Lett. 2004;358:83-6. Kemozek TW, LaWott EE, Dancisak MJ. Reliability of an in-shee pressure
- measurements system during treadmill walking. Foot Ankle Int. 1996;17:204–9.
 Galindo-Zamora V, Dziallas P, Wolf DC, Kramer S, Abdelhadi J, Lucas K, Nolte I, Wefstaedt P. Evaluation of thoracic limb loads, elbow movement, and morphology in dogs before and after arthroscopic management of unilateral medial coronoid process disease. Vet Surg. 2014;43:819–28.

 Basher A. Foot injuries in dogs and cats. Compend Cont Edu Pract vet 1b;
- 1994, p. 1159-76.
- Swaim SF, Management and bandaging of soft tissue injuries of dog and cat feet. J Am Anim Hosp Assoc. 1985;21:329–40.

 Marghitu DB, Swaim SF, Rumph PF, Cojonaru D, Gillette RL, Scardino MS. Dynamics Analysis of Ground Contact Pressure of English Pointer Dogs.
 Nonlinear Dyn. 2003;33:253–65.
 Johnston C, Back W. Hoof ground interaction: when biomechanical stimuli
- challenge the tissues of the distal limb. Equine Vet J. 2006;38:634-41. Han TR, Palk NJ, Irn MS. Quantification of the path of center of pressure (COP) using an F-scan in-shoe transducer. Gait Posture. 1999;10:248-54. Molsa SH, Hyytlainen HK, Hielm-Bjorkman AK, Laitinen-Vapaavuori OM.
- Long-term functional outcome after surgical repair of cranial cruciate ligament disease in dogs. BMC Vet Res. 2014;10:266.

 Oosterlinck M, Pille F, Back W, Dewulf J, Gasthuys F. A pressure plate study
- on fore and hindlimb loading and the association with hoof contact area in sound ponies at the walk and trot. Vet J. 2011;190:71–6.

Ready to submit your research? Choose BMC and benefit from:

- thorough peer review by experienced researchers in your field
- cation on acceptance · rapid put
- support for research data, including large and complex data types
- oration and increased citati · gold Open Access which fosters wider co m visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more blomedcentral.com/submis



TRATAMIENTO OA Y ARTÍCULOS

TRATAMIENTO OA Y ARTÍCULOS



4. TRATAMIENTO DE LA OA

4.1. Tratamientos convencionales

La estrategia terapéutica ha tratado, de forma clásica, de frenar o al menos ralentizar el proceso inflamatorio que en definitiva determina la aparición del dolor y la alteración funcional.

En primer lugar, debemos destacar aquellas medidas higiénicas y dietéticas, como la reducción de peso, el ejercicio de bajo impacto, así como la fisioterapia, láser de baja intensidad, acupuntura y ultrasonidos entre otros (Gudbergsen y cols, 2012; Cakir y cols, 2013; Fang y cols, 2013).

Desde el punto de vista del tratamiento basado en principios activos, los antiinflamatorios no esteroideos, así como los esteroideos de larga duración han formado de manera amplia el elenco de medicamentos clásicos a usar en estas situaciones. También los complementos nutricionales como los glucosaminoglicanos de origen diverso han demostrado su utilidad (Merashly y Uthman, 2012; Reid y cols, 2012; Godley y cols, 2013).

La evolución de los tratamientos clásicos, fundamentalmente los analgésicos no esteroideos han ido encaminando a conseguir moléculas con menos efectos colaterales, así como de mayor comodidad de administración. En este sentido cabe resaltar el uso de un inhibidor COX-2, el Mavacoxib (TrocoxilTM, Pfizer, MY, USA) (Penning y cols 1997). Este compuesto ha sufrido una serie de modificaciones bioquímicas que han conseguido hacerlo muy estable, y por lo tanto una vida media muy alta (Cox y cols, 2010)



Con eso se consigue por un lado mucha comodidad en su administración (una vez al mes), y por otro debido a su bajo índice de aclaramiento, que la dosis sea muy baja para obtener un óptimo efecto terapéutico. (Paulson y cols, 2001).

De modo general, debemos decir que las terapias anteriormente descritas solo obtienen un efecto sintomático, pero no van a impedir la evolución de la OA (Lawrence y cols, 2008; Cuervo y cols, 2016). Por este motivo, las terapias regenerativas están enfocadas en la aportación de células con capacidad de generar cartílago, así como también factores que promuevan la reparación tisular (Williams y cols, 2012).

4.2. Terapias regenerativas

Como decíamos anteriormente, estas terapias suponen una revolución terapéutica pues se adquiere una nueva perspectiva de tratamiento al buscar no solo la mejoría sintomática, sino que además aporta y estimula los factores y procesos presentes en el propio organismo de modo que, como mínimo, frenen o ralenticen el efecto degenerativo sin efectos nocivos al ser sustancias o células autólogas. (Wu y cols, 2007; Singh, 2012).

4.2.1 Células madre.

Las células mesenquimales o, más concretamente las células mesenquimales adultas son capaces de diferenciarse en las diversas variantes de tejido conectivo (adiposo, óseo, cartilaginoso) (Fortier y Travis, 2011; Diekman y Guilak, 2013). Estos dos últimos adquieren gran importancia desde el punto de vista clínico al formar parte de los componentes articulares.



Estas células mesenquimales se extraen principalmente del tejido adiposo (Black y cols, 2008; Yarak y Okamoto, 2010), aunque también se pueden obtener de medula ósea y/o hueso. Las células mesenquimales, y en concreto su eficacia en la OA está siendo objeto de múltiples estudios en la actualidad.

TRATAMIENTO OA Y ARTÍCULOS



Damia, E., Chicharro, D., Lopez, S., Cuervo, B., Rubio, M., Sopena, J.J., Vilar, J.M., Carrillo, J.M. <u>Adipose-derived mesenchymal stem cells:</u>

<u>Are they a good therapeutic strategy for osteoarthritis?</u>

Int J Mol Sci. 2018 Jun 30;19(7). doi: 10.3390/ijms19071926.

JCR Impact factor (2017): 3.687

Cuartil: Q2

Grupo: Chemistry, multidisciplinary

Posición: 52/171

TRATAMIENTO OA Y ARTÍCULOS





Revieu

Adipose-Derived Mesenchymal Stem Cells: Are They a Good Therapeutic Strategy for Osteoarthritis?

Elena Damia ^{1,2}, Deborah Chicharro ^{1,2}, Sergio Lopez ³, Belen Cuervo ^{1,2}, Monica Rubio ^{1,2}, Joaquin J. Sopena ^{1,2}, Jose Manuel Vilar ^{2,3}, ⁴ and Jose Maria Carrillo ^{1,2}

- Bioregenerative Medicine and Applied Surgery Research Group, Department of Animal Medicine and Surgery, CEU Cardenal Herrera University, CEU Universities, C/Tirant lo Blanc, 7, Alfara del Patriarca, 46115 Valencia, Spain; elena.damia@uchceu.es (E.D.); debora.chicharro@uchceu.es (D.C.); belen.cuervo@uchceu.es (B.C.); mrubio@uchceu.es (M.R.); jsopena@uchceu.es (J.J.S.); jcarrill@uchceu.es (J.M.C.)
- Garcia Cugat Foundation CEU UCH Chair of Medicine and Regenerative Surgery, 08006 Barcelona, Spain
- Department of Animal Pathology, Instituto Universitario de Investigaciones Biomédicas y Sanitarias, University of Las Palmas de Gran Canaria, 35416 Las Palmas de Gran Canaria, Spain; sergiolopezbarbeta@gmail.com
- * Correspondence: jose.vilar@ulpgc.es; Tel.: +34-928-457-244

Received: 28 May 2018; Accepted: 28 June 2018; Published: 30 June 2018



Abstract: Osteoarthritis (OA) is a major cause of disability in elderly population around the world. More than one-third of people over 65 years old shows either clinical or radiological evidence of OA. There is no effective treatment for this degenerative disease, due to the limited capacity for spontaneous cartilage regeneration. Regarding the use of regenerative therapies, it has been reported that one option to restore degenerated cartilage are adipose-derived mesenchymal stem cells (ASCs). The purpose of this review is to describe and compare the efficacy of ASCs versus other therapies in OA. Methods: Recent studies have shown that ASCs exert paracrine effects protecting against degenerative changes in chondrocytes. According to the above, we have carried out a review of the literature using a combination of osteoarthritis, stem cells, and regenerative therapies as keywords. Results: Conventional pharmacological therapies for OA treatment are considered before the surgical option, however, they do not stop the progression of the disease. Moreover, total joint replacement is not recommended for patients under 55 years, and high tibia osteotomy (HTO) is a viable solution to address lower limb malalignment with concomitant OA, but some complications have been described. In recent years, the use of mesenchymal stem cells (MSCs) as a treatment strategy for OA is increasing considerably, thanks to their capacity to improve symptoms together with joint functionality and, therefore, the patients' quality of life. Conclusions: ASC therapy has a positive effect on patients with OA, although there is limited evidence and little long-term follow-up.

Keywords: osteoarthritis; mesenchymal stem cells; regenerative medicine

1. Introduction

Osteoarthritis (OA) is a progressive degenerative disease of the joint characterized by gradual degradation of hyaline articular cartilage and sclerosis of bone. This cartilage is composed by type II collagen and proteoglycans. An alteration in the replacement of the proteoglycan and type II collagen network leads to the loss of function of the cartilage [1]. This disease, worldwide, is considered to be the fourth leading cause of disability [2] and the second cause of inability to work in men [3]. OA is the most common articular disease in adults, and knee OA is the most common location. Although, OA also affects other large-weight-bearing joints, such as hip, hands, feet, and spine [4]. Hip and knee

Int. J. Mol. Sci. 2018, 19, 1926; doi:10.3390/ijms19071926

www.mdpi.com/journal/ijms

Int. J. Mol. Sci. 2018, 19, 1926 2 of 14

OA are leading causes of disability worldwide [5]. The disease is characterized, at first, by a molecular derangement (alteration of joint tissue metabolism) followed by physiologic/anatomic damages (cartilage degradation, bone remodeling, osteophyte formation, joint inflammation), culminating in a loss of normal joint function [6].

In the United States, 27 millions of people suffer from clinical OA, and the treatment costs 185.5 billion dollars per year [7]. On the other hand, this pathology is the fourth leading cause of disability in Asia [2]. In addition, this chronic degenerative disease of articular cartilage has a current prevalence of 12% in the population over 60 years old, which will increase in the next 20 years [8,9]. It has also been reported that its incidence has doubled in women, and tripled in men, in recent years [10].

Several agents have been associated with a higher risk of suffering OA, such as genetic predisposition, obesity, previous trauma, and age. It has been demonstrated that the risk of developing post-traumatic OA increases by up to four times in people over the age of 50 [11]. As one ages, the chondrocytes, that contribute to 5% of the volume of the articular cartilage, decrease their regenerative response, leading to progressive loss of the articular surface resulting in cartilage degeneration with loss of matrix (which confers the biomechanical properties to the articular cartilage and constitutes the 95% of the tissue), which can result in a complete loss of joint surface. Moreover, chondrocytes produce mediators of inflammation (cytokines, chemokines, and proteolytic enzymes) that induce serious damage [12].

Pain is one of the first symptoms, leading to movement disability and impaired quality of life [9,13]. Synovial inflammation, cartilage breakdown, and bone remodeling are associated with OA chronic pain, and the mechanism responsible for pain involves structural changes and alterations in peripheral transduction and central processing of painful sensory inputs [14]. Consequently, ideal treatment should obtain analgesia, stopping progression of chondral degeneration; modify cartilage structure and revert damage; and finally, improve joint function [15].

While conventional therapies, such as physical therapy, glucosamine and chondroitin sulfate supplementation, arthroscopic surgery, or biological therapies, such as chondrocyte implantation, have little significant results, regenerative medicine (RM), has been demonstrated to be a great option in articular cartilage regeneration [16]. On the other hand, various surgical procedures have been performed to regenerate articular cartilage but have achieved limited success, including abrasion arthroplasty, subchondral drilling, and microfracture [17]. Mesenchymal stem cells (MSCs) are considered to be a promising candidate for cartilage regeneration, due to their ability to differentiate towards cartilage and bone cells and secrete trophic factors with regenerative functions [18]. The paracrine effect and anti-apoptotic, anti-inflammatory and anti-aging functions of these stem cells, is fundamental for the regeneration process. Recently, an anti-aging effect of the conditioned medium of adipose-derived mesenchymal stem cells (ASCs) on OA chondrocytes has been reported, featured by downregulation of senescence markers induced by inflammatory stress [19]. Stem cells promote biological processes, such as vascularization, cell proliferation, differentiation, and modulation of the inflammatory process [20], and can be isolated, among others, from bone marrow, adipose tissue, umbilical cord blood, and placenta [21,22]. It is currently admitted that there are MSCs within the connective tissue of virtually all organs [20]. In humans, ASCs showed a greater capacity for proliferation than the rest of the human MSCs [23], moreover, these cells maintain the differentiation potential after a longer time of culture [24], and the age of donors has less effect on the proliferation of them; this is important in elderly patients with osteoporosis [25].

ASCs were first identified in the early 2000s, and demonstrated to have self-renewal ability and multilineage differentiation potential [26]. These cells have a several benefits: faster and easier expansion in culture, more passage cells that retain stem cell phenotypes, pluripotency [27], less susceptibility to age, and less morbidity of patients [28], furthermore, compared with bone marrow-derived mesenchymal stem cells (BMMSCs), ASCs do have an equal potential to differentiate into cells and tissues of mesodermal origin, such as adipocytes, cartilage, bone, and skeletal muscle.

Int. J. Mol. Sci. 2018, 19, 1926 3 of 14

On the other hand, the easy and repeatable access to subcutaneous adipose tissue and the simple isolation procedures provide a clear advantage [29].

To establish the efficacy of treatment with this regenerative therapy and assess, in the case of OA, the quality and thickness of the cartilage, long-term patient controls are needed [30]. Different studies have shown that the application of MSCs as therapy in the treatment of OA has improved the symptoms suffered by patients, particularly after more than 6 months of follow-up [10,30,31].

Recent studies have focused on BMMSCs for chondrogenesis, but the clinical use of these cells has presented disadvantages, such as, donor site morbidity, pain and low cell number upon harvest [32]. On the other hand, ASCs are a positive alternative treatment for OA treatment as in vitro studies have proven they contain CD73, CD90, CD105, and CD106 markers, which are necessary for cell differentiation into cartilage, and moreover, in vivo studies have also reported good results [33]. For all of the abovementioned reasons, the aim of the present study is to review the application of MSCs in OA, with particular emphasis on the use of ASCs versus other therapies.

2. Results

The prevalence and incidence of OA have increased globally, but pharmaceutical or surgical therapies have limited efficacy in halting OA progression. Among conservative therapies, nonpharmacological treatment (physiotherapy, weight management), systemic pharmacological treatment (analgesics, nonsteroidal anti-inflammatory drugs, glucosamine, and chondroitin sulfate) and injections of intra-articular (IA) therapies are described. Local delivery of corticoids and hyaluronic acid (HA) are approved treatments by United States Food and Drug Administration (US FDA)/European Medicines Agency (EMA), however, some adverse effects have been described, such as inflammation or pain and septic arthritis at the site of injection [34].

When OA advances, some surgical treatments can rebuild the degenerated cartilage, but they do not stop the articular inflammatory process established. These treatments are arthroscopic debridement, microfracture/osteoplasty, and chondrocyte implantation techniques, such as autologous chondrocyte implantation and matrix-induced autologous chondrocyte implantation [35]. The latter is the only cell therapy approved by the FDA [36]. The application of such interventions remains limited, due to the necessity of additional surgery for harvesting the donor autograft cartilage, and the poor integration of the grafted defect with the surrounded cartilage [37].

Due to the limitations regarding OA conventional therapies, for example, the pharmacological therapy could produce serious gastrointestinal, renal, and cardiac adverse effects, and some of them can be a threat to life or they can leave a permanent disability. On the other hand, the surgical option, such as microfracture, has been used for the last 20 years, but hyaline cartilage has a limited capacity for regeneration. In recent years, the interest for new therapies, such as IA injection, autologous blood therapies, and MSCs is increasing considerably. Autologous conditioned serum (ACS) is a cell-free treatment obtained by incubating venous blood in a specialized syringe, where blood cells release anti-inflammatory cytokines and growth factors (GFs), such as transforming growth factor-\$\beta\$ (TGF-β) [38]. It has been demonstrate that ACS therapy is more effective than HA injection [39] and the treatment with ACS and physiotherapy reduce chronic pain in knee OA [40]. Plasma rich in growth factors (PRGFs), is a type of platelet rich plasma (PRP) preparation. It is an autologous product with a moderate concentration of platelets, multitude of GFs, and absence of leukocytes. It provides an anabolic effect on the resident cells, and due to its potential to inhibit inflammation, relieves OA symptoms [41]. When PRGF is injected IA, it provides a three-dimensional network in the joint composed of fibrin that contains binding sites for cell adhesion, as well as proteins that form the microenvironment leading to different adhesion molecules and cells that help biological cartilage repair [42]. Recently, it has been reported that PRGF injection is an effective option to decrease pain and improve function in patients with symptomatic mild to moderate knee OA, in the 6 month follow-up [43]. In recent years, the release of ASCs with PRP has been reported to improve the

Int. J. Mol. Sci. 2018, 19, 1926 4 of 14

proliferation and chondrogenesis of this type of stem cells [44,45], suggesting new applications in RM for the management of osteochondral defects.

Currently, RM, that aims to promote regenerative or reparative phenomena over the degenerative processes, is in full swing. Among these therapies are the already mentioned PRGF and the MSCs.

2.1. Mesenchymal Stem Cells

MSCs are defined as those cells that meet the criteria established by the International Society of Cellular Therapy. These criteria include an ability to adhere to plastic, the expression of a number of cell markers, including CD105, CD73, and CD90 while undergoing multilineage differentiation, and the ability to self-renew [46]. Those multipotent adult stem cells synthesize mediators (cytokines, neuroregulatory peptides, trophic factors) which participate in tissue repair and regulate inflammatory and immune responses [47]. These adult stem cells could be induced to differentiate exclusively into the adipocytic, chondrocytic, or osteocytic lineages. It has been identified that individual stem cells, when expanded to colonies, retained their multilineage potential [48].

MSCs can be obtained from various adult tissues, for example, bone marrow, umbilical cord, skeletal muscle, synovial capsule, and adipose tissue. This last origin, has a number of advantages over the others, because adipose tissue is abundant and easy to obtain, and can be obtained in large amounts, using local anesthesia and causing minimal discomfort [49]. Subcutaneous fat tissue is the most accessible source, but in recent studies, other sources for obtaining autologous ASCs as the supra- and infrapatellar fat pads have been described [50]. Bone marrow aspirate has a paucity of MSCs, comprising 0.001–0.02% of the mononucleated cell population, in comparison to ~1–7% of the mononucleated cell population within adipose tissue [51,52]. Moreover, adipose tissue is considered a primary source, because it contains 500 times more MSCs than the same volume of bone marrow [53]. Furthermore, bone marrow harvested from the iliac crest is painful, and increases the risk of infection [54]. Recently, some reports demonstrate that IA injection of allogenic ASCs combined with HA could stop OA progression and promote cartilage regeneration [55]. Additionally, a successful management of a post-traumatic chondral defect using IA autologous ASC therapy has been suggested [35].

The FDA regulates the use of adult stem cells. This agency adopted 21 CFR 1271, which modified its jurisdiction over human cells and tissues to include any "transfer into a human recipient". Previously, the code was specified transfer "into another human," excluding autologous cells. Since then, cells that are more than "minimally manipulated," even if they are intended for autologous use, and are subject to similar regulations as manufactured drugs [56]. More research studies on the use of MSCs in OA treatment would allow the FDA and physicians to provide patients with a more confidently alternative, minimally invasive treatment options that may significantly slow disease progression.

2.1.1. The Role of Mesenchymal Stem Cells in Osteoarthritis

The mechanism by which MSCs cause cartilage regeneration is not clear. It has been postulated that these cells may act on subchondral bone, forming the primary repair cartilage [57]. The injection of MSCs in joint cavity is a novel therapy that improves OA symptoms due to their ability to stimulate local repair and regeneration of damaged joint tissues, and to reduce inflammation and associated pain [16]. MSCs modulate the inflammatory response by causing the suppression of inflammatory T-cell proliferation and inhibition of monocyte and myeloid dendritic cell maturation [58]. Moreover, the anti-inflammatory capacity can be stimulated by various pro-inflammatory cytokines (IL-6, tumor necrosis factor and interferon gamma) [59]. Furthermore, these stem cells secrete reparative cytokines, including TGF- β , vascular endothelial growth factor (VEGF), and epidermal growth factor (EGF), which are responsible for a trophic effect that produces the local tissue repair [60]. Additional in vitro studies have demonstrated that GFs, such as TGF- β and insulin-like growth factor 1 (IGF-1), can stimulate MSCs towards chondrocytes. These chondrocytes, derived from MSCs, have the same

Int. J. Mol. Sci. 2018, 19, 1926 5 of 14

expression of type II collagen that mature adult chondrocytes, moreover, this type II collagen provides tensile strength in the joint [61].

It has been demonstrated that after IA injection of MSCs (Table 1), these cells were found in the synovial membrane and they expressed molecules with anti-inflammatory and chondrogenic properties. MSCs could help to establish a regenerative microenvironment at the site of release, which would improve the recruitment, activation, and differentiation of endogenous stem cells with the potential to repair the articular cartilage [62].

With the aim of achieving a prolonged regenerative activity in the OA joint, several studies propose using MSCs activated with biomolecules to potentially improve chondral and osteochondral lesion repair. The need to use bioactive scaffolds was proven by the fact that in the knee joint of experimental mice, only 15% of ASCs injected were detectable at 1 month post-injection, and this number decreased to 1.5% in 6 months [63]. This type of MSC co-delivery with scaffolds could have better retention, aggregation, and viability of these cells; moreover, the proliferation, migration, and chondrogenic differentiation improve in scaffolds with large pore size [64].

Some in vitro studies have demonstrated the beneficial effects of scaffolds (CDM, poly l-glutamic acid/chitosan, TGF- β 1-conjugated chitosan hydrogel), such as regeneration of hyaline cartilage, enhancing ASC chondrogenesis and reparation of full-thickness cartilage defects [65–67]. More scaffolds include fibrin, gelatin and collagen (protein-based scaffolds), and alginate or agarose, among others (carbohydrate-based scaffolds). Other scaffolds that maintain a three-dimensional structure include the hydrogel family and hydrophilic polymer [36].

Table 1. Research on osteoarthritis with MSC-based therapy.

Study	Model	MSCs Type	OA Location	Results		
Garay-Mendoza et al., 2018 [15]	Human	BMMSCs	Knee	Improvement in knee pain and quality or life since first evaluation until the last one at 6 months		
Sun et al., 2018 [68]	Rabbit	ASCs + TGF-β3 poly-lactic-co-glycolic Knee acid Microspheres		Promoted cartilage regeneration and lessened the severity of OA in vivo		
Desancé et al., 2018 [69]	Equine	UCBMSCs	In vitro	High proliferative capacity and differentiated into osteoblasts and chondrocytes. Have a great potential for cartilage tissue engineering		
Freitag et al., 2017 [35]	Human	Arthroscopy with removal of a chondral loose body + ASCs	Post-traumatic chondral defect of the patella	Complete regeneration of hyaline-like cartilage within the defect and improvement of the pain and function		
Abbas 2017 [70]	Human	BMMSCs + cartilage fragments	Osteochondral bone samples from patients with total knee arthroplasty and a central drill defect (human ex vivo osteochondral defect model)	Improvement in chondrogenic differentiation and positive staining for type II collagen antibodies		
Murphy et al., 2017 [71]	Human	BMMSCs	First Carpometacarpal joint	Functional and symptomatic relief for the patients		
Pers et al., 2016 [72]	Human	ASCs	Knee	Patients treated with ASCs experienced significant improvements in pain levels and function knee compared with baseline.		
Rich et al., 2015 [73]	Human	BMMSCs	Knee	Significantly improved the knee injury and Osteoarthritis Outcome Score and knee cartilage thickness (measured by magnetic resonance imaging), indicating that they may enhance the functional outcome as well as the structural component		

Int. J. Mol. Sci. 2018, 19, 1926 6 of 14

Table 1. Cont.

Study	Model	MSCs Type	OA Location	Results		
Jo et al., 2014 [74]	Human	ASCs	Knee	Improve function and pain of the knee joint without causing adverse events, and reduce cartilage defects by regeneration of hyaline-like articular cartilage		
Wu et al., 2014 [75]	Rat	SMSCs + fibrin/chitosan scaffold + TGF-β3	Temporomandibular Joint	Fibrocartilage formation with deposition of Col1 and Col2		
Chen et al., 2013 [76]	Rabbit	BMMSCs	Temporomandibular Joint	Enhance the regenerative process of cartilage repair at the early stage of Temporomandibular joint OA		

MSCs mesenchymal stem cells, BMMSCs bone marrow mesenchymal stem cells, OA osteoarthritis, TGF- β 3 transforming growth factor β 3, UCBMSCs umbilical cord blood mesenchymal stem cells, ASCs adipose-derived stromal cells, SMSCs synovial mesenchymal stem cells, Col1 and Col2 type 1 and type 2 collagen.

2.1.2. Mesenchymal Stem Cells Exosomes in Osteoarthritis

Exosomes are a type of secreted membrane vesicles produced by different cells. Some types of exosomes have been shown to confer immunosuppressive effects in different disease models, among others, rheumatoid arthritis [77]. MSC exosomes are accepted as the principal therapeutic agents present in MSC secretion, and are adequate to mediate the many reported therapeutic options of MSCs [78]. Zhang et al., (2016) have reported that human MSC exosomes promote cartilage regeneration in an immunocompetent rat osteochondral defect model, so MSC exosomes help to regenerate the damaged articular cartilage in OA. The mechanism of action of MSC exosomes in that study were accelerated neotissue filling and enhanced matrix synthesis of type II collagen and sulphated glycosaminoglycan [79].

Another study demonstrates that weekly IA injections of human embryonic MSC exosomes induced a regeneration cartilage and subchondral bone over a period of 12 weeks in an adult immunocompetent rat model [79]. Definitely, the efficacy of MSC-based therapies has been assigned to the paracrine secretion of trophic factors, and exosomes have a fundamental role in mediating tissue repair, thus, exosomes represent a novel therapeutic option for OA.

3. Discussion

ASCs have a series of advantages over other types of cells, because adipose tissue is abundant and easy to obtain. On the other hand, these cells have a high in vitro proliferation capacity and fibroblastic morphology, and they can adhere well to the culture plate. Furthermore, they have a low risk of rejection [80]. ASCs can be isolated from the stromal vascular fraction (SVF) of adipose tissue. The cells are obtained by liposuction, followed by collagenase digestion, centrifugation, and dilution. SVF includes ASCs, as well as other cells, including pericytes, vascular adventitia cells, fibroblasts, preadipocytes, monocytes, macrophages, and red blood cells. The SVF product has 500,000 to 2,000,000 cells per gram, of which 1 to 10% are considered ASCs [81].

It has been calculated that there are approximately $1\times10^{5-6}$ ASCs in 1 ml of lipoaspirate, while there are 50–675 BMMSCs in 1 mL of bone marrow aspirate [82]. Another alternative for OA treatment with MSCs are human umbilical cord-derived MSCs (hUC-MSCs). These cells enhance the proliferation of OA chondrocytes and downregulate the expression of inflammatory cytokines, moreover, the co-culture of hUC-MSCs and OA chondrocytes may to be a therapeutic option for OA [22]. hUC-MSCs could be an alternative to BMMSCs for clinical applications, due to their easy preparation and low risk of viral contamination. They can differentiate into the three germ layers that promote tissue and organ repair and modulate immune responses [83]. In this review, we have reported the different types of treatment that are used conventionally in joint degenerative disease. Our discussion focuses on comparing ASCs with others stem cells most commonly used in the OA treatment in recent years.

Int. I. Mol. Sci. 2018, 19, 1926 7 of 14

3.1. Adipose-Derived Mesenchymal Stem Cells

In human medicine, it has been shown that MSCs are a clinical promise for articular cartilage regeneration. Several authors reported studies that demonstrate the effectiveness of MSCs in OA treatment. In relation to ASCs, the first case report was published in 2001 [21]. During the last decade, these cells have attracted great interest because they have been demonstrated to be safe and efficient for articular cartilage regeneration in several trials. In recent years, IA injection of ASCs in knee OA showed clinical, radiological, arthroscopic, and histological evidence at 6-month follow-up [74]. Among other studies, the IA injection of these stem cells (isolated from abdominal subcutaneous fat tissue) in severe knee OA, reported that clinical outcomes (pain, function knee, return to sport) of the low- and medium-dose groups tended to deteriorate after 1 year, while those of the high-dose group tended to plateau after 1 year, until 2 years [10]. Recently, Spasovski et al. (2018) have demonstrated that the use of ASCs from subcutaneous fat in knee OA improves clinical symptoms and reduces pain at 3 months, obtaining the best results at 6 months [30]. ASC therapy in OA has shown chondrogenesis potential, both for the infrapatellar- and suprapatellar-derived ASCs [50,84]. A greater chondrogenesis potential has been reported by infrapatellar ASCs compared to suprapatellar in vitro and in vivo [84,85]. In addition, the suprapatellar-derived ASCs transplantation in a severe knee OA mouse model diminished inflammation and cartilage degenerative grade, increasing the synthesis of glycosaminoglycan and inducing endogenous chondrogenesis [50]. These effects may be due to ASCs-mediated reduction of pro-inflammatory cytokines and chemokines, apoptosis of chondrocytes, hypertrophic and fibrotic chondrocyte phenotypes, and collagenases [86].

One of the limitations of the studies that describe the use of ASCs in OA is the short follow-up period, Joe at al. (2014) and Pers et al. (2016) reported the efficacy of IA injections of these cells for the treatment of knee OA, but their follow-up period was only 24 weeks [72,74]. However, Song et al. (2018) have reported the first study that has demonstrated the efficacy of ASCs therapy in knee OA with long-term follow-up of 96 weeks with repeated injections. These patients showed improvement in pain, function, and cartilage volume of the knee joint with repeated IA injections of these cells [87].

3.2. Bone Marrow Mesenchymal Stem Cells

Pittenger et al. (1999) isolated MSCs from bone marrow adult cells and since then they have been used to treat chondral defects [48]. Some studies in patients with OA treated with BMMSCs obtained good results, reducing the symptoms and, therefore, increasing patient satisfaction. In a study carried out in 24 patients with knee OA infiltrated with BMMSCs, histological and arthroscopic improvement was observed [88]. Another report by Kuroda et al. (2002) concluded that the transplantation of autologous BMMSCs promotes the repair of large defects of focal articular cartilage in young and active patients [89]. Recently, safety of IA injection of BMMSCs was confirmed in 12 OA patients. They showed pain relief and improvement of cartilage quality at 2 years post-treatment [90]. Regarding the regulation of the inflammatory process in OA, Zhang et al. (2016) reported that the co-cultivation of BMMSCs with chondrocytes from patients with OA increases cell proliferation of chondrocytes and inhibits inflammatory activity in OA [91]. In a study of knee OA treatment in 13 patients with in vitro expanded BMMSCs at 12 months, a significant improvement in the thickness of knee cartilage in the femoral and tibial plates was shown [73]. Good results have also been reported with the application of BMMSCs in large [92] and small joints [71] with microfracture.

3.3. Human Umbilical Cord-Derived Mesenchymal Stem Cells

Some studies have demonstrated that chondrocytes secrete the same cytokines and induce human stem cells to differentiate into chondrocytes [93,94]. It has been reported that the hUC-MSCs improve the viability of OA-degenerated chondrocytes. A study carried out in Shangai suggested that the secretion of hUC-MSCs enhanced chondrocyte proliferation and showed that these stem cells increased expression of chondrogenic genes (aggrecan, sox-9, collagen II), indicating chondrocytes

Int. J. Mol. Sci. 2018, 19, 1926 8 of 14

promoted chondrogenic differentiation of hUC-MSCs, compared to the control group. They also postulated that hUC-MSCs inhibited inflammatory activity in OA chondrocytes [22]. In the same line, Zheng et al. (2013) reported the chondrogenic differentiation of hUC-MSCs by co-culture with rabbit chondrocytes [95]. In other studies, hUC-MSCs were reported to inhibit the expression of some inflammatory factors [96,97]. Definitely, hUC-MSCs could regulate inflammatory activity and the proliferation of chondrocytes in OA.

Therapeutic options of OA depend on each individual case and multiple factors, such as the disease progression, degeneration degree of articular cartilage, affected joint, and patient's expectations. The gold standard in the treatment of OA is total joint replacement, but surgery is not suitable for patients under 55 years [98], however, HTO is intended to transfer the mechanical axis from medial to slightly lateral, to the midline of the knee, to decrease the load and subsequently delay OA [99] in 40–60 years old patients [100]. Some studies showed that the regenerative process began after realignment [99], but different complications have been described, such as aseptic nonunion and deep infection [101]. MSCs may be a safe and alternative treatment strategy for OA, due to that this therapy has different advantages: it is applied in all joints, the injections are repeatable, and it is minimally invasive [12].

4. Materials and Methods

The authors searched PubMed English languages articles using a combination of "osteoarthritis", "stem cells", "regenerative therapies", and "adipose-derived mesenchymal stem cells" as keywords. After the first selection of the main articles based on OA and conventional treatments, studies of OA and stem cells were selected, and there were a total of about 900 articles in the last decade. Special attention has been drawn to original analyses and studies, around 300 studies, of which we chose about 80 articles dedicated to ASCs and were published in the last 10 years. Other searches were executed using bibliographies of articles found in the primary and secondary search. One limitation in this review, is the fact that our methods, while rigorous, did not follow any formal guidelines for a systematic review (e.g., the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines).

5. Conclusions

OA is a major health problem, especially in the elderly population. Cellular therapy is an emerging modality for the treatment of OA, even the combination of conventional treatments with the application of MSCs is a therapeutic option to improve the quality of life of patients with OA. In recent years, the interest in MSCs as a therapeutic option in OA is due to the facility of harvesting, preparation, and implantation without surgery, the capacity to stimulate local repair and regeneration of damaged joint tissues, and the ability to reduce inflammation and associated pain. ASCs have different advantages, including easy cryopreservation, faster expansion in culture, more passage cells that retain stem cell phenotypes and pluripotency, and less susceptibility to aging together with lower morbidity of patients. Moreover, the obtention of adipose tissue is much less expensive than bone marrow, with less invasive intervention and available in greater quantities.

ASCs in OA may offer an exciting possibility to improve function, pain, and cartilage volume of the joint, suggestive of a good therapeutic strategy for OA. Despite this, further long-term studies are needed to prove and evaluate the effectiveness of ASCs in OA treatment and their safety capacity. However, it is important to establish a standardized therapeutic protocol for this biological therapy, and assess each patient and each pathology individually.

Author Contributions: All authors contributed equally. All authors read and approved the final manuscript.

Funding: This work was funded by García Cugat foundation CEU-UCH chair and CEU Cardenal Herrera University, CEU Universities.

Acknowledgments: The authors are grateful to the Fundación Garcia Cugat, CEU Cardenal Herrera University and CEU Universities for their technical support.

Int. J. Mol. Sci. 2018, 19, 1926 9 of 14

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ACS Autologous conditioned serum ASCs Adipose-derived mesenchymal stem cells

EGF Epidermal growth factor
EMA European Medicines Agency
FDA Food and Drug Administration

GFs Growth factors HA Hyaluronic acid

hUC-MSCs Human umbilical cord mesenchymal stem cells

IA Intra-articular

IGF-I Insulin-like growth factor-I MSCs Mesenchymal stem cells

OA Osteoarthritis

PRGFs Plasma rich in growth factors PRP Platelet rich plasma RM Regenerative medicine

TGF-β Transforming growth factor-beta VEGF Vascular endothelial growth factor

HTO High tibia osteotomy

References

- Maldonado, M.; Nam, J. The role of changes in extracellular matrix of cartilage in the presence of inflammation on the pathology of osteoarthritis. Biomed. Res. Int. 2013, 2013, 2013, 208873. [CrossRef] [PubMed]
- Fransen, M.; Bridgett, L.; March, L.; Hoy, D.; Penserga, E.; Brooks, P. The epidemiology of osteoarthritis in Asia. Int. J. Rheum. Dis. 2011, 14, 113–121. [CrossRef] [PubMed]
- Orozco, L.; Munar, A.; Soler, R.; Alberca, M.; Soler, F.; Huguet, M.; Sentis, J.; Sanchez, A.; Garcia-Sancho, J.
 Treatment of knee osteoarthritis with autologous mesenchymal stem cells: A pilot study. *Transplantation*2013, 95, 1535–1541. [CrossRef] [PubMed]
- Van der Kraan, P.M.; van den Berg, W.B. Chondrocyte hypertrophy and osteoarthritis: Role in initiation and progression of cartilage degeneration? Osteoarthr. Cartil. 2012, 20, 223–232. [CrossRef] [PubMed]
- Cross, M.; Smith, E.; Hoy, D.; Nolte, S.; Ackerman, I.; Fransen, M.; Bridgett, L.; Williams, S.; Guillemin, F.; Hill, C.L.; et al. The global burden of hip and knee osteoarthritis: Estimates from the global burden of disease 2010 study. Ann. Rheum. Dis. 2014, 73, 1323–1330. [CrossRef] [PubMed]
- Goldring, S.R.; Goldring, M.B. Changes in the osteochondral unit during osteoarthritis: Structure, function and cartilage-bone crosstalk. Nat. Rev. Rheumatol. 2016. 12, 632–644. [CrossRef] [PubMed]
- Kotlarz, H.; Gunnarsson, C.L.; Fang, H.; Rizzo, J.A. Insurer and out-of-pocket costs of osteoarthritis in the US: Evidence from national survey data. Arthrit. Rheumatol. 2009, 60, 3546–3553. [CrossRef] [PubMed]
- Neogi, T.; Zhang, Y. Epidemiology of osteoarthritis. Rheum. Dis. Clin. N. Am. 2013, 39, 1–19. [CrossRef] [PubMed]
- Glyn-Jones, S.; Palmer, A.J.; Agricola, R.; Price, A.J.; Vincent, T.L.; Weinans, H.; Carr, A.J. Osteoarthritis. Lancet 2015, 386, 376–387. [CrossRef]
- Jo, C.H.; Chai, J.W.; Jeong, E.C.; Oh, S.; Shin, J.S.; Shim, H.; Yoon, K.S. Intra-articular Injection of Mesenchymal Stem Cells for the Treatment of Osteoarthritis of the Knee: A 2-Year Follow-up Study. Am. J. Sports Med. 2017, 45, 2774–2783. [CrossRef] [PubMed]
- Martin, J.A.; Brown, T.; Heiner, A.; Buckwalter, J.A. Post-traumatic osteoarthritis: The role of accelerated chondrocyte senescence. *Biorheology* 2004, 41, 479–491. [PubMed]
- Rahmati, M.; Nalesso, G.; Mobasheri, A.; Mozafari, M. Aging and osteoarthritis: Central role of the extracellular matrix. Ageing Res. Rev. 2017, 40, 20–30. [CrossRef] [PubMed]
- Puljak, L.; Marin, A.; Vrdoljak, D.; Markotic, F.; Utrobicic, A.; Tugwell, P. Celecoxib for osteoarthritis. Cochrane Database Syst. Rev. 2017, 5, CD009865. [CrossRef] [PubMed]

De Lange-Brokaar, B.J.; Ioan-Facsinay, A.; Yusuf, E.; Visser, A.W.; Kroon, H.M.; van Osch, G.J.;
 Zuurmond, A.M.; Stojanovic-Susulic, V.; Bloem, J.L.; Nelissen, R.G.; et al. Association of pain in knee osteoarthritis with distinct patterns of synovitis. *Arthrit. Rheumatol.* 2015, 67, 733–740. [CrossRef] [PubMed]

- Garay-Mendoza, D.; Villarreal-Martinez, L.; Garza-Bedolla, A.; Perez-Garza, D.M.; Acosta-Olivo, C.; Vilchez-Cavazos, F.; Diaz-Hutchinson, C.; Gomez-Almaguer, D.; Jaime-Perez, J.C.; Mancias-Guerra, C. The effect of intra-articular injection of autologous bone marrow stem cells on pain and knee function in patients with osteoarthritis. *Int. J. Rheum. Dis.* 2018, 21, 140–147. [CrossRef] [PubMed]
- Wehling, P.; Evans, C.; Wehling, J.; Maixner, W. Effectiveness of intra-articular therapies in osteoarthritis: A literature review. Ther. Adv. Musculoskelet. Dis. 2017, 9, 183–196. [CrossRef] [PubMed]
- Sakata, K.; Furumatsu, T.; Abe, N.; Miyazawa, S.; Sakoma, Y.; Ozaki, T. Histological analysis of failed cartilage repair after marrow stimulation for the treatment of large cartilage defect in medial compartmental osteoarthritis of the knee. Acta Med. Okayama 2013, 67, 65–74. [PubMed]
- Vinatier, C.; Guicheux, J. Cartilage tissue engineering: From biomaterials and stem cells to osteoarthritis treatments. Ann. Phys. Rehabil. Med. 2016, 59, 139–144. [CrossRef] [PubMed]
- Platas, J.; Guillen, M.I.; Perez Del Caz, M.D.; Gomar, F.; Castejon, M.A.; Mirabet, V.; Alcaraz, M.J. Paracrine
 effects of human adipose-derived mesenchymal stem cells in inflammatory stress-induced senescence
 features of osteoarthritic chondrocytes. Aging 2016, 8, 1703–1717. [CrossRef] [PubMed]
- Meirelles Lda, S.; Fontes, A.M.; Covas, D.T.; Caplan, A.I. Mechanisms involved in the therapeutic properties
 of mesenchymal stem cells. Cytokine Growth Factor Rev. 2009, 20, 419–427. [CrossRef] [PubMed]
- Zuk, P.A.; Zhu, M.; Mizuno, H.; Huang, J.; Futrell, J.W.; Katz, A.J.; Benhaim, P.; Lorenz, H.P.; Hedrick, M.H. Multilineage cells from human adipose tissue: Implications for cell-based therapies. *Tissue Eng.* 2001, 7, 211–228. [CrossRef] [PubMed]
- 22. Wang, H.; Yan, X.; Jiang, Y.; Wang, Z.; Li, Y.; Shao, Q. The human umbilical cord stem cells improve the viability of OA degenerated chondrocytes. *Mol. Med. Rep.* **2018**, *17*, 4474–4482. [CrossRef] [PubMed]
- Kern, S.; Eichler, H.; Stoeve, J.; Kluter, H.; Bieback, K. Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. Stem. Cells 2006, 24, 1294–1301. [CrossRef] [PubMed]
- Dmitrieva, R.I.; Minullina, I.R.; Bilibina, A.A.; Tarasova, O.V.; Anisimov, S.V.; Zaritskey, A.Y. Bone marrowand subcutaneous adipose tissue-derived mesenchymal stem cells: Differences and similarities. *Cell Cycle* 2012, 11, 377–383. [CrossRef] [PubMed]
- Chen, H.T.; Lee, M.J.; Chen, C.H.; Chuang, S.C.; Chang, L.F.; Ho, M.L.; Hung, S.H.; Fu, Y.C.; Wang, Y.H.; Wang, H.I.; et al. Proliferation and differentiation potential of human adipose-derived mesenchymal stem cells isolated from elderly patients with osteoporotic fractures. *J. Cell. Mol. Med.* 2012, 16, 582–593. [CrossRef] [PubMed]
- Murphy, J.M.; Fink, D.J.; Hunziker, E.B.; Barry, F.P. Stem cell therapy in a caprine model of osteoarthritis. Arthrit. Rheum. 2003, 48, 3464–3474. [CrossRef] [PubMed]
- Zhu, Y.; Liu, T.; Song, K.; Fan, X.; Ma, X.; Cui, Z. Adipose-derived stem cell: A better stem cell than BMSC. Cell Biochem. Funct. 2008, 26, 664–675. [CrossRef] [PubMed]
- Mirsaidi, A.; Kleinhans, K.N.; Rimann, M.; Tiaden, A.N.; Stauber, M.; Rudolph, K.L.; Richards, P.J. Telomere length, telomerase activity and osteogenic differentiation are maintained in adipose-derived stromal cells from senile osteoporotic SAMP6 mice. J. Tissue Eng. Regen. Med. 2012, 6, 378–390. [CrossRef] [PubMed]
- Schaffler, A.; Buchler, C. Concise review: Adipose tissue-derived stromal cells-basic and clinical implications for novel cell-based therapies. Stem Cells 2007, 25, 818–827. [CrossRef] [PubMed]
- Spasovski, D.; Spasovski, V.; Bascarevic, Z.; Stojiljkovic, M.; Vreca, M.; Andelkovic, M.; Pavlovic, S. Intra-articular injection of autologous adipose-derived mesenchymal stem cells in the treatment of knee osteoarthritis. J. Gene Med. 2018, 20, e3002. [CrossRef] [PubMed]
- Cui, G.H.; Wang, Y.Y.; Li, C.J.; Shi, C.H.; Wang, W.S. Efficacy of mesenchymal stem cells in treating patients with osteoarthritis of the knee: A meta-analysis. Exp. Ther. Med. 2016, 12, 3390–3400. [CrossRef] [PubMed]
- 32. Gupta, P.K.; Das, A.K.; Chullikana, A.; Majumdar, A.S. Mesenchymal stem cells for cartilage repair in osteoarthritis. *Stem Cell. Res. Ther.* **2012**, *3*, 25. [CrossRef] [PubMed]
- Hurley, E.T.; Yasui, Y.; Gianakos, A.L.; Seow, D.; Shimozono, Y.; Kerkhoffs, G.; Kennedy, J.G. Limited evidence for adipose-derived stem cell therapy on the treatment of osteoarthritis. Knee Surg. Sports Traumatol. Arthrosc. 2018. [CrossRef] [PubMed]

 Smith, E.; Hoy, D.G.; Cross, M.; Vos, T.; Naghavi, M.; Buchbinder, R.; Woolf, A.D.; March, L. The global burden of other musculoskeletal disorders: Estimates from the Global Burden of Disease 2010 study. Ann. Rheum. Dis. 2014, 73, 1462–1469. [CrossRef] [PubMed]

- Freitag, J.; Li, D.; Wickham, J.; Shah, K.; Tenen, A. Effect of autologous adipose-derived mesenchymal stem cell therapy in the treatment of a post-traumatic chondral defect of the knee. BMJ Case Rep. 2017, 2017, bcr-2017. [CrossRef] [PubMed]
- Mirza, Y.H.; Oussedik, S. Is there a role for stem cells in treating articular injury? Br. J. Hosp. Med. 2017, 78, 372–377. [CrossRef] [PubMed]
- Niemeyer, P.; Steinwachs, M.; Erggelet, C.; Kreuz, P.C.; Kraft, N.; Kostler, W.; Mehlhorn, A.; Sudkamp, N.P. Autologous chondrocyte implantation for the treatment of retropatellar cartilage defects: Clinical results referred to defect localisation. Arch. Orthop. Trauma Surg. 2008, 128, 1223–1231. [CrossRef] [PubMed]
- 38. Meijer, H.; Reinecke, J.; Becker, C.; Tholen, G.; Wehling, P. The production of anti-inflammatory cytokines in whole blood by physico-chemical induction. *Inflamm. Res.* 2003, 52, 404–407. [CrossRef] [PubMed]
- 39. Baltzer, A.W.; Moser, C.; Jansen, S.A.; Krauspe, R. Autologous conditioned serum (Orthokine) is an effective treatment for knee osteoarthritis. Osteoarthr. Cartil. 2009, 17, 152–160. [CrossRef] [PubMed]
- Baselga Garcia-Escudero, J.; Miguel Hernandez Trillos, P. Treatment of Osteoarthritis of the Knee with a Combination of Autologous Conditioned Serum and Physiotherapy: A Two-Year Observational Study. PLoS ONE 2015, 10, e0145551. [CrossRef] [PubMed]
- Anitua, E.; Sanchez, M.; Aguirre, J.J.; Prado, R.; Padilla, S.; Orive, G. Efficacy and safety of plasma rich in growth factors intra-articular infiltrations in the treatment of knee osteoarthritis. *Arthroscopy* 2014, 30, 1006–1017. [CrossRef] [PubMed]
- Anitua, E.; Sanchez, M.; Orive, G. Potential of endogenous regenerative technology for in situ regenerative medicine. Adv. Drug Deliv. Rev. 2010, 62, 741–752. [CrossRef] [PubMed]
- Raeissadat, S.A.; Rayegani, S.M.; Ahangar, A.G.; Abadi, P.H.; Mojgani, P.; Ahangar, O.G. Efficacy of Intra-articular Injection of a Newly Developed Plasma Rich in Growth Factor (PRGF) Versus Hyaluronic Acid on Pain and Function of Patients with Knee Osteoarthritis: A Single-Blinded Randomized Clinical Trial. Clin. Med. Insights Arthritis Musculoskelet. Disord. 2017, 10, 1179544117733452. [CrossRef] [PubMed]
- 44. Li, G.; Fu, N.; Xie, J.; Fu, Y.; Deng, S.; Cun, X.; Wei, X.; Peng, Q.; Cai, X.; Lin, Y. Poly(3-hydroxybutyrate-co-4-hydroxybutyrate) Based Electrospun 3D Scaffolds for Delivery of Autogeneic Chondrocytes and Adipose-Derived Stem Cells: Evaluation of Cartilage Defects in Rabbit. J. Biomed. Nanotechnol. 2015, 11, 105–116. [CrossRef] [PubMed]
- Scioli, M.G.; Bielli, A.; Gentile, P.; Cervelli, V.; Orlandi, A. Combined treatment with platelet-rich plasma and insulin favours chondrogenic and osteogenic differentiation of human adipose-derived stem cells in three-dimensional collagen scaffolds. J. Tissue Eng. Regen. Med. 2017, 11, 2398–2410. [CrossRef] [PubMed]
- Ikebe, C.; Suzuki, K. Mesenchymal stem cells for regenerative therapy: Optimization of cell preparation protocols. Biomed. Res. Int. 2014, 2014, 951512. [CrossRef] [PubMed]
- Bashir, J.; Sherman, A.; Lee, H.; Kaplan, L.; Hare, J.M. Mesenchymal stem cell therapies in the treatment of musculoskeletal diseases. PM R 2014, 6, 61–69. [CrossRef] [PubMed]
- Pittenger, M.F.; Mackay, A.M.; Beck, S.C.; Jaiswal, R.K.; Douglas, R.; Mosca, J.D.; Moorman, M.A.; Simonetti, D.W.; Craig, S.; Marshak, D.R. Multilineage potential of adult human mesenchymal stem cells. Science 1999, 284, 143–147. [CrossRef] [PubMed]
- Noth, U.; Steinert, A.F.; Tuan, R.S. Technology insight: Adult mesenchymal stem cells for osteoarthritis therapy. Nat. Clin. Pract. Rheumatol. 2008, 4, 371–380. [CrossRef] [PubMed]
- Munoz-Criado, I.; Meseguer-Ripolles, J.; Mellado-Lopez, M.; Alastrue-Agudo, A.; Griffeth, R.J.; Forteza-Vila, J.; Cugat, R.; Garcia, M.; Moreno-Manzano, V. Human Suprapatellar Fat Pad-Derived Mesenchymal Stem Cells Induce Chondrogenesis and Cartilage Repair in a Model of Severe Osteoarthritis. Stem Cells Int. 2017, 2017, 4758930. [CrossRef] [PubMed]
- 51. Peng, L.; Jia, Z.; Yin, X.; Zhang, X.; Liu, Y.; Chen, P.; Ma, K.; Zhou, C. Comparative analysis of mesenchymal stem cells from bone marrow, cartilage, and adipose tissue. *Stem Cells Dev.* **2008**, *17*, 761–773. [CrossRef] [PubMed]

 Alvarez-Viejo, M.; Menendez-Menendez, Y.; Blanco-Gelaz, M.A.; Ferrero-Gutierrez, A.; Fernandez-Rodriguez, M.A.; Gala, J.; Otero-Hernandez, J. Quantifying mesenchymal stem cells in the mononuclear cell fraction of bone marrow samples obtained for cell therapy. *Transplant Proc.* 2013, 45, 434–439. [CrossRef] [PubMed]

- Hass, R.; Kasper, C.; Bohm, S.; Jacobs, R. Different populations and sources of human mesenchymal stem cells (MSC): A comparison of adult and neonatal tissue-derived MSC. Cell Commun. Signal 2011, 9, 12. [CrossRef] [PubMed]
- Macrin, D.; Joseph, J.P.; Pillai, A.A.; Devi, A. Eminent Sources of Adult Mesenchymal Stem Cells and Their Therapeutic Imminence. Stem Cell Rev. 2017, 13, 741–756. [CrossRef] [PubMed]
- Feng, C.; Luo, X.; He, N.; Xia, H.; Lv, X.; Zhang, X.; Li, D.; Wang, F.; He, J.; Zhang, L.; et al. Efficacy and Persistence of Allogeneic Adipose-Derived Mesenchymal Stem Cells Combined with Hyaluronic Acid in Osteoarthritis After Intra-articular Injection in a Sheep Model. Tissue Eng. Part A 2018, 24, 219–233. [CrossRef] [PubMed]
- Jevotovsky, D.S.; Alfonso, A.R.; Einhorn, T.A.; Chiu, E.S. Osteoarthritis and stem cell therapy in humans: A systematic review. Osteoarthr. Cartil. 2018, 26, 711–729. [CrossRef] [PubMed]
- Pelttari, K.; Steck, E.; Richter, W. The use of mesenchymal stem cells for chondrogenesis. *Injury* 2008, 39 (Suppl. 1), S58–S65. [CrossRef] [PubMed]
- Caplan, A.I. Why are MSCs therapeutic? New data: New insight. J. Pathol. 2009, 217, 318–324. [CrossRef] [PubMed]
- Maumus, M.; Roussignol, G.; Toupet, K.; Penarier, G.; Bentz, I.; Teixeira, S.; Oustric, D.; Jung, M.; Lepage, O.;
 Steinberg, R.; et al. Utility of a Mouse Model of Osteoarthritis to Demonstrate Cartilage Protection by IFNgamma-Primed Equine Mesenchymal Stem Cells. Front. Immunol. 2016, 7, 392. [CrossRef] [PubMed]
- 60. Caplan, A.I.; Correa, D. The MSC: An injury drugstore. Cell Stem Cell 2011, 9, 11–15. [CrossRef] [PubMed]
- Longobardi, L.; O'Rear, L.; Aakula, S.; Johnstone, B.; Shimer, K.; Chytil, A.; Horton, W.A.; Moses, H.L.; Spagnoli, A. Effect of IGF-I in the chondrogenesis of bone marrow mesenchymal stem cells in the presence or absence of TGF-beta signaling. J. Bone Min. Res. 2006, 21, 626–636. [CrossRef] [PubMed]
- Ozeki, N.; Muneta, T.; Koga, H.; Nakagawa, Y.; Mizuno, M.; Tsuji, K.; Mabuchi, Y.; Akazawa, C.; Kobayashi, E.; Matsumoto, K.; et al. Not single but periodic injections of synovial mesenchymal stem cells maintain viable cells in knees and inhibit osteoarthritis progression in rats. Osteoarthr. Cartil. 2016, 24, 1061–1070. [CrossRef] [PubMed]
- Maumus, M.; Manferdini, C.; Toupet, K.; Peyrafitte, J.A.; Ferreira, R.; Facchini, A.; Gabusi, E.; Bourin, P.; Jorgensen, C.; Lisignoli, G.; et al. Adipose mesenchymal stem cells protect chondrocytes from degeneration associated with osteoarthritis. Stem Cell Res. 2013, 11, 834–844. [CrossRef] [PubMed]
- 64. Almeida, H.V.; Cunniffe, G.M.; Vinardell, T.; Buckley, C.T.; O'Brien, F.J.; Kelly, D.J. Coupling Freshly Isolated CD44⁺ Infrapatellar Fat Pad-Derived Stromal Cells with a TGF-β3 Eluting Cartilage ECM-Derived Scaffold as a Single-Stage Strategy for Promoting Chondrogenesis. Adv. Healthc. Mater. 2015, 4, 1043–1053. [CrossRef] [PubMed]
- Kang, H.; Peng, J.; Lu, S.; Liu, S.; Zhang, L.; Huang, J.; Sui, X.; Zhao, B.; Wang, A.; Xu, W.; et al. In vivo cartilage repair using adipose-derived stem cell-loaded decellularized cartilage ECM scaffolds. J. Tissue Eng. Regen. Med. 2014, 8, 442–453. [CrossRef] [PubMed]
- 66. Choi, B.; Kim, S.; Fan, J.; Kowalski, T.; Petrigliano, F.; Evseenko, D.; Lee, M. Covalently conjugated transforming growth factor-β1 in modular chitosan hydrogels for the effective treatment of articular cartilage defects. Biomater. Sci. 2015, 3, 742–752. [CrossRef] [PubMed]
- Zhang, K.; Yan, S.; Li, G.; Cui, L.; Yin, J. In-situ birth of MSCs multicellular spheroids in poly(L-glutamic acid)/chitosan scaffold for hyaline-like cartilage regeneration. *Biomaterials* 2015, 71, 24–34. [CrossRef] [PubMed]
- Sun, Q.; Zhang, L.; Xu, T.; Ying, J.; Xia, B.; Jing, H.; Tong, P. Combined use of adipose derived stem cells and TGF-β3 microspheres promotes articular cartilage regeneration in vivo. *Biotech. Histochem.* 2018, 93, 168–176. [CrossRef] [PubMed]
- Desance, M.; Contentin, R.; Bertoni, L.; Gomez-Leduc, T.; Branly, T.; Jacquet, S.; Betsch, J.M.; Batho, A.; Legendre, F.; Audigie, F.; et al. Chondrogenic Differentiation of Defined Equine Mesenchymal Stem Cells Derived from Umbilical Cord Blood for Use in Cartilage Repair Therapy. *Int. J. Mol. Sci.* 2018, 19, 537. [CrossRef] [PubMed]

 Abbas, M. Combination of bone marrow mesenchymal stem cells and cartilage fragments contribute to enhanced repair of osteochondral defects. *Bioinformation* 2017, 13, 196–201. [CrossRef] [PubMed]

- 71. Murphy, M.P.; Buckley, C.; Sugrue, C.; Carr, E.; O'Reilly, A.; O'Neill, S.; Carroll, S.M. ASCOT: Autologous Bone Marrow Stem Cell Use for Osteoarthritis of the Thumb-First Carpometacarpal Joint. *Plast. Reconstr. Surg. Glob. Open* 2017, 5, e1486. [CrossRef] [PubMed]
- Pers, Y.M.; Rackwitz, L.; Ferreira, R.; Pullig, O.; Delfour, C.; Barry, F.; Sensebe, L.; Casteilla, L.; Fleury, S.;
 Bourin, P.; et al. Adipose Mesenchymal Stromal Cell-Based Therapy for Severe Osteoarthritis of the Knee: A
 Phase I Dose-Escalation Trial. Stem Cells Transl. Med. 2016, 5, 847–856. [CrossRef] [PubMed]
- Al-Najar, M.; Khalil, H.; Al-Ajlouni, J.; Al-Antary, E.; Hamdan, M.; Rahmeh, R.; Alhattab, D.; Samara, O.; Yasin, M.; Abdullah, A.A.; et al. Intra-articular injection of expanded autologous bone marrow mesenchymal cells in moderate and severe knee osteoarthritis is safe: A phase I/II study. J. Orthop. Surg. Res. 2017, 12, 190. [CrossRef] [PubMed]
- Jo, C.H.; Lee, Y.G.; Shin, W.H.; Kim, H.; Chai, J.W.; Jeong, E.C.; Kim, J.E.; Shim, H.; Shin, J.S.; Shin, I.S.; et al. Intra-articular injection of mesenchymal stem cells for the treatment of osteoarthritis of the knee: A proof-of-concept clinical trial. Stem Cells 2014, 32, 1254–1266. [CrossRef] [PubMed]
- Wu, Y.; Gong, Z.; Li, J.; Meng, Q.; Fang, W.; Long, X. The pilot study of fibrin with temporomandibular joint derived synovial stem cells in repairing TMJ disc perforation. *Biomed. Res. Int.* 2014, 2014, 454021. [CrossRef] [PubMed]
- Cui, D.; Li, H.; Xu, X.; Ye, L.; Zhou, X.; Zheng, L.; Zhou, Y. Mesenchymal Stem Cells for Cartilage Regeneration of TMJ Osteoarthritis. Stem Cells Int. 2017, 2017, 5979741. [CrossRef] [PubMed]
- Yang, C.; Robbins, P.D. Immunosuppressive exosomes: A new approach for treating arthritis. Int. J. Rheumatol. 2012, 2012, 573528. [CrossRef] [PubMed]
- Toh, W.S.; Lai, R.C.; Hui, J.H.P.; Lim, S.K. MSC exosome as a cell-free MSC therapy for cartilage regeneration: Implications for osteoarthritis treatment. Semin. Cell Dev. Biol. 2017, 67, 56–64. [CrossRef] [PubMed]
- Zhang, S.; Chu, W.C.; Lai, R.C.; Lim, S.K.; Hui, J.H.; Toh, W.S. Exosomes derived from human embryonic mesenchymal stem cells promote osteochondral regeneration. *Osteoarthr. Cartil.* 2016, 24, 2135–2140. [CrossRef] [PubMed]
- McIntosh, K.R.; Frazier, T.; Rowan, B.G.; Gimble, J.M. Evolution and future prospects of adipose-derived immunomodulatory cell therapeutics. Expert Rev. Clin. Immunol. 2013, 9, 175–184. [CrossRef] [PubMed]
- Baer, P.C. Adipose-derived mesenchymal stromal/stem cells: An update on their phenotype in vivo and in vitro. World J. Stem. Cells 2014, 6, 256–265. [CrossRef] [PubMed]
- 82. Li, J.; Wong, W.H.; Chan, S.; Chim, J.C.; Cheung, K.M.; Lee, T.L.; Au, W.Y.; Ha, S.Y.; Lie, A.K.; Lau, Y.L.; et al. Factors affecting mesenchymal stromal cells yield from bone marrow aspiration. *Chin. J. Cancer Res.* **2011**, 23, 43–48. [CrossRef] [PubMed]
- Ding, D.C.; Chang, Y.H.; Shyu, W.C.; Lin, S.Z. Human umbilical cord mesenchymal stem cells: A new era for stem cell therapy. Cell Transpl. 2015, 24, 339–347. [CrossRef] [PubMed]
- 84. Tangchitphisut, P.; Srikaew, N.; Numhom, S.; Tangprasittipap, A.; Woratanarat, P.; Wongsak, S.; Kijkunasathian, C.; Hongeng, S.; Murray, I.R.; Tawonsawatruk, T. Infrapatellar Fat Pad: An Alternative Source of Adipose-Derived Mesenchymal Stem Cells. Arthritis 2016, 2016, 4019873. [CrossRef] [PubMed]
- Hindle, P.; Khan, N.; Biant, L.; Peault, B. The Infrapatellar Fat Pad as a Source of Perivascular Stem Cells with Increased Chondrogenic Potential for Regenerative Medicine. Stem Cells Transl. Med. 2017, 6, 77–87.
 [CrossRef] [PubMed]
- Yun, S.; Ku, S.K.; Kwon, Y.S. Adipose-derived mesenchymal stem cells and platelet-rich plasma synergistically ameliorate the surgical-induced osteoarthritis in Beagle dogs. J. Orthop. Surg. Res. 2016, 11, 9. [CrossRef] [PubMed]
- Song, Y.; Du, H.; Dai, C.; Zhang, L.; Li, S.; Hunter, D.J.; Lu, L.; Bao, C. Human adipose-derived mesenchymal stem cells for osteoarthritis: A pilot study with long-term follow-up and repeated injections. *Regen. Med.* 2018, 13, 295–307. [CrossRef] [PubMed]
- 88. Wakitani, S.; Imoto, K.; Yamamoto, T.; Saito, M.; Murata, N.; Yoneda, M. Human autologous culture expanded bone marrow mesenchymal cell transplantation for repair of cartilage defects in osteoarthritic knees. Osteoarthr. Cartil. 2002, 10, 199–206. [CrossRef] [PubMed]

 Kuroda, R.; Ishida, K.; Matsumoto, T.; Akisue, T.; Fujioka, H.; Mizuno, K.; Ohgushi, H.; Wakitani, S.; Kurosaka, M. Treatment of a full-thickness articular cartilage defect in the femoral condyle of an athlete with autologous bone-marrow stromal cells. Osteoarthr. Cartil. 2007, 15, 226–231. [CrossRef] [PubMed]

- Orozco, L.; Munar, A.; Soler, R.; Alberca, M.; Soler, F.; Huguet, M.; Sentis, J.; Sanchez, A.; Garcia-Sancho, J.
 Treatment of knee osteoarthritis with autologous mesenchymal stem cells: Two-year follow-up results.
 Transplantation 2014, 97, e66–e68. [CrossRef] [PubMed]
- Zhang, Q.; Chen, Y.; Wang, Q.; Fang, C.; Sun, Y.; Yuan, T.; Wang, Y.; Bao, R.; Zhao, N. Effect of bone marrow-derived stem cells on chondrocytes from patients with osteoarthritis. Mol. Med. Rep. 2016, 13, 1795–1800. [CrossRef] [PubMed]
- Davatchi, F.; Sadeghi Abdollahi, B.; Mohyeddin, M.; Nikbin, B. Mesenchymal stem cell therapy for knee osteoarthritis: 5 years follow-up of three patients. Int. J. Rheum. Dis. 2016, 19, 219–225. [CrossRef] [PubMed]
- Wu, L.; Prins, H.J.; Helder, M.N.; van Blitterswijk, C.A.; Karperien, M. Trophic effects of mesenchymal stem cells in chondrocyte co-cultures are independent of culture conditions and cell sources. *Tissue Eng. Part A* 2012, 18, 1542–1551. [CrossRef] [PubMed]
- 94. Meretoja, V.V.; Dahlin, R.L.; Kasper, F.K.; Mikos, A.G. Enhanced chondrogenesis in co-cultures with articular chondrocytes and mesenchymal stem cells. *Biomaterials* 2012, 33, 6362–6369. [CrossRef] [PubMed]
- Zheng, P.; Ju, L.; Jiang, B.; Chen, L.; Dong, Z.; Jiang, L.; Wang, R.; Lou, Y. Chondrogenic differentiation of human umbilical cord bloodderived mesenchymal stem cells by coculture with rabbit chondrocytes. Mol. Med. Rep. 2013, 8, 1169–1182. [CrossRef] [PubMed]
- Zhu, Y.; Guan, Y.M.; Huang, H.L.; Wang, Q.S. Human umbilical cord blood mesenchymal stem cell transplantation suppresses inflammatory responses and neuronal apoptosis during early stage of focal cerebral ischemia in rabbits. *Acta Pharmacol. Sin.* 2014, 35, 585–591. [CrossRef] [PubMed]
- Min, F.; Gao, F.; Li, Q.; Liu, Z. Therapeutic effect of human umbilical cord mesenchymal stem cells modified by angiotensin-converting enzyme 2 gene on bleomycin-induced lung fibrosis injury. Mol. Med. Rep. 2015, 11, 2387–2396. [CrossRef] [PubMed]
- 98. Chapman, V.; Markides, H.; Sagar, D.R.; Xu, L.; Burston, J.J.; Mapp, P.; Kay, A.; Morris, R.H.; Kehoe, O.; El Haj, A.J. Therapeutic Benefit for Late, but Not Early, Passage Mesenchymal Stem Cells on Pain Behaviour in an Animal Model of Osteoarthritis. Stem Cells Int. 2017, 2017, 205104. [CrossRef] [PubMed]
- Akamatsu, Y.; Koshino, T.; Saito, T.; Wada, J. Changes in osteosclerosis of the osteoarthritic knee after high tibial osteotomy. Clin. Orthop. Relat. Res. 1997, 207–214. [CrossRef]
- Sabzevari, S.; Ebrahimpour, A.; Roudi, M.K.; Kachooei, A.R. High Tibial Osteotomy: A Systematic Review and Current Concept. Arch. Bone Jt. Surg. 2016, 4, 204–212. [PubMed]
- 101. Martin, R.; Birmingham, T.B.; Willits, K.; Litchfield, R.; Lebel, M.E.; Giffin, J.R. Adverse event rates and classifications in medial opening wedge high tibial osteotomy. Am. J. Sports Mat. 2014, 42, 1118–1126. [CrossRef] [PubMed]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).

			4	
TRATAMIENTO	OA	Y	ARTICULOS	



4.2.2. Plasma rico en Plaquetas

El plasma rico en plaquetas (PRP comenzó a utilizarse a principios de los 90 en cirugías cardiacas (Ferrari y cols, 1987), para posteriormente ampliarse su espectro de acción sobre todo en la cirugía maxilofacial y estética, y más adelante comenzó a utilizarse también en traumatología (Floryan y Berghoff 2004; Sampson y cols, 2008)

El PRP es un producto de origen biológico, que se obtiene mediante el procesado de la sangre del propio paciente. En este procedimiento se obtiene un derivado que contiene una concentración más o menos variable de plaquetas, así como factores plasmáticos de crecimiento (Marx, 2001).

4.2.2.1. Eficacia en Aparato locomotor

La eficacia de las terapias con PRP o sus derivados a la hora de la reparación de defectos óseos, condrales, musculares y/o tendinosos en animales ha quedado comprobada (Serra y cols, 2013; Thor y cols, 2013; Cugat y cols, 2017). Sin embargo, en el caso concreto de las fracturas, y el tratamiento de sus complicaciones en la consolidación ósea las publicaciones en medicina humana son muy escasas (Zhang y cols, 2003; Seijas y cols, 2010) y, en el campo de la medicina veterinaria, ausentes.

TRATAMIENTO OA Y ARTÍCULOS



López S, Vilar JM, Sopena JJ, Damià E, Chicharro D, Carrillo JM, Cuervo B, Rubio M. Assessment of the Efficacy of Platelet-Rich Plasma in the Treatment of Traumatic Canine Fractures.

Int J Mol Sci. 2019 Mar 1;20(5). doi: 10.3390/ijms20051075.

JCR Impact factor (2017): 3.687

Cuartil: Q2

Grupo: Chemistry, multidisciplinary

Posición: 52/171

TRATAMIENTO OA Y ARTÍCULOS







Article

Assessment of the Efficacy of Platelet-Rich Plasma in the Treatment of Traumatic Canine Fractures

Sergio López ¹, José M. Vilar ¹0, Joaquín J. Sopena ^{2,3}0, Elena Damià ², Deborah Chicharro ²0, José M. Carrillo ^{2,3},*, Belén Cuervo ²0 and Mónica Rubio ^{2,3}0

- Animal Pathology Department, Instituto Universitario de Investigaciones Biomédicas y Universitarias, Universidad de Las Palmas de Gran Canaria, 35416 Trasmontaña S/N, Arucas, Spain; sergiolopezbarbeta@gmail.com (S.L.); jose.vilar@ulpgc.es (J.M.V.)
- ² Bioregenerative Medicine and Applied Surgery Research Group, Animal Medicine and Surgery Department, Veterinary Faculty, Universidad Cardenal Herrera-CEU, CEU Universities, 46115 Valencia, Spain; jsopena@uchceu.es (J.J.S.); elena.damia@uchceu.es (E.D.); debora.chicharro@uchceu.es (D.C.); belen.cuervo@uchceu.es (B.C.); mrubio@uchceu.es (M.R.)
- ³ García Cugat Foundation CEU-UCH Chair of Medicine and Regenerative Surgery, 08006 Barcelona, Spain
- * Correspondence: jcarrill@uchceu.es; Tel.: +34-96-136-9000 (ext. 66216)

Received: 5 February 2019; Accepted: 27 February 2019; Published: 1 March 2019



Abstract: The role of platelet-rich plasma (PRP) in promoting the healing of bone fractures has not yet been clearly stated. The aim of this prospective clinical study was to evaluate the effectiveness of plasma rich in growth factors (PRGF, a PRP derivate) in the treatment of naturally-occurring bone fractures in dogs. With this objective, sixty-five dogs with radius/ulna or tibia/fibula bone fractures were randomly divided into two groups (PRGF and saline solution (SS) groups) and checked at days 0, 7, 14, 21, 28, 35, 42, 49, 56, 60, 63, 70, 120, and 180. All the fractures were treated with an external skeletal fixation, and pain was controlled with Carprofen. Healing was evaluated by physical examination, limb function, radiography, and by a Likert-type owner satisfaction questionnaire. A faster fracture healing was observed in the PRGF group, with statistically significant differences with respect to the SS group. Swelling at the fracture site was significantly greater at day 14 and 28 in animals injected with PRGF, and more pain on palpation was found in the area at day 28. The injection of PRGF in acute bone fractures accelerates bone healing.

Keywords: PRGF; Carprofen; dog; fracture; bone healing

1. Introduction

Plasma-rich growth factors (PRGF) are currently being used to promote bone healing in reconstructive surgeries [1–3]. In canine models, several experimental studies have published the effect of this platelet rich plasma derivate in osteoarthritis with differing results [4–7]. Platelets are very important in the wound healing process [1]; they rapidly arrive at the wound side and begin the coagulation process. In addition, they release multiple wound-healing growth factors and cytokines within 10 min [1–3]. Platelets are viable for seven days and will continue to release growth factors into the tissue during this time [8].

The use of PRGF is based on the assumption that higher platelet concentrations release significant quantities of growth factors, which aids in bone healing [9–11]. Specifically, growth factors are thought to be a contributing factor in bone regeneration and in increasing vascularization, which are vital features of the bone-healing process [6].

Treatments with PRGF have given excellent clinical results in oral and maxillofacial surgery in humans [9,12], and in bone and cartilage healing in animal studies [7,13,14]. Growth factors have also

Int. J. Mol. Sci. 2019, 20, 1075; doi:10.3390/ijms20051075

www.mdpi.com/journal/ijms

Int. J. Mol. Sci. 2019, 20, 1075

been used in the treatment of large wounds and skin defects in burn patients [15–17]. However, some controversial results can be found in the cited literature; therefore, the effectiveness of this technique requires further research.

To the authors' knowledge, articles discussing fresh fractures and delayed fracture healing are very scarce [18,19]. In the veterinary field, no publications were found regarding the use of PRGF in fractures.

In this study, the dogs used were clinical patients, but also clinical animal models. In the present study, we hypothesize that treating canine bone fractures with PRGF would accelerate bone healing. Thus, the aim of this clinical trial is to evaluate the use of PRGF in the treatment of naturally occurring bone fractures in dogs.

2. Results

A total of 68 dogs were initially evaluated; however, only 43 met the necessary requirements to be included in the study.

The dogs were randomly assigned to either PGRF or SS groups. Twenty dogs were included in the PRGF group (47%) and 23 in the SS group (53%). The results for each dog belonging to either the PRGF or the SS groups are summarized in their respective tables (Tables 1 and 2).

The mean weight for each group was 16.27 kg for the PRGF group and 13.07 kg for the SS group. Mean age was 40.85 and 57.17 months, respectively, with no statistical differences between groups in these parameters ($p \ge 0.08$).

During the study, all the animals received Carprofen as a rescue analgesia at least one time during the first seven days except for 2 and 4 patients in the PRGF and SS groups, respectively, with no statistical differences between groups ($p \ge 0.05$).

The time (mean \pm SD) for implant removal was 41.3 ± 11.73 days in the PRGF group and 49 ± 12.12 days in the SS group. This difference was statistically significant (p=0.03) (Figure 1).

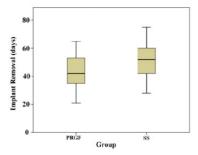


Figure 1. Boxplot corresponding to the days of implant removal for both PRGF and SS groups. Mean time was significantly higher in the SS group.

The time when full weight support was detected was 22.1 ± 13.64 days and 25.47 ± 14.9 days in the PRGF and SS groups, respectively; however, this difference was not statistically significant (p=0.45). All animals were sound within six months post-surgery.

Swelling in the fracture site was present in both groups up to day 14 without statistically significant differences between the groups. Between days 14 and 28, swelling was still present in the PRGF group (p < 0.048).

The joint movement evaluation showed almost 100% joint mobility without differences between groups in any of the checking periods.

The evaluation of pain on palpation showed statistically significant differences at day 28 between groups, where pain was still present in the PRGF group (p = 0.041).

Int. J. Mal. Sci. 2019, 20, 1075 3 of 10

Table 1. Individual data and main results for PRGF group.

Dog#	Breed	Gender	Weight (kg)	Age (months)	Fracture L	Configuration	Weight Support	Time I Removal (days) *	Complications	Analgesia
1	G DANE	M	53	11	U/R	TYPE IIB 2X3	7	35 (39)	NO	Y
2	CROSSBREED	F	4	24	U/R	TYPE IIB 1X2	7	42 (45)	NO	N
3	CROSSBREED	M	17,8	36	U/R	TYPE IIB 2x4	21	28 (32)	NO	Y
4	CROSSBREED	F	8,5	96	U/R	TYPE IIB 2X3	14	56 (60)	GE	Y
5	CROSSBREED	F	4	3	T/F	TYPE IIB 2X3	7	21 (21)	NO	Y
6	CROSSBREED	M	4	36	U/R	TYPE IIB 2X3	28	63 (63)	NO	Y
7	CROSSBREED	M	7	48	T/F	TYPE IIB 2X3	21	35 (40)	PL	Y
8	CROSSBREED	F	6	6	T/F	TYPE IIB	21	28 (30)	NO	Y
9	CROSSBREED	F	2,3	60	U/R	TYPE IIB 2X3	21	42 (42)	PL	Y
10	CROSSBREED	F	22	12	U/R	TYPE IIB 2X3	14	35 (36)	NO	Y
11	CROSSBREED	F	72	20	U/R	TYPE IIB 2X3	60	56 (56)	NO	Y
12	CROSSBREED	M	6,3	48	U/R	TYPE IIB 2X3	60	42 (45)	NO	Y
13	CROSSBREED	M	4	12	U/R	TYPE IIB 2X3	7	56 (56)	NO	Y
14	BELG SHEPH	F	16	96	U/R	TYPE IIB 2X3	21	49 (49)	PL	Y
15	CROSSBREED	M	4,5	24	U/R	TYPE IIB 2X3	21	28 (30)	NO	Y
16	PODENCO	M	22	56	U/R	TYPE IIB 2X3	28	28 (28)	NO	Y
17	CROSSBREED	M	6	70	U/R	TYPE IIB 2X3	21	63 (65)	NO	Υ
18	RAT VAL	F	2	24	U/R	TYPE IIB 2X3	21	49 (50)	NO	Y
19	MASTIFF	M	52	36	T/F	TYPE IIB 2X3	28	35 (40)	NO	Y
20	CROSSBREED	M	12	99	U/R	TYPE IIB 2X3	14	35 (35)	NO	N

^{*} The first number references the checking day when stage 4/5 was reached radiographically and the implant was ready for removal; the number in parenthesis refers to the day the implant was removed. RAT VAL: Rationero Valenciano.

Int. J. Mal. Sci. 2019, 20, 1075

Table 2. Individual data and main results for SS group.

Dog#	Breed	Gender	Weight (kg)	Age (months)	Fracture L	Configuration	Weight Support	Time I Removal (days) *	Complications	Analgesia
1	SIB HUSK	M	27	15	U/R	TYPE IIB	60	63 (69)	PL	Y
2	RAT VAL	F	1,7	6	U/R	TYPE IIB 1X2	21	35 (35)	NO	N
3	CROSSBREED	F	5,5	12	T/F	TYPE IIB1.5X2	7	28 (34)	NO	N
4	CROSSBREED	F	5,5	12	U/R	TYPE IIB1.5X2	7	28 (31)	PL	N
- 5	AM STAFFORD	M	30	72	U/R	TYPE IIB 2X3	21	35 (36)	NO	Y
- 6	CROSSBREED	M	4,5	60	U/R	TYPE IIB 2X3	7	56 (58)	NO	Y
7	CROSSBREED	M	20	72	U/R	TYPE IIB 2X3	21	70 (75)	NO	Y
- 8	GRIFFON	M	15	50	U/R	TYPE IIB 2X3	28	42 (42)	NO	Y
9	GER SHEPH	F	34	70	T/F	TYPE IIB 2X3	21	42 (44)	NO	Y
10	W HIGH W TERR	F	5,6	48	U/R	TYPE IIB 2X3	21	42 (42)	GE	Y
11	CROSSBREED	M	18	48	U/R	TYPE IIB 2X3	21	42 (45)	NO	Υ
12	CROSSBREED	F	12	60	T/F	TYPE IIB 2X3	21	42 (45)	NO	Y
13	CROSSBREED	F	3	24	U/R	TYPE IIB 2X3	21	56 (57)	NO	Y
14	MALTESE	F	9	192	T/F	TYPE IIB 2X3	60	56 (60)	NO	Y
15	RAT VAL	F	5	111	T/F	TYPE IIB	21	63 (68)	NO	Y
16	BELG SHEPH	M	34	86	T/F	TYPE IIB	28	63 (65)	NO	N
17	BELG SHEPH	F	9	20	T/F	TYPE IIB 2X3	28	63 (64)	NO	Y
18	YORKSHIRE	F	1,5	35	U/R	TYPE IIB 2X3	21	35 (38)	PL	Y
19	POODLE	F	8	122	U/R	TYPE IIB 2X3	21	49 (50)	NO	Y
20	CROSSBREED	M	25	75	U/R	TYPE IIB 2X3	21	56 (60)	NO	Y
21	DALMATIAN	M	22	46	T/F	TYPE IIB 2X3	21	49 (52)	NO	Y
22	YORKSHIRE	F	1,5	24	U/R	TYPE IIB 2X3	60	56 (60)	NO	Y
23	CROSSBREED	M	4	55	U/R	TYPE IIB 2X3	28	56 (60)	NO	Y

^{*} The first number references the checking day when stage 4/5 was radiographically reached and the implant was ready for removal; the number in parenthesis refers to the day the implant was removed. SIB HUSK: Siberian Husky; RAT VAL: Ratonero Valenciano; AM STAFFORD: American Staffordshire Terrier; GER SHEPH: German Shepherd; W HIGH W TERR: West Highland White Terrier; BELG SHEPH: Belgian Shepherd.

Int. J. Mol. Sci. 2019, 20, 1075 5 of 10

No significant differences were found in the assessment of owner satisfaction at implant removal, with a satisfaction between 4 (24% in PRGF, 25% in SS) and 5 (76% in PRGF, 75% in SS).

Complications were recorded. One dog suffered gastroenteritis, and three dogs had pins become loose in the PRGF group. The same number of complications occurred in SS group (Tables 1 and 2).

3. Discussion

In the present study, the beneficial effect of PRGF in acute ulna/radius and tibia/fibula fracture healing has been proven, achieving a faster healing compared with controls. However, in all cases, a primary and non-complicated healing was present.

To the authors' knowledge, there is no published clinical research discussing the use of PRGF in fractures in a canine model. Experimentally, some studies proved there was faster bone regeneration when PRGF or other autologous platelet concentrates were applied [20,21]. In human medicine, there was only one clinical study evaluating the healing of fresh fractures using PRGF with no positive effect [18]. On the contrary, a clinical case with a delayed union fracture treated with autologous PRGF showed a favorable healing and concluded to be a safe technology for patients [19].

PRGF has also been used by other authors in combination with other therapeutics, showing positive results. Ya-dong Zhang et al. [22] proved that the use of PRGF combined with a degradable bioactive borate glass promotes functional bone repair. On the other hand, other authors [4] found no effect of PRGF on non-grafted implants in dogs; nevertheless, we cannot compare these results with our study because a different process was used to obtain the PRGF: using thrombin (100U/mL) to stimulate growth factor release rather than calcium chloride.

It is known that Carprofen is suitable alone or in combination with other NSAIDs for the control of pain and swelling in dogs [23,24]. Gastrointestinal inflammation and ulceration are among the most common side defects reported in the literature [25]. In our study, there were only two animals with gastroenteritis, and they responded positively to the conventional treatment.

In the present study, it has been observed that the surgical application of PRGF at the fracture site is associated with increased swelling and oedema during the first days, probably due to the activation of angiogenesis and cell activation [26]. The enhancement of the arrival and formation of blood vessels increases heat, pain, and redness of the area. This swelling associated with oedema has been effectively treated with oral Carprofen.

In any case, increased swelling did not affect the animals' gait nor the functional ability of the joint. In this sense, some papers reported the inhibitory effect of interleukins, which may be attributed to PRP [27]. This effect may be related to a reduction of acute pain in the fracture site, even though the activation of angiogenesis may cause an increased perception of discomfort and inflammation [26]. Thus, even if the application of PRGF increases oedema and swelling on the area, the limb's function was minimally affected during the first days.

Good results have been obtained using PRP to accelerate bone fusion [28]. In our case, the group receiving the PRGF injection presented an earlier implant removal, which is in agreement with those who state that chemotactic and mitogenic effect on mesenchymal cells (stem cells) and osteoblasts accelerate bone healing [29,30].

A rapid return to functionality is crucial for quick and correct healing; when the limb bears weight, a transmission of forces takes place that stimulates osteoinduction. Likewise, early activity boosts vascularization and avoids muscle atrophy, which are factors that clearly activate bone healing. Moreover, the Carprofen helped to control pain and acute swelling at the fracture site, facilitating an earlier return to functionality [29]. This shows that swelling control and post-surgical analgesia are fundamental for early functionality of the affected limb and represent an important parameter to be assessed by the pet owners.

Regarding external fixation, all the animals showed limb weight bearing 48 h after surgery. Very few complications arose in relation to the use of external skeletal fixation. One animal presented

Int. I. Mol. Sci. 2019, 20, 1075

a secondary infection, which is a usual side effect, and only six animals presented pin loosening. Other studies show a larger number of cases presenting pin loosening as the most frequent complication [31].

The present study has three main limitations. First, the use of dogs with a wide weight range potentially limited results that are more accurate. A narrow weight range could provide more reliable and accurate results, at least for a specific weight range. Second, a biomechanical analysis of gait could provide full objective results regarding limb function. Third, statistical analysis of the variable "swelling at the fracture site" could provide more accurate results if it is considered a continuous variable instead of categorical, avoiding detection, performance, and reporting biases; however, the presence of hematoma or callous formation at the fracture site could potentially hinder precise measurements.

4. Materials and Methods

A multicentric study was designed and formed by four surgeons in four different veterinary clinical centers.

4.1. Animal Model

A total of 68 dogs were evaluated. The follow-up of the animals took place until six months after treatment. The inclusion criteria required the presence of a fresh, single, closed fracture and the absence of significant muscular soft tissue damage or abrasions.

The exclusion criteria for the present study were the following:

- Animals presenting concurrent systemic disease (Leishmania spp., Ehrlichia spp., etc.).
- Animals with hematological disorders.
- Animals with multiple fractures.
- · Animals with internal lesions due to traumatism.
- Animals with open fractures or with significant damage to the surrounding soft-tissue.
- Animals with a significant weight loss or functional disabilities due to the treatment or other non-related causes.
- Animals needing different concurrent fixation methods due to the nature or clinical features of the fracture.

Fractures were classified according to the affected bone. In order to acquire similar healing conditions during the study, only tibia/fibula and radius/ulna fractures were included because of their poor vascularization due to their small surrounding muscular mass. The individual data of each dog for the PGRF and SS groups are summarized in Tables 1 and 2, respectively.

4.2. Fracture Treatment

All fractures were treated with conventional open or closed reduction and external fixation. The external skeletal fixation configuration frame was the most appropriate for each fracture, using type IIa or type IIb [32]. In all cases smooth pins of different diameters, connecting bars, and Meynard clamps were used.

After an initial clinical examination, animals were randomly assigned to one of the following groups depending on the treatment received:

- PRGF group: A single infiltration of PRGF in the fracture site during the surgery.
- SS group: A single infiltration of saline solution in the fracture site during the surgery.

All groups were treated with morphine (0.2 mg IM every 6 h), and Carprofen 4 mg/kg IV (Rimadyl[®], Zoetis[®], Spain) for 24 h. Cephalexin was administered as a post-surgery antibiotic.

After 24 h, the owners were allowed to give Carprofen (4 mg/kg/day) as a rescue analgesic if their pet presented clear signs of distress or discomfort. This fact should be reported during the clinical follow-up.

Int. J. Mol. Sci. 2019, 20, 1075 7 of 10

4.3. PRGF Preparation

For the present study, the extraction, isolation, activation, and administration model of the PRGF was standardized in all clinics following Anitua's technique [9]. Briefly, 20 mL of blood were aseptically collected in four 4.5 mL citrate tubes, then centrifuged during 8 min at 460 G. Care has to be taken to avoid the buffy coat. Before the infiltration, the PRGF was activated with 5% of its volume with 10% calcium chloride. This obtained PRP derivate is enriched in platelets 2-fold over peripheral blood and less than 0.2 leucocytes $\times\,10^6/\text{mL}.$

4.4. Evaluation

The limb function was evaluated on days 0 (pre-surgery), 7, 14, 21, 28, 60, 120, and 180 after the treatment began. This parameter was assessed by the same researcher evaluating animals when standing (1: weight-bearing; 2: no weight-bearing; or 3: no limb support), by observing swelling on the fracture site (0: presence or 1: absence), pain on palpation (0: presence or 1: absence), and joint movement (1: <40%; 2: 40-70%; 3: 70-90%; or 4: >90%).

The same radiologist, unaware of the group of treatment, patient, and surgeon involved, examined all radiographs. Each radiograph was evaluated by a stage score of 1–5 points (1: not visible callus formation; 2: barely visible callus formation; 3: scattered, not homogeneous callus; 4: uniform, mature callus formation; 5: very active, hyperthrophic callus formation). Radiographical examination started for each dog at day 21 and for every two weeks thereafter until the animal reached stage 2; beyond this period, radiographs were taken weekly, coinciding with the checkpoints for the other parameters. When a final score of 4/5 was achieved, implant removal was performed and recorded. The researcher who performed the evaluation of limbs and who read the radiographies were blind to the given treatment (PGRF or SS).

The use of the rescue analgesic and the presence of side effects were registered by the owner. The level of owner satisfaction with the clinical outcome of their pets during the first 28 days and at implant removal was evaluated with the following questionnaire referring to the level of satisfaction measured with a Likert-type scale (Table 3).

How do you	consider the	lameness of (na	me of the pet)	has progressed?
Excellent	Good	Average	Fair	Poor
5	4	3	2	1
Do you think	the treatmen	t given to (nam	e of the pet) ha	s been effective?
Strongly agree	Agree	Neutral	Disagree	Strongly disagree
How Do You	Think (Nam	e of the Pet) ha	s Responded to	the Treatment?
Excellent	Good	Average	Fair	Poor
5	4	3	2	1

Table 3. Likert-type questionnaire of satisfaction for dog owners at time of implant removal.

4.5. Statistical Analysis

Statistical analysis was performed with the computer program SPSS $18^{\$}$ for Windows (IBM Co., Chicago, IL, USA). A value of p < 0.05 was considered statistically significant. The descriptive study of the population was shown as the mean \pm SD. To determine the differences between the groups for non-categorical variables (weight, age, and total doses of Carprofen), a Kruskal–Wallis and Mann–Whitney test was done. To determine the effect of PRGF on implant removal time, a Kaplan–Meier curve and a log-rank test were used. The impact evaluation of total doses of Carprofen, age, weight, and bone fractured, within time to implant removal, a multivariable analysis was made using a Cox regression. Categorical variables (evaluation when walking, evaluation when standing, swelling, pain on palpation, joint movement, use of the recue analgesic, owner satisfaction, and

Int. J. Mol. Sci. 2019, 20, 1075 8 of 10

presence of side effects) were assessed using crosstabs with chi square, contingency coefficient, or the Fisher's exact test used when necessary in each variable.

The experimental procedure was approved by the ethics committee of the Research Institute in Biomedical and Health Sciences (ULPGC, Spain). The owners were informed about the aims of the study, and a written consent was required before including their pets in the study.

5. Conclusions

The use of PRGF for bone repair accelerates fracture consolidation and simultaneously promoted healing, achieving clearly shorter implant removal times.

Author Contributions: M.R., E.D., J.J.S., and J.M.C. performed and designed the experiment; S.L. and D.C. wrote the manuscript and participated in performing the experiment; M.R. performed statistical analysis; B.C. and S.L. participated in performing the experiment; and J.M.V. proofread the manuscript and gave approval of the final version.

Acknowledgments: The authors would like to acknowledge the pet owners for their cooperation in this study. Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

PRGF Plasma rich in growth factors

SS Saline solution
Fracture L Fracture location
U/R Ulna/radius
T/F Tibia/fibula

Time I removal Time for implant removal

GE Gastroenteritis
PL Pin loosening
Y Yes
N No

References

- Anitua, E. The use of plasma-rich growth factors (PRGF) in oral surgery. Pract. Proc. Aesth. Dent. 2001, 13, 487–493.
- Anitua, E.A. Enhancement of osseointegration by generating a dynamic implant surface. J. Oral Implantol. 2006, 32, 72–76. [CrossRef] [PubMed]
- Taschieri, S.; Rosano, G.; Weinstein, T.; Bortolin, M.; Del Fabbro, M. Treatment of through-and-through bone lesion using autologous growth factors and xenogeneic bone graft: A case report. Oral Maxillofac. Surg. 2012, 16, 57–64. [CrossRef] [PubMed]
- Vilar, J.M.; Morales, M.; Santana, A.; Spinella, G.; Rubio, M.; Cuervo, B.; Cugat, R.; Carrillo, J.M. Controlled, blinded force platform analysis of the effect of intraarticular injection of autologous adipose-derived mesenchymal stem cells associated to PRGF-Endoret in osteoarthritic dogs. BMC Vet. Res. 2013, 9, 131. [CrossRef] [PubMed]
- Jensen, T.B.; Bechtold, J.E.; Chen, X.; Vestermark, M.; Soballe, K. No effect of autologous growth factors (AGF) around ungrafted loaded implants in dogs. Int. Orthop. 2010, 34, 925–930. [CrossRef] [PubMed]
- Li, N.Y.; Chen, L.Q.; Chen, T.; Jin, X.M.; Yuan, R.T. Effect of platelet-rich plasma and latissimus dorsi
 myofascia with blood vessel on vascularization of tissue engineered bone in dogs. *Hua Xi Kou Qiang Yi Xue*Za Zhi 2007, 25, 408–411. [PubMed]
- Thor, A.L.; Hong, J.; Kjeller, G.; Sennerby, L.; Rasmusson, L. Correlation of platelet growth factor release in jawbone defect repair—A study in the dog mandible. Clin. Implant Dent. Relat. Res. 2013, 15, 759–768. [CrossRef] [PubMed]
- Marx, R.E.; Carlson, E.R.; Eichstaedt, R.M.; Schimmele, S.R.; Strauss, J.E.; Georgeff, K.R. Platelet-rich plasma: Growth factor enhancement for bone grafts. Oral Surg. Oral Med. Oral Pathol. Oral Radiol. 1998, 85, 638–646.

Int. J. Mol. Sci. 2019, 20, 1075 9 of 10

Anitua, E.; Sanchez, M.; Zalduendo, M.M.; de la Fuente, M.; Prado, R.; Orive, G.; Andia, I. Fibroblastic response to treatment with different preparations rich in growth factors. Cell Prolif. 2009, 42, 162–170. [CrossRef] [PubMed]

- Marx, R.E.; Garg, A.K. The biology of platelets and the mechanism of platelet-rich plasma. In *Dental and Craniofacial Applications of Platelet-Rich Plasma*; Marx, R.E., Garg, A.K., Eds.; Quintessence Publishing Co.: Chicago, IL, USA, 2005; pp. 3–65.
- Mishra, A.; Pavelko, T. Treatment of chronic elbow tendinosis with buffered platelet-rich plasma. Am. J. Sports Med. 2006, 34, 1774–1778. [CrossRef] [PubMed]
- Anitua, E. Plasma rich in growth factors. Preliminary results of use in the preparation of future sites for implants. Int. J. Oral Maxillofac. Implants 1999, 14, 529–535. [PubMed]
- Serra, C.I.; Soler, C.; Carrillo, J.M.; Sopena, J.J.; Redondo, J.I.; Cugat, R. Effect of autologous platelet-rich plasma on the repair of full-thickness articular defects in rabbits. *Knee Surg. Sports Traumatol. Arthrosc.* 2013, 21, 1730–1736. [PubMed]
- Cugat, R.; Alentorn-Geli, E.; Steinbacher, G.; Alvarez-Diaz, P.; Cusco, X.; Seijas, R.; Barastegui, D.; Navarro, J.; Laiz, P.; Garcia-Balletbo, M. Treatment of Knee Osteochondral Lesions Using a Novel Clot of Autologous Plasma Rich in Growth Factors Mixed with Healthy Hyaline Cartilage Chips and Intra-Articular Injection of PRGF. Case rep. Orthop. 2017, 2017, 8284548. [CrossRef] [PubMed]
- Nicoletti, G.; Saler, M.; Villani, L.; Rumolo, A.; Tresoldi, M.M.; Faga, A. Platelet Rich Plasma Enhancement of Skin Regeneration in an ex-vivo Human Experimental Model. Front. Bioeng. Biotechnol. 2019, 7, 2. [CrossRef] [PubMed]
- Yang, H.S.; Shin, J.; Bhang, S.H.; Shin, J.Y.; Park, J.; Im, G.I.; Kim, C.S.; Kim, B.S. Enhanced skin wound healing by a sustained release of growth factors contained in platelet-rich plasma. *Exp. Mol. Med.* 2011, 43, 622–629. [CrossRef] [PubMed]
- Cieslik-Bielecka, A.; Choukroun, J.; Odin, G.; Dohan Ehrenfest, D.M. L-PRP/L-PRF in esthetic plastic surgery, regenerative medicine of the skin and chronic wounds. Curr. Pharm. Biotechnol. 2012, 13, 1266–1277. [CrossRef] [PubMed]
- Zhang, C.Q.; Yuan, T.; Zeng, B.F. Experimental study on effect of platelet-rich plasma in repair of bone defect. Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi 2003, 17, 355–358. [PubMed]
- Seijas, R.; Santana-Suarez, R.Y.; Garcia-Balletbo, M.; Cusco, X.; Ares, O.; Cugat, R. Delayed union of the clavicle treated with plasma rich in growth factors. Acta Orthop. Belgica 2010, 76, 689–693.
- Molina-Minano, F.; Lopez-Jornet, P.; Camacho-Alonso, F.; Vicente-Ortega, V. Plasma rich in growth factors and bone formation: A radiological and histomorphometric study in New Zealand rabbits. *Braz. Oral Res.* 2009, 23, 275–280. [CrossRef] [PubMed]
- Marcazzan, S.; Taschieri, S.; Weinstein, R.L.; Del Fabbro, M. Efficacy of platelet concentrates in bone healing: A systematic review on animal studies—Part B: Large-size animal models. *Platelets* 2018, 29, 338–346. [CrossRef] [PubMed]
- Zhang, Y.D.; Wang, G.; Sun, Y.; Zhang, C.Q. Combination of platelet-rich plasma with degradable bioactive borate glass for segmental bone defect repair. Acta Orthop. Belgica 2011, 77, 110–115.
- Karrasch, N.M.; Lerche, P.; Aarnes, T.K.; Gardner, H.L.; London, C.A. The effects of preoperative oral administration of carprofen or tramadol on postoperative analgesia in dogs undergoing cutaneous tumor removal. Can. Vet. J. 2015, 56, 817–822. [PubMed]
- Kalchofner Guerrero, K.S.; Schwarz, A.; Wuhrmann, R.; Feldmann, S.; Hartnack, S.; Bettschart-Wolfensberger, R. Comparison of a new metamizole formulation and carprofen for extended post-operative analgesia in dogs undergoing ovariohysterectomy. Vet. J. 2015, 204, 99–104. [CrossRef] [PubMed]
- Khan, S.A.; McLean, M.K. Toxicology of frequently encountered nonsteroidal anti-inflammatory drugs in dogs and cats. Vet. Clin. N. Am. Small Anim. Pract. 2012, 42, 289–306. [CrossRef] [PubMed]
- Martinez, C.E.; Smith, P.C.; Palma Alvarado, V.A. The influence of platelet-derived products on angiogenesis and tissue repair: A concise update. Front. Physiol. 2015, 6, 290. [PubMed]
- Tong, S.; Liu, J.; Zhang, C. Platelet-rich plasma inhibits inflammatory factors and represses rheumatoid fibroblast-like synoviocytes in rheumatoid arthritis. Clin. Exp. Med. 2017, 17, 441–449. [CrossRef] [PubMed]
- Kubota, G.; Kamoda, H.; Orita, S.; Inage, K.; Ito, M.; Yamashita, M.; Furuya, T.; Akazawa, T.; Shiga, Y.;
 Ohtori, S. Efficacy of Platelet-Rich Plasma for Bone Fusion in Transforaminal Lumbar Interbody Fusion.
 Asian Spine J. 2018, 12, 112–118. [CrossRef] [PubMed]

Int. J. Mol. Sci. 2019, 20, 1075

29. Holtsinger, R.H.; Parker, R.B.; Beale, B.S.; FRIEDMAN, R.L. The therapeutic efficacy of carprofen (Rimadyl-V[™]) in 209 clinical cases of canine degenerative joint disease. *Vet. Comp. Orthop. Traumatol.* **1992**, *5*, 140–144.

- Mehta, S.; Watson, J.T. Platelet rich concentrate: Basic science and current clinical applications. J. Orthop. Trauma 2008, 22, 432–438. [CrossRef] [PubMed]
- Gemmill, T.J.; Cave, T.A.; Clements, D.N.; Clarke, S.P.; Bennett, D.; Carmichael, S. Treatment of canine and feline diaphyseal radial and tibial fractures with low-stiffness external skeletal fixation. *J. Small Anim. Pract.* 2004, 45, 85–91. [PubMed]
- Piermattei, D.L.; Flo, G.L.; De Camp, C.E. Fractures: Classification, Diagnoses and Treatment. In Brinker, Piermattei and Flo's Handbook of Small Animal Orthopaedics and Fracture Repair; Piermattei, D.L., Flo, G.L., DeCamp, C.E., Eds.; Inter-Médica: Buenos Aires, Argentina, 2006; pp. 25–159.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).

TRATAMIENTO	OA	Y	ARTÍCULOS	

	O 4	T 7	A DOTÉC		
TRATAMIENTO	()A	Υ	ARTIC	IULOS	



4.2.2.2. Seguridad del PRP

Dentro de los efectos beneficiosos que produce el PRP, una parte significativa de ellos es atribuible a uno de los factores de crecimiento, en concreto al IGF-1. Este mediador de la hormona del crecimiento interviene de forma relevante en los procesos anabólicos del organismo (Philippou y cols, 2007). Teniendo en cuenta lo anterior, potencialmente podría producir un efecto ergogénico que, en teoría, si no estuviera conveniente regulado, podría desencadenar un proceso canceroso (Grimberg y Cohen, 2000; Grimberg, 2003; Renehan y cols, 2004; Lim y cols, 2015; Maniscalco y cols, 2015). Su efecto, tanto desde el punto de vista carcinogénico como sobre todo el ergogénico es objeto de discusión en la actualidad y, de hecho, este factor está incluido por este motivo entre la lista de sustancias prohibidas de la World Anti-Doping Agency (WADA).

Por lo tanto, se puede deducir que altos niveles de IGF-1 circulante serían los causantes del afecto a distancia, es decir, general, de los productos que contengan esta molécula como en este caso el PRP, utilizado con fines y a dosis terapéuticas. En este sentido, conviene destacar el trabajo de Vilar y cols (2017) donde se concluye que dosis terapéuticas de PRP no conllevan un aumento significativo de IGF-1 en sangre y, por lo tanto, es un producto seguro.

¿Y a nivel local? Este punto adquiere una enorme importancia ya que la mayoría de los tratamientos de PRP, especialmente aquellos que se usan en el aparato locomotor, se administran de forma parenteral. Por lo tanto, planteamos el siguiente estudio:

	O 4	T 7	A DOTÉC		
TRATAMIENTO	()A	Υ	ARTIC	IULOS	



Damiá E, Chicharro D, Rubio M, Carrillo JM, Sopena J, Cuervo B, López S, Vilar JM. Can Plasma Rich in Growth Factors Be Safe for Parental Use? A Safety Study in the Canine Model.

Int J Mol Sci. 2018 Sep 11;19(9). doi: 10.3390/ijms19092701.

JCR Impact factor (2017): 3.687

Cuartil: Q2

Grupo: Chemistry, multidisciplinary

Posición: 52/171

TRATAMIENTO				
TRATAMIENTO	OA	Y	ARTÍCULOS	





Article

Can Plasma Rich in Growth Factors Be Safe for Parental Use? A Safety Study in the Canine Model

Elena Damiá ¹, Deborah Chicharro ¹, Mónica Rubio ^{1,2}, José María Carrillo ^{1,2}, Joaquín Sopena ^{1,2}, Belén Cuervo ¹, Sergio López ³ and José Manuel Vilar ^{3,*}

- Bioregenerative Medicine and Applied Surgery Research Group, Department of Animal Medicine and Surgery, Faculty of Veterinary, Universidad Cardenal Herrera-CEU, CEU Universities, 46115 Valencia, Spain; elena.damia@uchceu.es (E.D.); debora.chicharro@uchceu.es (D.C.); mrubio@uchceu.es (M.R.); jcarrill@uchceu.es (J.M.C.); jsopena@uchceu.es (J.S.); belen.cuervo@uchceu.es (B.C.)
- García Cugat Foundation CEU-UCH Chair of Medicine and Regenerative Surgery, 08006 Barcelona, Spain
- ³ Department of Animal Pathology, Instituto Universitario de Investigaciones Biomédicas y Universitarias, Universidad de Las Palmas de Gran Canaria, 35416 Trasmontaña S/N. Arucas, Spain; sergiolopezbarbeta@gmail.com
- * Correspondence: jose.vilar@ulpgc.es; Tel.: +34-928-457-244

Received: 2 August 2018; Accepted: 10 September 2018; Published: 11 September 2018



Abstract: Low invasiveness is the main goal of modern surgery. The use of platelet-rich plasma (PRP) is known to be effective in a variety of applications, such as oral, maxillofacial, orthopedic, dermatologic and cosmetic surgeries. However, a potential ergogenic and carcinogenic effect of PRP derivatives by means of the insulin-like growth factor-1 (IGF-1) pathway has been suggested. Because of this notion, the purpose of this study is to assess the effect of a commercially available PRP-derivative intramuscular injection in the lumbar muscular tissue (local effect) and to determine the IGF-1 blood concentration (systemic effect) on healthy beagle dogs. Local effect was evaluated by computed tomography (CT) scan and echography, and systemic effect was calculated by blood testing on days 0, 14, 28, 42 and 56. No statistically significant changes were observed; thus, PRGF could be considered safe when using therapeutic doses.

Keywords: platelet-rich plasma (PRP); plasma rich in growth factors (PRGF); insulin-like growth factor-1 (IGF-1); canine

1. Introduction

The main goal of modern surgery is to reduce invasiveness and increase the healing process. Regenerative medicine is now one of the most attractive and interesting disciplines that aims to regenerate or repair damaged tissues [1]. Platelet-rich plasma (PRP) is currently used in different medical fields and involves a minimum risk of immune reactions and transmission of diseases [2]. The first descriptions of the development and use of PRP were in the early 1970s in the hematology field, followed by maxillofacial and oral surgery [3]. Subsequently, PRP has been used in a wide variety of disciplines, such as aesthetic dermatology [4], including alopecia [5] and skin rejuvenation [6]; the musculoskeletal field [7]; oral and maxillofacial surgery [8]; and ophthalmology [9].

In recent years, the use of regenerative therapies, such as plasma rich in growth factors (PRGF), a PRP derivative, is also gaining interest to promote healing in muscle injuries, and consequently, to enable the patient to resume daily and sports-related activities quickly without relapse. A considerable number of authors have reported that growth factors (GFs) and fibrin matrix are crucial for the muscle repair and regeneration process by promoting myogenesis, angiogenesis and fibrogenesis [10,11], and promising results have been proven with this novel biological approach in managing musculoskeletal pathologies [12].

Int. J. Mol. Sci. 2018, 19, 2701; doi:10.3390/ijms19092701

www.mdpi.com/journal/ijms

Int. J. Mol. Sci. 2018, 19, 2701

Besides the possible beneficial effects of PRP derivatives, several concerns have been raised regarding undesirable side effects. Some authors describe a potential carcinogenetic effect related to the insulin-like growth factor-1 in humans (IGF-1) [13–16]. In veterinary medicine, some studies demonstrated IGF-1 receptor expression and its role in canine osteosarcoma [17] and mammary gland carcinoma [18]. In this sense, there are different IGF-1 isoforms, such as IGF-1Ea, IGF-1Eb and IGF-1Ec. The IGF-1Ec isoform has an important role in physiology and cancer biology through its Ec peptide. After the tissue is damaged from mechanical stimuli, IGF-1Ec isoform and the Ec peptide levels are induced in the muscle, tendon and bone, and its secretion produces cellular proliferation [18–20].

Multiple GFs are secreted during muscle repair and hypertrophy, but only IGF-1 and its isoforms participate in muscle proliferation, differentiation and regeneration [21]. It has been demonstrated that IGF-1, at the onset of the mechanical stress on human skeletal muscle cells, increases IGF-1Ec isoform [22]. Moreover, the application of the Ec peptide in a rat model provided an increase in the expression of myofibroblasts in wound healing [23]. Overall, published scientific research supports that GFs included in PRP are unlikely to trigger a potent ergogenic effect. Regarding IGF-1, the doses in PRP are subtherapeutic, only 1% of the total IGF-1 is biologically available. IGF-1 in plasma has a low half-life (20 h) in humans [24], and although we could not find published data, dogs should be similar. To demonstrate this controversy, the use of PRP intramuscular injections in athletes was prohibited by the World Anti-Doping Agency (WADA) in 2010, despite the use of these biological therapies as the "gold standard" for muscle injury treatment, but again permitted in 2011 due to the limited evidence for systemic ergogenic effect of PRP. Nevertheless, the use of individual GFs in athletes continues to be prohibited under Section S2 of the 2018 WADA Prohibited List [25], particularly fibroblast growth factor (FGF), hepatocyte growth factor (HGF), insulin-like growth factor (IGF-1), platelet-derived growth factor (PDGF) and vascular endothelial growth factor (VEGF), because of concerns regarding their abuse as ergogenic substances [26].

For these reasons, along with the multiple applications of these therapies and several pieces of evidence for specific ergogenic (local) and carcinogenic (systemic) effects, the use of PRGF in muscle tissue remains of great interest.

Based on this, the purpose of our study was to evaluate (a) the local effect, measuring the cross-sectional area (CSA) of the lumbar muscles by using imaging systems, and (b) the systemic effect by blood IGF-1 determination in healthy beagle dogs, which were submitted to a PRGF intramuscular lumbar injection.

2. Results

2.1. Animals

The ages of the animals included in the study were (mean \pm SD) 73.5 \pm 38.8 months in the control group, 76.5 \pm 38.7 months in the PRGF group, and 79.4 \pm 38.9 months in the triple dose (HPRGF) group. Their weights were 16.9 \pm 3.3 kg in the control group, 14.8 \pm 3.1 kg in the PRGF-treated dogs, and 14.9 \pm 2.5 kg in the HPRGF group.

2.2. IGF-1 Evaluation

No statistically significant differences were observed along the studied times nor between groups in IGF-1 serum concentrations. As a result, IGF-1 serum concentrations remained stable throughout the study, showing the inability of PRGF intramuscular injection to have a systemic effect (Table 1).

Int. J. Mol. Sci. 2018, 19, 2701 3 of 10

Table 1. IGF-1 measurements (ng/mL) in the three study groups.

Time	Group	Mean	SD
	Control	166.3	31.1
Baseline	PRGF	132.7	49.1
	HPRGF	95.6	31.5
	Control	174.7	25.1
14 days	PRGF	114.5	40.2
-	HPRGF	117.1	30.4
	Control	157.9	26.4
28 days	PRGF	133.8	46.7
,	HPRGF	127.5	34.0
	Control	134.6	20.0
42 days	PRGF	143.9	48.5
	HPRGF	126.2	33.1
	Control	155.6	24.3
56 days	PRGF	139.4	49.9
	HPRGF	117.2	38.5

IGF-1: Insuline Growth Factor-1; PRGF: single dose of Plasma Rich in Growth Factors; HPRGF: triple dose of Plasma Rich in Growth Factors.

Regarding external factors affecting IGF-1 serum concentrations, in the three studied groups, weight influenced IGF-1 serum concentrations. In this way, the control group showed higher serum concentrations due to a larger weight in the animals during this phase. Conversely, older animals had lower IGF-1 serum concentrations compared to younger animals.

2.3. Computed Tomography and Echography Evaluation

Both CT-scan and echography images were carried out between the three studied groups and along the studied times. A correlation test was also realized between the two measures. No statistically significant differences were obtained between groups nor along the studied times in the studied anatomic level (L5) (Tables 2–4). As a result, no local effect and, therefore, no muscular hypertrophy were observed after PRGF injection.

Table 2. Ultrasound measurements of the muscular area at left L5 level in the three study groups.

Time	Group	Mean	SD
	Control	12.8	3.6
Baseline	PRGF	12.5	3.1
	HPRGF	12.8	3.5
	Control	13.4	3.3
14 days	PRGF	12.6	3.0
15.0	HPRGF	13.1	3.3
	Control	13.1	3.2
28 days	PRGF	12.7	3.0
,	HPRGF	13.3	3.1
	Control	13.0	3.1
42 days	PRGF	12.6	3.1
100	HPRGF	13.0	3.1
	Control	13.0	3.1
56 days	PRGF	12.4	3.1
	HPRGF	12.8	3.2

L5: fifth lumbar vertebra.

Int. J. Mol. Sci. 2018, 19, 2701 4 of 10

Table 3. CT-scan measurements of the muscular area at left L5 level in the three study groups.

Time	Group	Mean	SD
	Control	13.3	2.6
Baseline	PRGF	12.7	2.4
	HPRGF	12.8	2.9
	Control	12.9	2.7
14 days	PRGF	12.2	2.5
<i>(A)</i>	HPRGF	13.2	2.8
	Control	12.5	2.5
28 days	PRGF	12.3	3.3
	HPRGF	12.8	2.6
	Control	12.4	2.5
42 days	PRGF	12.1	2.7
,	HPRGF	13.1	3.1
	Control	12.9	2.6
56 days	PRGF	11.8	2.4
	HPRGF	12.4	3.1

Table 4. Pearson correlation between ultrasound and CT-scan measurements.

Correlations					
	L5 US	L5 CT scan			
Pearson correlation	1	0.928 *			
Sig. (2-tailed)		0.000			
N	210	105			
Pearson correlation	0.928 *	1			
Sig. (2-tailed)	0.000				
N	105	105			
	Pearson correlation Sig. (2-tailed) N Pearson correlation	Pearson correlation 1 Sig. (2-tailed) N 210 Pearson correlation 0.928 * Sig. (2-tailed) 0.000			

^{*} Correlation is significant at the 0.01 level (2-tailed).

3. Discussion

The aim of the present study was to determine if an intramuscular injection of PRGF increases circulating levels of the potentially ergogenic growth factor IGF-1 and thus induces skeletal muscle hypertrophy and, in the last instance, cancer.

Assuming that injections of PRGF within the injured muscle enhance healing and functional recovery [27], the question remains as to what is the correct dosage. In humans, Hamilton et al. (2010) demonstrated that a single injection of 3 mL PRP was effective for grade II semimembranosus strain injury, with a full recovery after 17 days post-injection [28]. Moreover, Hamid et al. (2012) used the same dose after a grade II hamstring injury, and the time needed to return to play in participants was 16 weeks [29]. In this sense, we decided to use a single injection of 1 mL of PRGF (normal-dose PRGF; PRGF group) and 3 mL of PRGF (high-dose PRGF; HPRGF group), taking into account the differences in size and weight between humans and dogs.

Particular attention is drawn to IGF-1 due to its potential ergogenic [14] and carcinogenetic effects [27]. In our study, a single intramuscular PRGF injection in healthy beagle dogs has no effect on circulating IGF-1 values, even when the standard PRGF concentration was increased three-fold. In reference to this systemic anabolic action of PRGF and in concordance with our results, several studies have also demonstrated that different commercial PRP systems do not increase IGF-1 concentrations over normal circulated blood levels [28–30]. Moreover, further scientific research supports the opinion that PRP is unlikely to promote an ergogenic effect in patients. This is due to subtherapeutic doses of IGF-1 in PRP. The isoform of IGF-1Ec in PRP is the isoform that causes muscular hypertrophy [31]. The unbound IGF-1 has too-short a half-life to exert systemic effects, and only 1% of IGF-1 is biologically available and active [32].

Int. J. Mol. Sci. 2018, 19, 2701 5 of 10

The last undesirable effect of IGF-1 suggested by other authors is a potential carcinogenic effect in humans [16,33] and in veterinary medicine [17,18]. Some authors [31,34] have suggested that growth factors, acting only on cell surface receptors, do not access the cell and do not promote cell DNA mutation. In agreement with Schippinger et al. [19], in our study neither the PRGF or HPRGF intramuscular injection showed an increase in IGF-1 serum concentrations. This suggests that PRGF application can be considered a safe method of treatment after 14 days, 28 days, 42 days and 56 days post-injection. Although long-term effects of multiple injections of PRGF were not examined in our study, the HPRGF used in one of the groups contained three-times the dose of normal PRGF, and no statistically significant differences were shown, suggesting that several applications over time would not alter IGF-1 circulating levels [35].

In reference to the effect of weight on serum IGF-1 concentrations, a positive correlation was observed [36], where animals from the control group had higher IGF-1 levels due to a greater weight. In the same way, high IGF-1 concentrations have been shown in obese dogs, which return to normal levels after weight loss [37]. Moreover, regarding the influence of age on IGF-1 circulating levels, with the exception of the control group, older animals show lower systemic IGF-1 concentrations. Moreover, our results are in agreement with other studies in humans [38] and veterinary medicine [39].

To assess the evolution of muscular fiber size after a PRGF injection, an imaging study was carried out with infiltrated lumbar muscles by echography and CT scan. No statistically significant differences were found between groups regarding muscle area measurements. As a result, the muscle size was similar in both infiltrated areas after intramuscular PRGF, HPRGF or saline solution, showing that intramuscular PRGF does not exert an anabolic effect even when injecting high doses.

4. Materials and Methods

4.1. Animal Model

A total of 24 healthy adult Beagle dogs were used in this study and were divided into three groups of eight dogs, five males and three females in each group, with ages ranging from 3-4 years and weights from 10-18 kg. Complete physical examination, haematology, and serum biochemical analyses were performed to ensure that animals were healthy.

The study protocol was approved by the Ethics Committee for Animal Welfare at the University CEU-Cardenal Herrera of Valencia (CEBA/2013).

4.2. Plasma Rich in Growth Factors (PRGF) Preparation and Infiltration

PRGF®-Endoret® technology (BTI Biotechnology Institute, Álava, Spain) was followed to obtain an autologous preparation of PRP [29]. Briefly, blood was collected from the external jugular vein of each dog under sterile conditions in Vacutainer sodium citrate 3.8% tubes (Blood-Collecting Tubes®, BTI Biotechnology Institute, Álava, Spain). The tubes were centrifuged at $460 \times g$ for eight minutes (PRGF® System III, Biotechnology Institute, Álava, Spain) to separate the different blood phases. The fraction located immediately above the buffy coat (white fraction) corresponded to PRGF, which was activated by adding 5% of calcium chloride (CaCl₂ 10%) just before infiltration to activate platelets for GF release.

After obtaining PRGF, the platelet concentrations and the presence of leukocytes between whole blood, PRGF, and plasma poor in growth factors (PPGF) were compared on the initial day of each of the 3 study groups. Regarding the concentration of platelets, in the 3 study groups, the authors observed an increase in the number of platelets between the blood, the PPGF, and the PRGF, showing PRGF platelet values of 1.5–2-times higher than the concentration in blood and PPGF, according to what has been previously described [29]. With regard to the concentration of leukocytes, there are statistically significant differences between blood, PRGF, and PPGF in the three groups of the study. These results confirm the absence of white blood cells after the centrifugation of the blood and the separation of the different types of plasma. These results coincide with a previous report [30], which defends the absence of leukocytes in the PRGF.

Int. J. Mol. Sci. 2018, 19, 2701 6 of 10

Every dog was injected in the left lumbar muscles (lumbar multifidus, latissimus dorsi lumbar, and iliocostal lumbar muscles) at the 5th lumbar vertebrae level with the following treatments:

- Treatment 1: single dose of 1 mL sterile saline solution activated with 0.05 mL CaCl₂ 10% (control group) [40].
- Treatment 2: single dose of 1 mL PRGF activated with 0.05 mL CaCl₂ 10% (PRGF group).
- Treatment 3: single dose of 3 mL PRGF activated with 0.15 mL CaCl₂ 10% (HPRGF group).

4.3. Determination of IGF-1 Concentrations

Under sterile conditions, blood samples were collected from the external jugular vein after intramuscular sedation with medetomidine (0.01 mg/kg), morphine (0.2 mg/kg), and midazolam (0.2 mg/kg). Samples were obtained at baseline, and 14 days, 28 days, 42 days and 56 days after injection of intramuscular PRGF. IGF-1 was analyzed by automated immunoassay system (Immulite 1000 IGF-1 assay; Diagnostic Products, Los Angeles, CA, USA) previously validated in dogs [41].

To evaluate the local effect of the intramuscular PRGF injections, ultrasound and CT-scan studies were performed.

4.4. Muscle Tissue Evaluation by Echography

Following the previous suggestions by other authors that echography has equal sensitivity to MRI for acute muscle injury (hamstring muscle) especially when performed within 2 weeks following injury [42], the ultrasound study was performed for each group (Control, PRGF, HPRGF) at baseline, and 14 days, 28 days, 42 days and 56 days after injection of the corresponding treatments.

The ultrasound images (Esaote mylab60, Genoa, Italy) were taken at left L5 level (midpoint 5th lumbar vertebrae) to calculate the muscular area average (Figure 1).

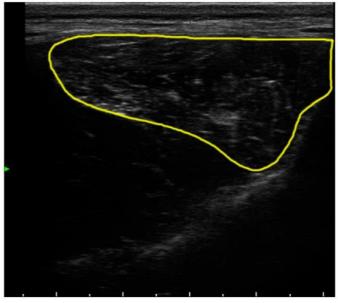


Figure 1. Ultrasonogram of one dog. The approximate contour of the measured lumbar area is delineated in yellow.

Int. J. Mol. Sci. 2018, 19, 2701 7 of 10

4.5. Muscle Tissue Evaluation by Computed Tomography

A CT scan (CT-max, General Electric, Madrid, Spain) was performed every 14 days within the study; therefore, measurements were taken at baseline, 14, 28, 42, and 56 days (i.e., the same as ultrasound examination) under sedation with medetomidine (0.01 mg/kg), morphine (0.2 mg/kg), and midazolam (0.2 mg/kg).

CT-scan images were performed at the same anatomic level as the ultrasound study, and three corresponding measurements were taken from lumbar muscles at left side (Figure 2).

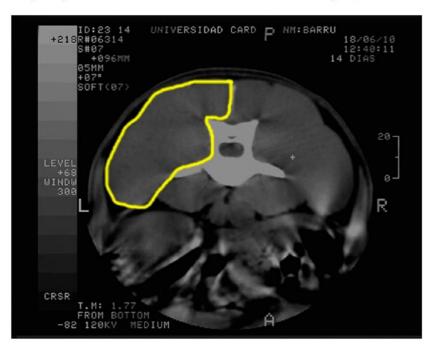


Figure 2. CT scan of one dog. The approximate contour of the measured lumbar area is delineated in yellow.

4.6. Image Processing

Once the ultrasound and CT-scan images were collected, the contours were traced. Measurements from the designated muscular lumbar area were determined via quantitative morphometry using Image Pro Plus software (for Windows 2000, Silver Spring, MD, USA). The median value of the three measurements was considered as long as the measurements differed <10%. When the difference was >10%, new measurements were obtained.

4.7. Statistical Analysis

The data were processed using the SPSS 15.0 for Windows (Chicago, IL, USA). A descriptive study of the mean, standard deviation, and confidence intervals was made for each variable. A value of $p \leq 0.05$ was considered significant. The result of each parameter was evaluated with a nonparametric Kolgomorov–Smirnov test for normality and log transformed if necessary. ANOVA repeated-measures and post-hoc Tukey tests were performed to assess differences with the baseline. A one-way ANOVA

Int. J. Mol. Sci. 2018, 19, 2701 8 of 10

was conducted each time, to assess differences between groups, and a post-hoc Tukey test was carried out when necessary. A Pearson correlation between echography and CT-scan measures was obtained.

5. Conclusions

A single intramuscular application of PRGF does not significantly increase systemic IGF-1 levels nor increase muscle mass, even when three-times the normal dose in canine species was used. Therefore, in the canine species, a single application of PRGF is safe for parental use with respect to local and systemic IGF-1 levels and cancer risk. Despite this, further studies are needed to prove and evaluate the safety of this therapy in humans.

Author Contributions: M.R., E.D., J.S. and J.M.C. performed the experiment and designed it; D.C. and E.D. wrote the manuscript and participated in performing the experiment; M.R. performed statistical analysis; B.C. and S.L. participated in performing the experiment; J.M.V. proofread and gave final approval of the manuscript.

Funding: This research received no external funding.

Acknowledgments: We would like to acknowledge the García Cugat Foundation for biomedical research.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

PRGF plasma rich in growth factors
HPRGF high-dose plasma rich in growth factors
PRP platelet-rich plasma
IGF-1 insulin-like growth factor-1
CRP C-reactive protein

CRP C-reactive protein
WADA World Anti-Doping Agency
VEGF vascular endothelial growth factor
FGF fibroblast growth factor

FGF fibroblast growth factor
HGF hepatocyte growth factor
PDGF platelet-derived growth factor

References

- Giannini, S.; Cielo, A.; Bonanome, L.; Rastelli, C.; Derla, C.; Corpaci, F.; Falisi, G. Comparison between PRP, PRGF and PRF: Lights and shadows in three similar but different protocols. Eur. Rev. Med. Pharmacol. Sci. 2015, 19, 927–930. [PubMed]
- Marques, L.F.; Stessuk, T.; Camargo, I.C.; Sabeh Junior, N.; dos Santos, L.; Ribeiro-Paes, J.T. Platelet-rich
 plasma (PRP): Methodological aspects and clinical applications. *Platelets* 2015, 26, 101–113. [CrossRef]
 [PubMed]
- Andia, I.; Abate, M. Platelet-rich plasma: Underlying biology and clinical correlates. Reg. Med. 2013, 8, 645–658. [CrossRef] [PubMed]
- Leo, M.S.; Kumar, A.S.; Kirit, R.; Konathan, R.; Sivamani, R.K. Systematic review of the use of platelet-rich plasma in aesthetic dermatology. J. Cosmet. Dermatol. 2015, 14, 315–323. [CrossRef] [PubMed]
- Chen, J.X.; Justicz, N.; Lee, L.N. Platelet-Rich Plasma for the Treatment of Androgenic Alopecia: A Systematic Review. Facial Plast. Surg. 2018. [CrossRef] [PubMed]
- Ulusal, B.G. Platelet-rich plasma and hyaluronic acid-an efficient biostimulation method for face rejuvenation.
 J. Cosmet. Dermatol. 2017, 16, 112–119. [CrossRef] [PubMed]
- Mlynarek, R.A.; Kuhn, A.W.; Bedi, A. Platelet-Rich Plasma (PRP) in Orthopedic Sports Medicine. Am. J. Orthop. (Belle Mead NJ) 2016, 45, 290–326. [PubMed]
- Del Corso, M.; Vervelle, A.; Simonpieri, A.; Jimbo, R.; Inchingolo, F.; Sammartino, G.; Dohan Ehrenfest, D.M.
 Current knowledge and perspectives for the use of platelet-rich plasma (PRP) and platelet-rich fibrin (PRF)
 in oral and maxillofacial surgery part 1: Periodontal and dentoalveolar surgery. Curr. Pharm. Biotechnol.
 2012, 13, 1207–1230. [CrossRef] [PubMed]
- Alio, J.L.; Rodriguez, A.E.; Abdelghany, A.A.; Oliveira, R.F. Autologous Platelet-Rich Plasma Eye Drops for the Treatment of Post-LASIK Chronic Ocular Surface Syndrome. J. Ophthalmol. 2017. [CrossRef] [PubMed]

Int. J. Mol. Sci. 2018, 19, 2701 9 of 10

 Borselli, C.; Storrie, H.; Benesch-Lee, F.; Shvartsman, D.; Cezar, C.; Lichtman, J.W.; Vandenburgh, H.H.; Mooney, D.J. Functional muscle regeneration with combined delivery of angiogenesis and myogenesis factors. *Proc. Natl. Acad. Sci. USA* 2010, 107, 3287–3292. [CrossRef] [PubMed]

- 11. Hammond, J.W.; Hinton, R.Y.; Curl, L.A.; Muriel, J.M.; Lovering, R.M. Use of autologous platelet-rich plasma to treat muscle strain injuries. *Am. J. Sports Med.* 2009, 37, 1135–1142. [CrossRef] [PubMed]
- Anitua, E.; Sanchez, M.; Orive, G. Potential of endogenous regenerative technology for in situ regenerative medicine. Adv. Drug Deliv. Rev. 2010, 62, 741–752. [CrossRef] [PubMed]
- Glass, D.J. Skeletal muscle hypertrophy and atrophy signaling pathways. Int. J. Biochem. Cell Biol. 2005, 37, 1974–1984. [CrossRef] [PubMed]
- Tentori, L.; Graziani, G. Doping with growth hormone/IGF-1, anabolic steroids or erythropoietin: Is there a
 cancer risk? *Pharmacol. Res.* 2007, 55, 359–369. [CrossRef] [PubMed]
- Rowlands, M.A.; Tilling, K.; Holly, J.M.; Metcalfe, C.; Gunnell, D.; Lane, A.; Davis, M.; Donovan, J.; Hamdy, F.; Neal, D.E.; et al. Insulin-like growth factors (IGFs) and IGF-binding proteins in active monitoring of localized prostate cancer: A population-based observational study. Cancer Cause Control 2013, 24, 39–45. [CrossRef] [PubMed]
- Grimberg, A. Mechanisms by which IGF-I. may promote cancer. Cancer Biol. Ther. 2003, 2, 630–635. [CrossRef] [PubMed]
- Maniscalco, L.; Iussich, S.; Morello, E.; Martano, M.; Gattino, F.; Miretti, S.; Biolatti, B.; Accornero, P.; Martignani, E.; Sanchez-Cespedes, R.; et al. Increased expression of insulin-like growth factor-1 receptor is correlated with worse survival in canine appendicular osteosarcoma. Vet. J. 2015, 205, 272–280. [CrossRef] [PubMed]
- Lim, H.Y.; Im, K.S.; Kim, N.H.; Kim, H.W.; Shin, J.I.; Yhee, J.Y.; Sur, J.H. Effects of Obesity and Obesity-Related Molecules on Canine Mammary Gland Tumors. Vet. Pathol. 2015, 52, 1045–1051. [CrossRef] [PubMed]
- Schippinger, G.; Oettl, K.; Fankhauser, F.; Spirk, S.; Domej, W.; Hofmann, P. Influence of Intramuscular Application of Autologous Conditioned Plasma on Systemic Circulating IGF-1. J. Sports Sci. Med. 2011, 10, 439–444. [PubMed]
- Wang, Y.; Cheng, Z.; Elalieh, H.Z.; Nakamura, E.; Nguyen, M.T.; Mackem, S.; Clemens, T.L.; Bikle, D.D.; Chang, W. IGF-1R signaling in chondrocytes modulates growth plate development by interacting with the PTHrP/Ihh pathway. J. Bone Miner. ResNLM 2011, 26, 1437–1446. [CrossRef] [PubMed]
- Zanou, N.; Gailly, P. Skeletal muscle hypertrophy and regeneration: Interplay between the myogenic regulatory factors (MRFs) and insulin-like growth factors (IGFs) pathways. Cell Mol. Life Sci. 2013, 70, 4117–4130. [CrossRef] [PubMed]
- Yang, S.Y.; Goldspink, G. Different roles of the IGF-I Ec peptide (MGF) and mature IGF-I in myoblast proliferation and differentiation. FEBS Lett. 2002, 522, 156–160. [CrossRef]
- Tong, Y.; Feng, W.; Wu, Y.; Lv, H.; Jia, Y.; Jiang, D. Mechano-growth factor accelerates the proliferation and osteogenic differentiation of rabbit mesenchymal stem cells through the PI3K/AKT pathway. BMC Biochem. 2015, 16, 1. [CrossRef] [PubMed]
- Grahnen, A.; Kastrup, K.; Heinrich, U.; Gourmelen, M.; Preece, M.A.; Vaccarello, M.A.; Guevara-Aguirre, J.;
 Rosenfeld, R.G.; Sietnieks, A. Pharmacokinetics of recombinant human insulin-like growth factor I given
 subcutaneously to healthy volunteers and to patients with growth hormone receptor deficiency. *Acta Paediatr.*1993, 82, 9–13. [CrossRef]
- World Anti-Doping Agency (WADA). The 2018 Prohibited List. 2018. Available online: https://www.wada-ama.org/en/content/what-is-prohibited (accessed on 1 January 2018).
- Wasterlain, A.S.; Braun, H.J.; Harris, A.H.; Kim, H.J.; Dragoo, J.L. The systemic effects of platelet-rich plasma injection. Am. J. Sports Med. 2013, 41, 186–193. [CrossRef] [PubMed]
- Rowlands, M.A.; Gunnell, D.; Harris, R.; Vatten, L.J.; Holly, J.M.; Martin, R.M. Circulating insulin-like growth factor peptides and prostate cancer risk: A systematic review and meta-analysis. *Int. J. Cancer* 2009, 124, 2416–2429. [CrossRef] [PubMed]
- Conway, K.; Price, P.; Harding, K.G.; Jiang, W.G. The molecular and clinical impact of hepatocyte growth factor, its receptor, activators, and inhibitors in wound healing. Wound Repair Regen. 2006, 14, 2–10. [CrossRef] [PubMed]

Int. J. Mol. Sci. 2018, 19, 2701

 Anitua, E.; Andia, I.; Sanchez, M.; Azofra, J.; del Mar Zalduendo, M.; de la Fuente, M.; Nurden, P.; Nurden, A.T. Autologous preparations rich in growth factors promote proliferation and induce VEGF and HGF production by human tendon cells in culture. *J. Orthop. Res.* 2005, 23, 281–286. [CrossRef] [PubMed]

- Sanchez, M.; Anitua, E.; Azofra, J.; Andia, I.; Padilla, S.; Mujika, I. Comparison of surgically repaired Achilles tendon tears using platelet-rich fibrin matrices. Am. J. Sports Med. 2007, 35, 245–251. [CrossRef] [PubMed]
- Creaney, L.; Hamilton, B. Growth factor delivery methods in the management of sports injuries: The state of play. Br. J. Sports Med. 2008, 42, 314–320. [CrossRef] [PubMed]
- 32. Foster, T.E.; Puskas, B.L.; Mandelbaum, B.R.; Gerhardt, M.B.; Rodeo, S.A. Platelet-rich plasma: From basic science to clinical applications. *Am. J. Sports Med.* 2009, 37, 2259–2272. [CrossRef] [PubMed]
- Chen, B.; Liu, S.; Xu, W.; Wang, X.; Zhao, W.; Wu, J. IGF-I and IGFBP-3 and the risk of lung cancer: A meta-analysis based on nested case-control studies. J. Exp. Clin. Cancer Res. 2009, 28, 89. [CrossRef] [PubMed]
- Marx, R.E. Platelet-rich plasma: Evidence to support its use. J. Oral Maxillofac. Surg. 2004, 62, 489–496.
 [CrossRef] [PubMed]
- Vilar, J.M.; Damiá, E.; Rubio, M.; Santana, A.; Sopena, J.; Ceron, J.; Tvarijonaviciute, A.; Cugat, R.; Carrillo, J.M.
 Therapeutic doses of plasma rich in growth factors cannot provoke cancer by means of the IGF-1 pathway or inflammation in dogs. J. Appl. Anim. Res. 2016, 45, 490–493. [CrossRef]
- Favier, R.P.; Mol, J.A.; Kooistra, H.S.; Rijnberk, A. Large body size in the dog is associated with transient GH excess at a young age. J. Endocrinol. 2001, 170, 479–484. [CrossRef] [PubMed]
- Blanchard, G.; Nguyen, P.; Gayet, C.; Leriche, I.; Siliart, B.; Paragon, B.M. Rapid weight loss with a high-protein low-energy diet allows the recovery of ideal body composition and insulin sensitivity in obese dogs. J. Nutr. 2004, 134 (Suppl. 8), 21485–2150S. [CrossRef] [PubMed]
- Coronado, R.; Diez, M.P.; Chévez, D. Correlación de edad, niveles séricos de IGF-1 e índice de masa muscular, y su influencia como determinantes de las variables isocinéticas en pacientes con osteoporosis. Cirugia y Cirujanos 2005, 73, 457–463.
- Greer, K.A.; Hughes, L.M.; Masternak, M.M. Connecting serum IGF-1, body size, and age in the domestic dog. Age (Dordr.) 2011, 33, 475–483. [CrossRef] [PubMed]
- Fernandez-Sarmiento, J.A.; Dominguez, J.M.; Granados, M.M.; Morgaz, J.; Navarrete, R.; Carrillo, J.M.; Gomez-Villamandos, R.J.; Munoz-Rascon, P.; Martin de Las Mulas, J.; Millan, Y.; et al. Histological study of the influence of plasma rich in growth factors (PRGF) on the healing of divided Achilles tendons in sheep. J. Bone Jt. Surg. Am. 2013, 95, 246–255. [CrossRef] [PubMed]
- Tvarijonaviciute, A.; Tecles, F.; Carrillo, J.M.; Rubio, M.; Ceron, J.J. Serum insulin-like growth factor-1
 measurements in dogs: Performance characteristics of an automated assay and study of some sources of
 variation. Can. J. Vet. Res. 2011, 75, 312–316. [PubMed]
- Connell, D.A.; Schneider-Kolsky, M.E.; Hoving, J.L.; Malara, F.; Buchbinder, R.; Koulouris, G.; Burke, F.; Bass, C. Longitudinal study comparing sonographic and MRI assessments of acute and healing hamstring injuries. AJR Am. J. Roentgenol. 2004, 183, 975–984. [CrossRef] [PubMed]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).

TRATAMIENTO OA Y ARTÍCULOS

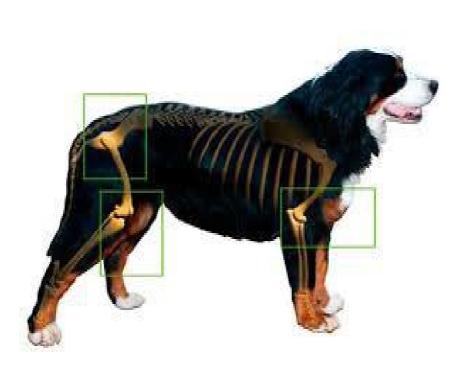
TRATAMIENTO OA Y ARTÍCULOS



CONCLUSIONES

6. CONCLUSIONES

- 1) El análisis de la trayectoria del COP en los miembros de animales que presentan cojera unilateral por OA de codo ha permitido demostrar que en los miembros con cojera dicha trayectoria es más corta, está situada en posición más craneal y con menor desplazamiento mediolateral respecto a los miembros sanos; también se ha demostrado que en ambos miembros el COP describe una trayectoria craneomedial y no el eje longitudinal del miembro. Por último, la progresión del COP no se desarrolla a una velocidad constante durante la fase de apoyo.
- 2) El PRGF acelera el tiempo de consolidación ósea, por lo que acorta el tiempo de retirada de los implantes, en este caso fijadores externos.
- 3) La aplicación intramuscular de PGRF a dosis terapéuticas, e incluso triplicando esta, no incrementa los niveles de igf-1 en sangre ni incrementa la masa muscular. Esto determina que es segura en términos de no provocar efectos locales ni generales (cáncer) a través de aumentos significativos de los niveles de IGF-1 en sangre.



RESUMEN



7. RESUMEN

La osteoartritis (OA) constituye hoy en día una de las patologías del aparato locomotor de mayor morbilidad no solo en la especie humana, sino también en el campo de la veterinaria. Por lo tanto, a la hora de su diagnóstico en las especies animales, el principal caballo de batalla es el de establecer de una manera objetiva el déficit funcional que el animal está padeciendo para, si un tratamiento es instaurado, conocer de forma igualmente objetiva el verdadero nivel de eficacia.

Por todo ello, en este campo las técnicas de evaluación (diagnóstico) basadas en la biomecánica han ido adquiriendo especial relevancia en los últimos años. Entre ellos, los análisis con las plataformas de presión proporcionan multitud de parámetros que en conjunto pueden dar una visión global de las características y alteraciones de la locomoción tanto en animales sanos como en los que presentan cojera o, incluso, déficits neurológicos.

De este elenco de parámetros, el estudio de la trayectoria del centro de presiones (COP) está presente en estudios actuales, tanto en humana como en veterinaria.

Dentro de las estrategias para el tratamiento de la OA adquieren especial relevancia en la actualidad las terapias regenerativas-reparativas, principalmente a partir de células madre mesenquimales y plasma rico en plaquetas. En este último caso, su uso es muy variado en campos como la dermatología, oftalmología, y el aparato locomotor, aunque no existenestudios



de su eficacia en fracturas en veterinaria. La terapia a base de PRP, a pesar de ser autóloga, es objeto de polémica en la actualidad por su supuesto efecto ergogénico e incluso carcinogénico por vía de uno de los factores de crecimiento, en concreto el IGF-1. Siendo de un uso muy frecuente en los campos anteriormente citados, parece procedente realizar estudios directos e indirectos sobre su seguridad.

Por todo esto, el objetivo común de esta tesis doctoral consiste en estudiar aspectos de diagnóstico, y tratamiento en animales con osteoartritis y fracturas. Y por este motivo, se han diseñado tres estudios diferentes:

- Un primer estudio utilizando 10 perros con OA unilateral de codo para valorar la eficacia del estudio del COP en la detección de cojeras utilizando una plataforma de presión.
- Un segundo estudio utilizando 65 perros de diversas razas con fracturas en huesos largos, para comprobar la eficacia del PRP en la aceleración de la consolidación ósea.
- Por último, en tercer estudio sobre la seguridad del PRP a dosis terapéuticas, utilizando 24 perros de raza Beagle a los que se les inyecto PRP por vía intramuscular y se evaluó los niveles de igf-1 en sangre, además de técnicas de imagen para evaluar si existía desarrollo muscular.

Adicionalmente, se aporta un estudio de revisión sobre la eficacia de las células madre mesenquimales en el tratamiento de la OA.











SUMMARY

8. SUMMARY

Osteoarthritis (OA) is today one of the pathologies of the locomotor system of greater morbidity not only in the human species, but also in the field of veterinary medicine. Therefore, at the time of diagnosis in animal species, the main workhorse is to establish in an objective manner the functional deficit that the animal is suffering for, if a treatment is established, to know equally objectively the true level of effectiveness.

For all these reasons, in this field the evaluation techniques (diagnosis) based on biomechanics have acquired special relevance in recent years. Among them, the analyzes with the pressure platforms provide a multitude of parameters that together can give a global view of the characteristics and alterations of the locomotion in healthy animals as well as in those that present lameness or, even, neurological deficits.

From this list of parameters, the study of the trajectory of the pressure center (COP) is present in current studies, both human and veterinary.

Among the strategies for the treatment of OA, regenerative-reparative therapies are especially relevant today, mainly from mesenchymal stem cells and platelet-rich plasma. In the latter case, its use is very varied in fields such as dermatology, ophthalmology, and the locomotor system, although there are no studies of its effectiveness in veterinary fractures. PRP therapy, despite being autologous, is currently controversial due to its supposed ergogenic and even carcinogenic effect via one of the growth factors, specifically IGF-1. Being of



very frequent use in the aforementioned fields, it seems appropriate to carry out direct and indirect studies on its safety.

For all this, the common objective of this doctoral thesis is to study aspects of diagnosis, and treatment in animals with osteoarthritis and fractures.

For this, three different studies have been designed:

- a first study using 10 dogs with unilateral elbow OA to assess the effectiveness of the COP study in the detection of lameness using a pressure platform.
- A second study using 65 dogs of different breeds with fractures in long bones, to check the effectiveness of PRP in the acceleration of bone consolidation.
- Finally, in the third study on the safety of PRP at therapeutic doses, using 24 Beagle dogs to which PRP was injected intramuscularly and blood igf1 levels were evaluated, in addition to imaging techniques for evaluate if there was muscle development.

Additionally, a review study on the efficacy of mesenchymal stem cells in the treatment of OA is provided.



BIBLIOGRAFÍA







Abrantes, J. & Santos, L., 2012. Plantar pressure assessment: a new tool for postural instability diagnosis in multiple sclerosis. En: Jorge, R., Tavares, J., Barbosa, M., Slade, A. (eds). Technologies for medical sciences. Springer, The Netherlands. 179-206. ISBN: 978-94-007-4067-9.

Adams, D., 2004. Canine anatomy a systematic study. 4a. ed. Wiley-Blackwell, Iowa. 41-8; 71-81.

Adrian, A.M., Barrett, M.F., Werpy, N.M., Kawcak, C.E., Chapman, P.L., Goodrich, L.R., 2017. A comparison of arthroscopy to ultrasonography for identification of pathology of the equine stifle. Equine Vet J. 49(3), 314-321.

Adrian, M.J., Rog, W.E. & Karpovich, D.V. Normal gait of the dog: An electrogoniometric study. Am J. Vet. Res. 27 (116), 90-95.

Anitua, E., Sánchez, M., Nurden, A.T., Zalduendo, M.M., de la Fuente, M., Azofra, J. & Andia, I., 2007a. Platelet---released growth factors enhance the secretion of hyaluronic acid and induce hepatocyte growth factor production by synovial fibroblasts from arthritic patients. Rheumatology. 46 (12), 1769--1772.

Anjos, D.M., Gomes, L.P., Sampaio, L.M., Correa, J.C. & Oliveira C.S., 2010. Assessment of plantar pressure and balance in patients with diabetes. Arch Mmed Sci. 6 (1), 43-48.

Anker, L.C., Weerdesteyn, V., van Nes, I.J.W., Nienhuis, B., Straatman, H. & Geurts, A.C.H., 2008. The relation between postural stability and weight distribution in healthy subjects. Gait Posture. 27 (3), 471-477.

Baratto, L., Morasso, P.G., Re, C. & Spada, G., 2002. A new look at posturographic analysis in the clinical context: sway-density versus other parameterization techniques. Motor control. 6 (3), 246-270.

Barone, R., 1980. Anatomie Comparée des mammifères domestiques. 2da ed. Vigot, Paris. Tomo 2.



Barrey, E., Galloux, P., Valette J.P., Auvinet, B. & Wolter, R., 1993. Stride characteristics of overground versus treadmill locomotion in the saddle horse. Acta Anat (Basel). 146 (2-3), 90-94.

Bhosale, A. M. & Richardson, J. B., 2008. Articular cartilage: structure, injuries and review of management. Br Med Bull. 87(1), 77--95.

Bialski, D., Lanovaz, J.L., Bohart, G.V., Mullineaux, D.R. & Clayton, H.M., 2003. Effect of detomidine on postural sway in horses. Comp Exerc Physiol. 1 (1), 45–50.

Birch, H.L., McLaughlin, L., Smith, R.K. & Goodship, A.E. 1999a. Treadmill exercise-induced tendon hypertrophy: assessment of tendons with different mechanical functions. Equine Vet J. 31 (S30), 222-226.

Black, L.L., Gaynor, J., Adams, C., Dhupa, S., Sams, A.E., Talor, R., Harman, S., Ginggerich, D.A. & Harman, R., 2008. Effect of intraarticular injection of autologous adipose--- derived mesenchymal stem and regenerative cells on clinical signs of chronic osteoarthritis of the elbow joint in dogs. Vet Ther. 9 (3), 192--200.

Bland, S.D., 2015. Canine osteoarthritis and treatments: a review. Vet Sci Dev. 5 (1), 84-89.

Bockstahler, B., Tichy, A. & Aigner, P., 2016. Compensatory load redistribution in Labrador retrievers when carrying different weights-a non-randomized prospective trial. BMC Vet Res. 12 (1), 92. doi:10.1186/s12917-016-0715-7.

Buchner, H.H., Savelberg, H.H., Schamhardt, H.C., & Merkens, H.W., 1994. Kinematics of treadmill vs overground locomotion in horses. Vet Q. 16 (2), S87-S90.

Buchner, H.H., Savelberg, H.H., Schamhardt, H.C. & Barneveld, A., 1996. Head and trunk movement adaptations in horses with experimentally induced fore-or hindlimb lameness. Equine Vet J. 28 (1), 71-76.



Budsberg, S.C., Rytz, U. & Johnston, S.A., 1999. Effects of acceleration on ground reaction forces collected in healthy dogs at a trot. Vet Comp Orthop Traumatol. 12 (1), 15-19.

Burr, D. B., 2004. Anatomy and physiology of the mineralized tissues: role in the pathogenesis of osteoarthrosis. Osteoarthritis Cartilage. 12, 20-30.

Cakir, S., Hepguler, S., Ozturk, C., Korkmarz, M., Isleten, B. & Atamaz, F.C., 2014. Efficacy of Therapeutic Ultrasound for the Management of Knee Osteoarthritis: A Randomized, Controlled, and Double---Blind Study. Am J Phys Med Rehabil. 93 (5), 405-412.

Cardinet, G. H., Kass, P. H., Wallace, L.J., Guffy, M. M., 1997. Association between pelvic muscle mass and canine hip dysplasia. JAVMA. 210 (10), 1466-1473.

Carr, B.J., Canapp, S.O. & Zink, M.C., 2015. Quantitative Comparison of the Walk and Trot of Border Collies and Labrador Retrievers, Breeds with Different Performance Requirements. PLoS ONE. 10 (12), e0145396. doi:10.1371/journal.pone.0145396

Castañon, F., 2015. Estudios comparativo de las técnicas quirúrgicas, TTA clásicas Securos ®, TTA Porous ® y TTA Porous ® con PRP, para el tratamiento de la rotura del ligamento cruzado anterior en el perro. Tesis Doctoral,León.

Cimino Brown, D., Boston, R.C., Coyne. J.C. & Farrar, J. T., 2007. Development and psychometric testing of an instrument designed to measure chronic pain in dogs with osteoarthritis. Am J Vet Res. 68 (6), 631-637.

Cole, K. & Abbs, J.H., 1986. Coordination of three-joint digit movements for rapid fingerthumb grasp. J. Neurophysiol. 55 (6), 1407-1423.

Cordeiro, T.L., Frade, M.A., Barros, A.R. & Foss, N.T. 2014. Postural balance control of the leprosy patient with plantar sensibility impairment. Occup Med Health Aff. 2, 3.



Cuervo, B., Rubio, M., Sopena, J., Domínguez, J.M., Vilar, J., Morales, M., Cugat, R., Carrillo, J.M., 2014. Hip osteoarthritis in dogs: a randomized study using mesenchymal stem cells from adipose tissue and plasma rich in growth factors. Int J Mol Sci. 15 (8), 13437-13460.

Cuervo, B. 2015. "Estudio Clínico Multicéntrico del Efecto de la Aplicación de Células Mesenquimales de Grasa (CMG), Plasma Rico en Factores de Crecimiento (PRGF) y la Combinación de Ambos en el Tratamiento de la Enfermedad Degenerativa Articular en Perro", Tesis Doctoral, Valencia.

Cruz, A.M., Maninchedda, U.E., Burger, D., Wanda, S., Vidondo, B., 2017. Repeatability of gait pattern variables measured by use of extremity-mounted inertial measurement units in nonlame horses during trotting. Am J Vet Res. 78 (9), 1011-1018. Doi: 10.2460/ajvr. 78.9.1011.

Damiá, E., Chicharro, D., Rubio, M., Carrillo, J.M., Sopena, J., Cuervo, B., López, S., Vilar, J.M., 2018. Can Plasma Rich in Growth Factors Be Safe for Parental Use? A Safety Study in the Canine Model. Int J Mol Sci. 19 (9), 2701.

Diekman, B.O. & Guilak, F., 2013. Stem cell---based therapies for osteoarthritis: challenges and opportunities. Curr Opin Rheumatol. 25 (1), 119--126.

Drevemo, S., Dalin, G., Fredricson, I., Hjertén, G. & Hjertén, G., 1980. Equine locomotion: 1. The analysis of linear and temporal stride characteristics of trotting standardbreds. Equine Vet J. 12 (2), 60-65.

Durante, E.J., Brusa, M.C., 1998. Algunos aspectos de la displasia del codo de los caninos. Analecta Veterinaria. 18 (1-2), 59-70.

Duval, J.M., Budsberg, S.C., Flo, G.L. & Sammarco, J.L., 1999. Breed, sex, and body weight as risk factors for rupture of the cranial cruciate ligament in young dogs. J Am Vet Med Assoc. 215(6), 811-814.

Elkins, A.D., Pechman, R., Kearney, M. & Herron, N., 1991. A retrospective study evaluating the degree of degenerative joint disease in the stifle e joint of

dogs following surgical repair of anterior cruciate ligament rupture. J Am Anim Hosp Assoc. 27(5), 533-540.

Evans, H.E., 1993. Arthrology. En: Evans, H.E., (Eds.), Miller's anatomy of the dog, ed 3, WB Sunders, Philadelphia.

Evans, H.E., DelaHunta, A., 2000. Disección del perro. 5ta ed. McGraw-Hill-Interamericana, México. 384.

Evans, R., Horstman, C. & Conzemius, M., 2005. Accuracy and optimization of force platform gait analysis in Labradors with cranial cruciate disease evaluated at a walking gait. Vet Surg. 34 (5), 445-449.

Fahie, M.A., Ortolano, G.A., Guercio, V., Schaffer, J.A., Johnston, G., AU, J., Hettlich, B.A., Phillips, T., Allen, M.J. & Bertone, A.L., 2013. A randomized controlled trial of the efficacy of autologous platelet therapy for the treatment of osteoarthritis in dogs. J Am Vet Med Assoc. 243 (9), 1291--1297.

Fang, J., Zheng, N., Wang, Y., Cao, H., Sun, S., Dai, J., Li, Q. & Zhang, Y. 2013. Understanding Acupuncture Based on ZHENG Classification from System Perspective. Evid Based Complement Alternat Med. 2013, 956967-10.

Fortier, L.A., Barker, J.U., Strauss, E.J., McCarrel, T.M. & Cole, B.J., 2011. The role of growth factors in cartilage repair. Clin Orthop Relat Res. 469 (10), 2706-2715.

Fossum, T. W., 2009. Cirugía en pequeños animales. 3ra Ed. Elsevier España, Barcelona. 1632 pp.

Godley, D.R., 2013. Assessment, diagnosis, and treatment of developmental dysplasia of the hip. JAAPA. 26 (3), 54--58.

Gomes-Costa, M., Roupa, I., Pequito, M., Prazeres, J., Gaivão, M., Abrantes, J. & Clayton, H.M., 2015. The use of pressure plates for static Center of Pressure Analysis in horses. J Equine Vet Sci. 35 (4), 315–320.



Gómez Álvares C.B., Gustås P., Bergh A. & Rhodin M., 2017. Vertical head and pelvic movement symmetry at the trot in dogs with induced supporting limb lameness. Vet J. 229, 13-18.

Gordon, W.J., Conzemius, M.G., Riedesel, E., Besancon, M. F., Evans, R., Wilke, V. & Ritter, 2003. The relationship between limb function and radiographic osteoarthrosis in dogs with stifle osteoarthrosis. Vet Surg. 32 (5), 451-454.

Gudbergsen, H., Boesen, M., Lohmander, L.S., Christensen, R., Henriksen, M., Bartels, E.M., Christensen, P., Rindel, L., Aaboe, J., Danneskiold--Samsoe, B., Riecke, B.F. & Bliddal, H., 2012. Weight loss is effective for symptomatic relief in obese subjects with knee osteoarthritis independently of joint damage severity assessed by high--field MRI and radiography. Osteoarthritis Cartilage. 20 (6), 495--502.

Gustås, P., Pettersson, K., Honkavaara, S., Lagerstedt, A., Byström, A & Sveriges, I., 2003. Kinematic and temporospatial assessment of habituation of Labrador retrievers to treadmill trotting. Vet J. 198(1), 114-119. Doi.org/10.1016/j.tvjl.2013.09.044

Harasen, G., 2002. Diagnosing rupture of the cranial cruciate ligament. Can Vet J. 43 (6), 475–476.

Heffron, L.E., Campbell, J.R., 1978. Morphology, histology and functional anatomy of the canine cranial cruciate ligament. Vet Rec. 102, 280-283.

Heikkilä, H.M., Hielm-Björkman, A.K., Morelius, M., Larsen, S., Honkavaara, J., Innes, J. F. & Laitinen- Vapoavouri, O.M., 2014. Intra-articular botulinum toxin A for the treatment of osteoarthritic joint pain in dogs: A randomized, double-blinded, placebocontrolled clinical trial. Vet Journal. 200 (1), 162-169.

Hielm-Björkman, A.K., Kuusela, E., Liman, A., Markkola, A., Saarto, E., Huttunen, P., Leppäluoto, J., Tulamo, R. & Reakallio, M., 2003. Evaluation of methods for assessment of pain associated with chronic osteoarthritis in dogs. J Am Vet Med Assoc. 222 (11), 1552-1558.



Hielm-Björkman, A.K., Kapatkin, A.S. & Rita, H.J., 2011. Reliability and validity of a visual analogue scale used by owners to measure chronic pain attributable to osteoarthritis in their dogs. Am J Vet Res. 72 (5), 601-607.

Hielm-Björkman, A.K., Roine, J., Elo, K., Lappalainen, A., Junnila, J. & Laitinen- Vapaavuari, O., 2012. An un-commissioned randomized, placebo-controlled double-blind study to test the effect of deep sea fish oil as a pain reliever for dogs suffering from canine OA. BMC Vet Res. 8 (1), 157.

Horstam, C.L., Conzemius, M.G., Evans, R. & Gordon, W. J., 2004. Assessing the efficacy of perioperative oral carprofen after cranial cruciate surgery using noninvasive, objective pressure platform gait analysis. Vet Surg. 33(3), 286-292.

Hudson, J.T., Slater, M.R., Taylor, L., Scott, H.M. & Kerwin, S.C., 2004. Assessing repeatability and validity of a visual analoque scale questionnaire for use in assessing pain and lameness in dogs. Am J Vet Res. 65 (12), 1634-1643.

Innes, J.F., Barr, A.R.S., 1998a Clinical natural history of the postsurgical cruciate deficient canine stifle joint: year 1. J Small Anim Pract. 39 (7), 325-332.

Innes, J.F., Barr, A.R.S., 1998b. Can owners assess outcome following treatment of canine cruciate ligament deficinency? J Small Anim Pract. 39 (8), 373-378.

Ishihara, A., Reed, S.M., Rajala-Schultz, P.J., Robertson, J.T. & Bertone, A.L., 2009. Use of kinetic gait analysis for detection, quantification, and differentiation of hindlimb lameness and spinal ataxia in horses. J Am Vet Med Ass. 234 (5), 644–651.

Johnson, J.A., Austin, C., Breur, G.J., 1994. Incidence of canine appendicular musculoskeletal disorders in 16 veterinary teaching hospitals from 1980 to 1989. Vet Comp Orthop Traumatol. 7, 56-59.



Karpovich, P.V., Herden, E.L. & Asa, M.M., 1960. Electrogoniometric study of joints. U.S. Armed Forces Med. J. 11, 425-450.

Keegan, K.G., Wilson, D.A., Smith, B.K. & Wilson, D.J., 2000. Changes in kinematic variables observed during pressure-induced forelimb lameness in adult horses trotting on a treadmill. Am J Vet Res. 61(6), 612-619.

Keegan, K.G., Yonezawa, Y., Pai, P.F. & Wilson, D.A., 2002. Accelerometer-based system for the detection of lameness in horses. Biomed Sci Instrum. 38, 107-112.

Kelmer, G., Keegan, K.G., Kramer, J., Wilson, D.A., Pai, F.P. & Singh, P., 2005. Computer-assisted kinematic evaluation of induced compensatory movements resembling lameness in horses trotting on a treadmill. Am J Vet Res. 66 (4), 646-655.

Kirberger, G., 2003. Elbow dysplasia in the dog- What is it? IEWG. 15, 15-19.

Kiberger, R., Barr, F., 2006. Manual of canine and musculosketetal imaging. BSAVA, London. 220.

Kim, Y.M., Joo, Y.B., 2018. Anteromedial Meniscofemoral Ligament of the Anterior Horn of the Medial Meniscus: Clinical, Magnetic Resonance Imaging, and Arthroscopic Features. Arthroscopy. 34(5), 1590-1600.

King, M.R., Haussler, K.K., Kawcak, C.E., Mcllwraith, C.W. & Reiser R.F., 2013. Effect of underwater treadmill exercise on postural sway in horses with experimentally induced carpal joint osteoarthritis. Am J Vet Res. 74 (7), 971-982.

Khumsap, S., Lanovaz, J.L., Clayton, H.M. 2004 Verification of skin-based markers for 3-dimentional kinematic analysis of the equine tarsal joint. Equine Vet J. 36 (8), 655-658.



Komsta, R., Debiak, P., Twardowski, P. 2008. Radiographic evaluation of joints in dogs with elbow dyplasia- clinical observations. Bull Vet Inst Pulawy. 52, 179-183.

König, H. & Liebich, H., 2008. Introducción. En: König, H. & Liebich, H. (eds.) Anatomía de los animales domésticos. Aparato locomotor. 2ª ed. Montevideo, Uruguay: Editorial médica panamericana.

Kramer, J., Keegan, K.G., Kelmer, G. & Wilson D.A., 2004. Objetive determination of pelvic movement during hind limb lameness by use of a signal decomposition method and pelvic height differences. Am J Vet Res. 65(6), 741-747.

Kramer, J. & Keegan, K., 2007. Capítulo 13: Cinemática de la claudicación. En: medicina y cirugía en los equinos de deporte. Kkenneth W. Hinchcliff, Andris J. Kaneps, Raymond J. Geor. (eds). Editorial Inter-Médica S.A.I.C.I. 266-282. ISBN: 978-950-555-327-3.

Langlois B., Froideveaux J., Lamarche L., Legault, C., Legault, P., Tassencourt, L. & Théret, M., 1978. Analyse des liaisons entre la morphologie et l'aptitude au gallop, au trot et au saut d'obstacles chez le cheval. Annales de Génétique et de Sélection Animale, 10 (3), 443-474.

Leach, D.H. & Cymbaluk, N.F., 1986. Relationship between stride length, stride frequency, velocity and morphometrics of foals. Am J Vet Res. 47 (9), 2090-2097.

Leach, D.H. & Dyson, S., 1988. Instant centres of rotation of equine limbjoints and their relationship to standard skin marker locations. Equine Vet J. 20 (6), 113-119.

Lippincott, C.L. Femoral head and neck excision in the management of canine hip dysplasia. Vet Clin North Am- Small Anim Pract. 1992; vol. 22 (3):. 721-737.



Luzio Quiroga, A., Campos Oyarce, P., Troncoso Toro, I., Fischer Wiethunter, C., Gili Graf, R., 2014. Evaluación clínica y radiológica de la articulación húmero- radio-ulnar en perros de trabajo policial en la ciudad de Concepción, Chile. Rev Med Vet. 27, 121- 131.

Madsen, J. S., Svalastoga, E. Inclination and anteversion of collum femoris in hip dysplasia and coxarthritis. Acta Vet Scand. 1994; vol. 35 (2):. 115-119.

Madsen, J. S., Svalastoga, E. Early diagnosis of hip dyspasia- A stress radiographic study. Vet Comp Orthop Traumatol. 1995; vol 8:. 114-117.

Malek, S., Sample, S. J., Schwartz, Z., Nemke, B., Jaconson, P. B., Cozzi, E. M., Schaefer, S. L., Bleedorm, J. A., Holzman, G. & Muir, P., 2012. Effect of analgesic therapy on clinical outcome measures in a randomized controlled trial using client---owned dogs with hip osteoarthritis. BMC Vet Resch. 8, 185.

Maliye, S., Voute, L., Lund, D. & Marshall, J.F., 2013. An inertial sensor-based system can objectively assess diagnostic anesthesia of the equine foot. Equine Vet J. 45 (45) 26-30. Doi: 10.1111/evj.12158.

Malouin, F. & Bedard, P., 1983. Evaluation of head motility and posture in cats with horizontal torticollis. Exp. Neurol. 81(3) 559-570.

Manera, M., Carrillo, J., Batista, M., Rubio, M., Sopena, J., Santana, A. and Vilar, J. (2017). Static Posturography: A New Perspective in the Assessment of Lameness in a Canine Model. *PLOS ONE*, 12(1)

Martinez- Méndez R., Sekine, M. & Tamura, T., 2011a. Postural Sway parameters using a triaxial accelerometer: comparing elderly and young healthy adults. Comput Methods Biomech Biomed Engin. 15 (9), 899-910.

Martinez- Méndez, R., Sekine, M. & Tamura, T., 2011b. Detection of anticipatory postural adjustments prior to gait initiation using inertial wearable sensors. J Neuroeng Rehabil. 8 (1), 17.

Martinez-Méndez, R. & Huertas, M.R., 2013. Uso de sensores inerciales en la medición y evaluación de movimiento humano para aplicaciones en la salud. Universidad Autónoma del Estado de México, Facultad de Ingeniería.



Merashly, M. & Uthman, I., 2012. Management of knee osteoarthritis: an evidence---based review of treatment options. J Med Liban. 60 (4), 237--242.

Moleman, M., van Heel, M.C., van Weeren, P.R. & Back, W., 2006. Hoof growth between two shoeing sessions leads to a substantial increase of the moment about the distal, but not the proximal, interphalangeal joint. Equine Vet J. 38 (2), 170–174.

Morgan, J., Wind, A., Davidson, A., 1999. Bone dysplasias in the labrador retriever: A radiographic study. J Am Anim Hosp Assoc. 35 (4), 332-340.

Mölsä, S.H., Hielm-Bjorkman, A. K & Laitinen-Vapaavuori, O.M., 2010. Force platform in clinically healthy rottweilers: comparison with labrador. Vet Surg. 39 (6), 701-107.

Nelson, S.A., Krotscheck, U., Rawlinson, J., Todhunter, R.J., Zhang, Z. & Mohammed, H., 2013. Long-term functional outcome of tibial plateau leveling osteotomy versus extracapsular repair in a heterogeneous population of dogs. Vet Surg. 42 (1), 38-50.

Oldershaw, R.A., 2012. Cell sources for the regeneration of articular cartilage: the past, the horizon and the future. Int J Exp Pathol. 93 (6), 389--400.

Oosterlinck, M., Pille, F., Back, W., Dewulf, J. & Gasthuys, F., 2010a. Use of a stand-alone pressure plate for the objective evaluation of forelimb symmetry in sound ponies at walk and trot. Vet J. 183 (3), 305-309.

Oosterlinck, M., Pille, F., Huppes, T., Gasthuys, F. & Back, W., 2010b. Comparison of pressure plate and force plate gait kinetics in sound warmbloods at walk and trot. Vet J. 186 (3), 347–351.

Paulson, S.K., Zhang, J.Y., Jessen, S.M., Lawal, Y., Liu, N.W., Dudkowski, C.M., Wang, Y.F., Chang, M., Yang, D., Findlay, J.W., Berge, M.A., Markos, C.S., Breau, A.P., Hribar, J.D. & Yuan, J., 2000. Comparison of celecoxib metabolism and excretion in mouse, rabbit, dog, cynomolgus monkey and rhesus monkey. Xenobiotica. 30 (7), 731-744.



Paulson, S.K., Vaughn, M.B., Jessen, S.M., Lawal, Y., Gresk, C.J., Yan, B., Maziasz, T.J., Cook, C.S. & Karim, A., 2001. Pharmacokinetics of celecoxib after oral administration in dogs and humans: effect of food and site of absorption. J Pharmacol Exp Ther. 297 (2), 638-645.

Penning, T.D., Talley, J.J., Bertenshaw, S.R., Carter, J.S., Collins, P.W., Docter, S., Graneto, M.J., Lee, L.F., Malecha, J.W., Miyashiro, J.M., Rogers, R.S., Rogier, D.J., Yu, S.S., Anderson, G.D., Burton, E.G., Cogburn, J.N., Gregory, S.A., Koboldt, C.M., Perkins, W.E., Seibert, K., Veenhuizen, A.W., Zhang, Y.Y. & Isakson, P.C., 1997. Synthesis and biological evaluation of the 1, 5-diarylpyrazole class of cyclooxygenase 2 inhibitors: identification of 4-[5-(4-methylphenyl)-3-trifluoromethyl)- 1H-pyrazol-1-y1]benze nesulfonamide (SC-58635, celecoxib). J Med Chem. 40 (9), 1347-1365.

Piermattei, D. L., Flo, G.L. & DeCamp, C.E., 2006. The Stifle Joint. En: Brinker, Permattei & Flo's (eds.) Handbook of Small Animal Orthopedics and Fracture Repair, St. Louis: Elsevier Inc, 562-632.

Ramírez-Flores, G.I., Del Angel-Caraza, J., Quijano-Hernández, I.A., Hulse, D.A., Beale, B.S., Victoria-Mora, J.M., 2017. Correlation between osteoarthritic changes in the stifle joint in dogs and the results of orthopedic, radiographic, ultrasonographic and arthroscopic examinations. Vet Res Commun. 41(2), 129-137.

Ratzlaff, M., 1989. Quantitative Methods for the Analysis of Equine Locomotion and Their Applications to Other Species. American Zoologist. 29(1), 267-285.

Reid, M.C., Shengelia, R. & Parker, S.J., 2012. Pharmacologic Management of Osteoarthritis-- Related Pain in Older Adults: A Review Shows that Many Drug Therapies Provide Small--to--Modest Pain Relief. HSS J. 8 (2), 159--164.

Riser, W. H. A half century of canine hip dysplasia. Semin Vet Med Surg (Small Anim). 1987; vol. 2 (2):. 87-91.



Rhodin, M., Bergh, A., Gustås, P. & Gómez-Álvarez, C.B., 2017. Inertial sensorbased system for lameness detection in trotting dogs with induced lameness. Vet J. 222, 54-59.

Riggs, C.M., DeCamp, C.E., Soutas-Little, R.W., Braden, T.D. & Richter, M.A., 1993. Effects of subject velocity on force plate-measured ground reaction forces in healthy Greyhounds at the trot. Am J Vet Res. 54 (9), 1523-1526.

Rodriguez, J. 2003. La displasia de cadera en el dogo canario: Determinación del índice de distracción. Tesis Doctoral, Las Palmas de Gran Canaria.

Ruhe, A., Fejer, R. & Walker, B., 2010. The test-retest reliability of centre of pressure measures in bipedal static task conditions- a systematic review of the literature. Gait Posture. 32 (4), 436-445.

Rychel, J. K., 2010. Diagnosis and treatment of osteoarthritis. Top Companion Anim Med. 25 (1), 20-25.

Saito, I., Koshino, T., Nakashima, K., Uesugi, M. & Saito, T., 2002. Increased cellular infiltrate in inflammatory synovia of osteoarthritic knees. Osteoarthritis Cartilage. 10 (2), 156--162.

Sanchez-Bustinduy, M., de Medeiros, M.A., Radke, H., Langley-Hobbs, S., McKinley, T., Jeffery, N., 2010. Comparison of kinematic variables in defining lameness caused by naturally occurring rupture of the cranial cruciate ligament in dogs. Vet Surg. 39 (4), 523–530.

Sandoval J. 2000. Tratado de anatomía Veterinaria Tomo II. Aparato Locomotor. Imprenta Sorles, León.

Schnabl-Feichter, E., Tichy, A. & Bockstahler, B., 2017. Coefficients of variation of ground reaction force measurement in cats. PLoS One. 12 (3),e0171946.

Smith, G.K. Advances in diagnosing canine hip dysplasia. JAVMA. 1997; vol. 210 (10):. 1451-1457.



Singh, J.A., 2012. Stem cells and other innovative intra---articular therapies for osteoarthritis: what does the future hold? BMC Med. 10 (1), 44.

Skinner, O.T., Kim, S.E., Lewis, D.D. & Pozzi, A., 2013. In vivo femorotibial subluxation during weight-bearing and clinical outcome following tibial tuberosity advancement for cranial cruciate ligament insufficiency in dogs. Vet J. 196 (1), 86-91.

Souza, A.N., Tatarunas, A.C. & Matera, J.M., 2014. Evaluation of vertical forces in the pads of Pitbulls with cranial cruciate ligament rupture. BMC Vet Res. 10 (1), 51.

Taylor, B. M., Tipton, C.M., Adrian, M. & Karpovich P.V., 1966. Action of certain joints in the legs of the horse recorded electrogoniometrically. Am. J. Vet. Res. 27 (116), 85-89.

Tobias, Karen M., Johnston, Spencer A., 2011. Veterinary Surgery: Small Animal. 1st ed. Saunders. 2332.

Todhunter, R. & Johnston, S., 2006. Osteoartritis. En: Slatter, D. (ed.) Tratado de Cirugía en Pequeños Animales. 3ª ed. Buenos Aires, República Argentina: Inter--Médica.

Tomas, A., Marcellin-Little, D.J., Roe, S.C., Motsinger-Reif, A. & Lascelles, B.D., 2014. Relationship between mechanical thresholds and limb use in dogs with coxofemoral joint oa-associated pain and the modulating effects of pain alleviation from total hip replacement on mechanical thresholds. Vet Surg. 43 (5), 542-548.

Uhlir, C., Licka, T., KÜbber, P., Peham, C., Scheidl M. & Girtler, D., 1997. Compensatory movements of horses with a stance phase lameness. Equine Vet J. Supplement. 29 (S23), 102-105.

Upchurch, D.A., Renberg, W.C., Roush, J.K., Milliken, G.A. & Weiss, M.L., 2016. Effects of administration of adipose-derived stromal vascular fraction and



platelet-rich plasma to dogs with osteoarthritis of the hip joints. Am J Vet Res. 77 (9), 940-945.

Van Heel, M.C., Moleman, M., Barneveld, A., Van Weeren, P.R. & Back, W., 2005. Changes in location of Centre of pressure and hoof-unrollment pattern in relation to an 8-week shoeing interval in the horse. Equine Vet J. 37 (6), 536–540.

van Weeren, P.R., van den Bogert, A.J. & Barneveld, A., 1990. A quantitative analysis of skin displacement in the trotting horse. Equine Vet J. 22 (9), 101-109.

Vilar, J.M., Miró, F., Santana, A. & Spinella, G., 2010a. Biokinematics Under Competitive Racing Conditions in Young Standardbred Trotter Horses: a Preliminary Report. 30(8), 432-435. doi: 10.1016/j.jevs.2010.07.002.

Vilar, J.M., Ramirez, G., Spinella, G. & Martinez, A., 2010b. Kinematic characteristics of myositis ossificans of the semimembranosus muscle in a dog. Can Vet J. 51 (3), 289-292.

Vilar, J.M., Morales, M., Santana, A., Batista, M., Miró, F. & Spinella, G., 2013a. Long-term valuation of oral mavacoxib in osteoarthrosic dogs using force platform analysis. Pakistan Vet J. 33 (2), 229-233.

Vilar, J. M., Rubio, M., Carrillo, J.M., Domínguez, A.M., Mitat, A. & Batista, M., 2016a. Biomechanic characteristics of gait of four breeds of dogs with different conformations at walk on a treadmill. J Appl Anim Res. 44 (1), 252-257.

Vilar, J.M., Cuervo, B., Rubio, M., Sopena, J., Domínguez, J.M., Santana, A., Carrillo, J.M., 2016b. Effect of intraarticular inoculation of mesenchymal stem cells in dogs with hip osteoarthritis by means of objective force platform gait analysis: concordance with numeric subjective scoring scales. BMC Vet Res. 12(1), 223.

Walton, M.B., Cowderoy, E., Lasclles, D. & Innes, J. F., 2013. Evaluation of construct and criterion validity for the "Liverpool osteoarthritis in dogs" (LOAD)



clinical metrology instrument and comparison to two other instruments. PLoS ONE. 8(3), e58125. doi:10.1371/journal.pone.0058125

Walton, M.B., Cowderoy, E.C., Wustefeld-Janssens, B., Lascelles, B.D. & Innes, J.F., 2014. Mavacoxib and meloxicam for canine osteoarthritis: a randomised clinical comparator trial. Vet Rec. 175 (11), 280.

Watanabe, T., Saito, H., Koike, E. & Nitta, K., 2011. A preliminary test of measurement of joint angles and stride length with wireless inertial sensors for wearable gait evaluation system. Comp Intel Neurosc. 975193, 12.

Wernham, B.G.J., Trumpatori, B., Hash. J., Lipsett, J., Davidson, G., Wackerow, P., Thomson, A. & Lascelles, B., 2011. Dose reduction of meloxicam in dogs with osteoarthritis-associated pain and impaired mobility. J Vet Intern Med. 25 (6), 1298-1305.

Whitehair, J.G., Vasseur, P.B., Willits, N.H., 1993. Epidemiology of cranial cruciate ligament rupture in dogs. J Am Vet Med Assoc. 203, 1016-1019.

Winter, D.A., 1991. Kinetics. En: The biomechanics and motor control of human gait: normal, elderly and pathological. 2nd ed. Waterloo, University of Waterloo Press. 35-53.

Winter, D.A., Prince, F., Frank, J.S., Powell, C. & Zabjek K.F., 1996. Unified theory regarding A/P and M/L balance in quiet stance. J Neurophysiol. 75 (6), 2334-2343.

Wu, W., Chen, F., Liu, Y., Ma, Q. & Mao, T., 2007. Autologous injectable tissue--engineered cartilage by using platelet---rich plasma: experimental study in a rabbit model. J Oral Maxillofac Surg. 65 (10), 1951--1957.

Wucherer, K.L., Conzemius, M.G., Evans, R. & Wilke, V.L., 2013. Short-term and long-term outcomes for overweight dogs with cranial cruciate ligament rupture treated surgically or nonsurgically. J Am Vet Med Assoc. 242 (10), 1364-1372.

BIBLIOGRAFÍA



Yarak, S. & Okamoto, O.K., 2010. Human adipose---derived stem cells: current challenges and clinical perspectives. An Bras Dermatol. 85 (5), 647--656.

