

Markus Amann and Jose A. L. Calbet

J Appl Physiol 104:861-870, 2008. First published Oct 25, 2007; doi:10.1152/jappphysiol.01008.2007

You might find this additional information useful...

This article cites 128 articles, 82 of which you can access free at:

<http://jap.physiology.org/cgi/content/full/104/3/861#BIBL>

This article has been cited by 1 other HighWire hosted article:

Is peripheral locomotor muscle fatigue during endurance exercise a variable carefully regulated by a negative feedback system?

S. Marcora

J. Physiol., April 1, 2008; 586 (7): 2027-2028.

[Full Text] [PDF]

Updated information and services including high-resolution figures, can be found at:

<http://jap.physiology.org/cgi/content/full/104/3/861>

Additional material and information about *Journal of Applied Physiology* can be found at:

<http://www.the-aps.org/publications/jappl>

This information is current as of April 14, 2008 .

HIGHLIGHTED TOPIC | *Fatigue Mechanisms Determining Exercise Performance*

Convective oxygen transport and fatigue

Markus Amann¹ and Jose A. L. Calbet²

¹John Rankin Laboratory of Pulmonary Medicine, University of Wisconsin-Madison Medical School, Madison, Wisconsin; and ²Department of Physical Education, University of Las Palmas de Gran Canaria, Spain

Amann M, Calbet JA. Convective oxygen transport and fatigue. *J Appl Physiol* 104: 861–870, 2008. First published October 25, 2007; doi:10.1152/jappphysiol.01008.2007.—During exercise, fatigue is defined as a reversible reduction in force- or power-generating capacity and can be elicited by “central” and/or “peripheral” mechanisms. During skeletal muscle contractions, both aspects of fatigue may develop independent of alterations in convective O₂ delivery; however, reductions in O₂ supply exacerbate and increases attenuate the rate of accumulation. In this regard, peripheral fatigue development is mediated via the O₂-dependent rate of accumulation of metabolic by-products (e.g., inorganic phosphate) and their interference with excitation-contraction coupling within the myocyte. In contrast, the development of O₂-dependent central fatigue is elicited 1) by interference with the development of central command and/or 2) via inhibitory feedback on central motor drive secondary to the peripheral effects of low convective O₂ transport. Changes in convective O₂ delivery in the healthy human can result from modifications in arterial O₂ content, blood flow, or a combination of both, and they can be induced via heavy exercise even at sea level; these changes are exacerbated during acute and chronic exposure to altitude. This review focuses on the effects of changes in convective O₂ delivery on the development of central and peripheral fatigue.

oxygenation; hypoxia; blood flow; exercise; hyperoxia

WHAT IS FATIGUE?

EXERCISE-INDUCED FATIGUE is defined as the reversible reduction in force- or power-generating capacity of the neuromuscular system (16, 47). Potentially, fatigue may originate in any of the elements that intervene in the planning, execution, and/or control of motor tasks. Classically, fatigue has been classified as “central” or “peripheral” (44, 46, 52, 53). Peripheral fatigue comprises processes at or distal to the neuromuscular junction, e.g., biochemical changes within the working muscle, leading to a failure to respond to neural excitation. Controversy exists about the extent to which significant failure of neuromuscular transmission, i.e., failure in the transmission of the neural signal, occurs during muscle fatigue in *healthy* humans (53). However, the reduction in force- or power-generating capacity can result from inadequate, or suboptimal, muscle activation—often, but not exclusively, in combination with failure of contractile mechanisms—resulting from a failure of the central nervous system (CNS) to excite—or “drive”—the motoneurons adequately (central fatigue). Several mechanisms that are not mutually exclusive have been suggested to underlie central and peripheral fatigue (2, 34, 44, 46, 52, 61, 95, 106), and the impaired motor performance is the result of a combination of various acute determinants of fatigue rather than just one aspect. Further-

more, insufficient O₂ delivery [blood flow × arterial O₂ content (Ca_{O₂})] may elicit fatigue by affecting both central and peripheral mechanisms (68, 105, 113).

The focus of this review is on factors causing changes in convective O₂ transport and their functional consequences on fatigue, rather than a discussion of diverse cellular mechanisms. We extend the definition of fatigue to include, as suggested by Enoka and Stuart (44), acute impairments of performance during exercise—as reflected in the failure to maintain a given force or power output (i.e., task failure)—which are attributable to mechanisms susceptible to alterations in convective O₂ transport.

How to Quantify Fatigue?

Methods to evaluate fatigue are numerous (23, 127, 133) and include techniques used to quantify biochemical changes known to cause reductions in force-generating capacity (41, 65, 71, 119). Assessment of neuromuscular functions are usually made *before* and immediately *after* fatiguing exercise, and the discrepancy between pre- vs. postexercise measures is used to quantify peripheral fatigue. These methods rely on the assumption that changes in neuromuscular function reflect fatigue induced during exercise and include either the assessment of effort-dependent contraction force [i.e., maximal voluntary contraction (MVC) force] or effort-independent force generated by evoked muscle contractions (i.e., force-frequency properties). Since the assessment of MVC force depends on the subject's voluntary effort (a major contributor to central fatigue), this method is not capable of exclusively quantifying

Address for reprint requests and other correspondence: M. Amann, The John Rankin Laboratory of Pulmonary Medicine, 4245 Medical Science Center, 1300 University Ave., Madison, WI 53706 (e-mail: amann@wisc.edu).

peripheral fatigue, a disadvantage that is eliminated with evoked contractions. The determination of voluntary muscle activation (97) during MVC maneuvers can be used to reveal the contribution of the central component of fatigue. Peripheral fatigue is often quantified based on measurements of isometric contractions of a single muscle *before* and *after* exercise, whereas dynamic contractions of many muscles were used *during* exercise.

To determine the development of fatigue *during* exercise, electromyography (EMG) has been utilized (8, 13, 16, 42). Although electromyography has its shortcomings (11), it can be used to estimate the development of both central (16, 17, 42, 44, 52, 124) and peripheral (13, 42) fatigue *during* exercise. In the case of peripheral fatigue, EMG activity/neural drive at a constant work output increases progressively over time, presumably to compensate for fatiguing muscle fibers (Fig. 1). In the case of central fatigue, a fall in muscle force or power is secondary to a centrally mediated reduction in motor drive (reduction in EMG activity), resulting in reduced motor unit recruitment. Finally, the rate of development of fatigue has

been estimated by means of the time to task failure—a typical indicator of muscular performance, which is defined as the point where the failure to maintain a given force or power (i.e., task) occurs.

CONVECTIVE O₂ TRANSPORT AND FATIGUE

Increases and decreases in systemic O₂ transport affect muscular performance and the rate of development of both central and peripheral fatigue. Blunted O₂ delivery exaggerates this rate, whereas an augmentation in O₂ delivery attenuates the rate of development (1, 6–9, 18, 24, 25, 32, 39, 43, 49–51, 71, 72, 81, 84, 85, 92, 107, 116–118, 120, 128, 136; see Figs. 1 and 2). The mechanisms by which changes in convective O₂ transport modify the rate of development of fatigue are complex, because the influences of altered O₂ supply occur throughout the organism. However, the highly sensitive effects of O₂ delivery on the rate of fatigue development are generally well accepted although not supported by all studies (14, 19, 22, 30, 45, 74, 82). By comparing the rate of decline in force-generating capacity during submaximal dynamic knee extensions in normoxia and hypoxia, Fulco et al. (50) reported a markedly faster rate of development of peripheral fatigue in hypoxia; however, the difference in exercise-induced reduction in MVC force was not significant between the two conditions until *minute 4* of exercise. These authors suggest that the failure to show greater reduction in MVC force under hypoxic conditions might be due to the use of high-intensity exercise of a duration shorter than 4 min (22, 30) or sustained static contractions in which muscle ischemia is likely to have markedly attenuated the effects of a reduced O₂ delivery on local muscle metabolism (19, 22). In support, studies on isolated canine muscle during which supramaximal electrical stimulation was used to evoke muscle contractions for 2–3 min under conditions of augmented as well as blunted O₂ delivery also reported no difference in end-exercise fatigue (73, 74). Thus under some experimental conditions, reduced oxygen delivery may accelerate the process leading to fatigue only when the duration of the exercise exceeds a certain threshold (>4 min). At lower exercise intensities under conditions of blunted CaO₂, compensatory increases in O₂ extraction fraction and blood flow are capable of maintaining adequate muscle O₂ supply and thus abolish or attenuate a potential difference in exercise-induced fatigue compared with normoxic conditions (12, 86, 87). At higher exercise intensities, cardiac output and leg blood flow approach their peak values and are no longer capable of compensating for the reduced CaO₂, and differences in fatigue resulting from alterations in O₂ delivery become more obvious.

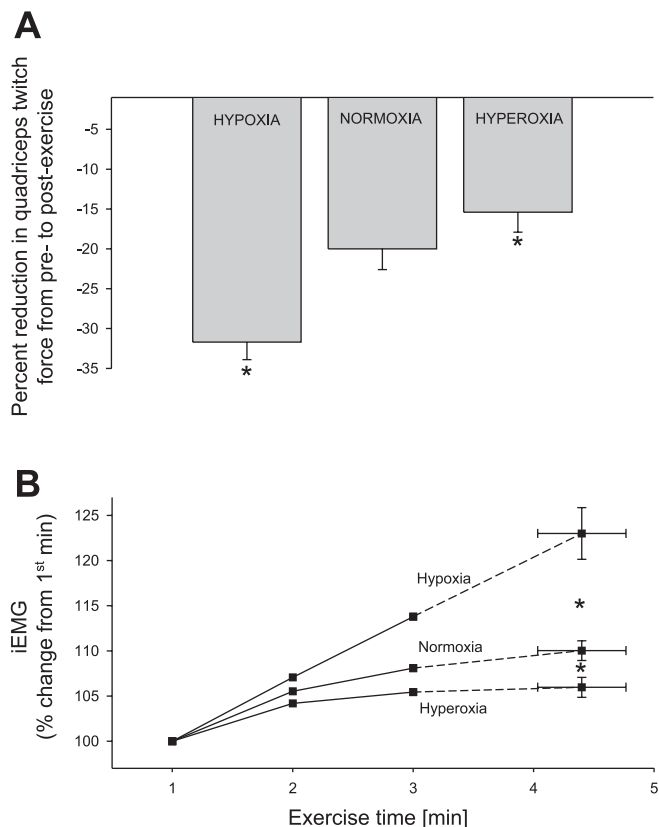


Fig. 1. Effects of constant-workload bicycling exercise for 4.5 ± 0.4 min at 314 ± 13 W ($\sim 94\%$ of normoxic maximal O₂ uptake) on the development of locomotor (quadriceps) muscle fatigue under various conditions of convective O₂ transport [Hypoxia: inspiratory O₂ fraction (F_IO₂) 0.15, S_aO₂ 82%, arterial O₂ content (CaO₂) 18 ml O₂/dl; Normoxia: F_IO₂ 0.15, S_aO₂ 95%, CaO₂ 21 ml O₂/dl; Hyperoxia: F_IO₂ 1.0, S_aO₂ 100%, CaO₂ 24 ml O₂/dl]. A: exercise-induced reduction in potentiated quadriceps twitch force, which was assessed via supramaximal magnetic femoral nerve stimulation before and again 2 min after exercise. B: myoelectrical activity [integrated EMG (iEMG)] of vastus lateralis *during* exercise under the three different conditions. Values are normalized to the mean of the first minute. Mean value for iEMG during each muscle contraction (cycle revolution) was calculated and averaged over each 60-s period. Adapted from Amann et al. (8). * $P < 0.05$ vs. Normoxia.

Peripheral Fatigue

Alterations in O₂ delivery to the working muscles affect the development of peripheral fatigue during *whole body* exercise via its effects on relative exercise intensity and changes in intracellular metabolism. Both of these factors alter the rate of accumulation of metabolites (e.g., H⁺, phosphates) known to cause failure of excitation-contraction coupling (ECC) within the muscle fiber, which has been identified as the main factor that evokes loss of tension development during the fatigue process occurring under conditions of high-intensity exercise (3, 46, 89, 91, 135). Briefly, ECC failure has been associated with an accumulation of metabolites that can directly inhibit

the contractile apparatus (48, 55, 125) or disrupt the Ca^{2+} release and uptake pathways in the sarcoplasmic reticulum (2, 41, 46). Although the rate of development of peripheral fatigue is faster with reduced—and slower with increased— O_2 delivery to the working muscle, the key underlying mechanism of fatigue (i.e., perturbations of ECC via diminished Ca^{2+} release and inhibition of the contractile apparatus) are the same in conditions from hypoxia to hyperoxia (123).

Relative exercise intensity. Alterations in convective O_2 transport during *whole body* exercise precipitate a change in peak exercise capacity [peak workload and maximal O_2 uptake ($\dot{V}\text{O}_{2\text{max}}$)] and therefore a shift of a given *absolute* work load to a different *relative* intensity of exercise, i.e., higher in conditions of a reduced O_2 delivery to the working muscles and lower with an increased O_2 delivery (6–8, 51, 85, 94, 116, 117, 126). An increase in relative exercise intensity subsequent to a reduction in O_2 delivery is known to increase the percentage of type II fibers activated during dynamic muscular contractions (96). Furthermore, reductions in systemic and muscular O_2 delivery may affect fiber type contribution by attenuating the sensitivity of type III/IV muscle afferents to a given stimulus [their stimulation is associated with a preferential recruitment of O_2 -dependent type I muscle fibers (10)]. Taken together, more type II fibers are activated under conditions of reduced O_2 transport to the working muscle to maintain a given constant workload. Since type II fibers are associated with an increased rate of accumulation of metabolites known to cause fatigue compared with type I fibers (42), the O_2 delivery-dependent change in relative exercise intensity might account for a significant proportion of the exaggerated rate of development of peripheral fatigue under conditions of reduced O_2 delivery to the muscle. This effect of altered O_2 delivery on the rate of development of peripheral muscle fatigue via alterations in relative work intensity as described above is not applicable when isolated isometric muscular contractions are utilized. This is due to the fact that maximal force output of the muscle under investigation is not affected by alterations in convective O_2 delivery under baseline resting conditions; hence, submaximal isometric muscular contractions at a given absolute force output are also carried out at the same relative work intensity when convective O_2 delivery is altered (35, 49, 50, 54, 81, 107, 136), and the difference in fatigue is mainly due to the effects of altered O_2 delivery on the rate of accumulation of metabolites known to cause fatigue.

Intracellular metabolism. Alterations in convective O_2 transport affect intracellular metabolism, including the rate of accumulation of metabolites that have been identified as major determinants of peripheral fatigue (1, 46, 71). Since the rate of accumulation of protons (1, 71, 75) and the hydrolysis of phosphocreatine and concomitant cytoplasmic inorganic phosphate (P_i) accumulation (65, 66, 71) are faster under conditions of reduced—and slower under conditions of increased— O_2 delivery to the working muscle, both metabolites have been considered to play a key role in the development of peripheral fatigue in conditions of altering systemic O_2 delivery. Although an increased $[\text{H}^+]$ is traditionally suggested as the main cause of skeletal muscle fatigue, recent *in vitro* studies have questioned the deleterious role of $[\text{H}^+]$ in metabolic fatigue of mammalian muscles at physiological temperatures (21, 108, 109), and the question regarding the relative contribution of protons to muscle fatigue remains controversial (88). More

recently, increased P_i rather than acidosis appears to be the most important cause of peripheral fatigue during high-intensity exercise (134). Briefly, cytoplasmic P_i is thought to enter the sarcoplasmic reticulum and bind to Ca^{2+} to form a precipitate (CaP_i), thus reducing the amount of releasable Ca^{2+} , which results in perturbations of ECC (33, 41, 48). Overall, P_i appears to offer a more attractive explanation of the different rates of development of peripheral fatigue with alterations in systemic O_2 transport (134). For a more detailed reading on determinants of peripheral fatigue, we refer the reader to other reviews published as a part of this highlighted topic.

Causes of Alterations in Convective O_2 Transport and Associated Functional Consequences on Peripheral Fatigue

Alterations of convective O_2 transport to the working muscles in *healthy* humans are the result of changes in CaO_2 and/or limb blood flow. First, alterations in CaO_2 can result from changes in the inspiratory O_2 fraction ($\text{F}_{\text{I}\text{O}_2}$), inspiratory partial pressure of oxygen ($\text{P}_{\text{I}\text{O}_2}$; i.e., exposure to altitude), or hemoglobin concentration (i.e., anemia). In some, but not all, well-trained athletes, significant reductions in CaO_2 can—even under sea-level normoxic conditions—result from a substantial high-intensity exercise-induced widening of the alveolar-arterial PO_2 difference combined with a blunted hyperventilatory response to exercise, resulting in a decrease in arterial PO_2 (PaO_2) and correspondingly arterial hemoglobin saturation (SaO_2) from rest, a phenomenon referred to as exercise-induced arterial hypoxemia (EIAH; see Ref. 38). It is important to notice that *severe* EIAH ($\text{SaO}_2 < 88\%$) occurs only in a minority of athletes due to a reduction in PaO_2 ; more commonly, EIAH results from a rightward shift in the oxyhemoglobin dissociation curve mediated by acidosis and temperature (38). A mild level of EIAH (SaO_2 93–95%) is observed in most humans during sea level exercise, and a $>3\%$ reduction in SaO_2 from rest has been shown to have a significant detrimental effect on $\dot{V}\text{O}_{2\text{max}}$ (63). Second, hyperventilation of heavy sustained exercise ($>85\% \dot{V}\text{O}_{2\text{max}}$) causes substantial increases in respiratory muscle work, leading to diaphragm and expiratory muscle fatigue (37). Accumulation of metabolites in these muscles activates unmyelinated group IV phrenic afferents, which in turn increase sympathetic vasoconstrictor activity in the working limb via a supraspinal reflex (37). The result is a work of breathing-induced reduction in limb blood flow, a corresponding reduction in O_2 delivery to the working muscles, and (presumably) an increase in blood flow to the respiratory muscles, indicating a competitive relationship for a limited cardiac output (62).

As a side note, a *severely* reduced convective O_2 transport to the working muscle via EIAH is experienced only by a subgroup of highly trained endurance athletes and can develop even at submaximal exercise intensities, whereas a reduction in convective O_2 transport via respiratory muscle fatigue and the subsequent reduction in blood flow to the working limb muscles occur in healthy untrained and trained subjects but only at sustained, high-intensity endurance exercise ($>85\% \dot{V}\text{O}_{2\text{max}}$; see Ref. 7).

CaO_2 . Alterations in CaO_2 are followed by reciprocal changes in $\dot{V}\text{O}_{2\text{max}}$ and endurance performance (27); however, the effect on endurance is more accentuated than that on $\dot{V}\text{O}_{2\text{max}}$, especially after acclimatization to high altitude (80, 94, 122, 129).

The latter is likely due to a profound effect of Ca_{O_2} on muscle metabolism during submaximal exercise. For example, the blood lactate accumulation curve is shifted to the left, meaning that for a given absolute exercise intensity, the glycolytic rate and associated rate of accumulation of metabolites known to cause fatigue (e.g., H^+) is increased (28, 86). Even a relatively small reduction in Sa_{O_2} (5–8% from rest) associated with EIAH affects the magnitude of locomotor muscle fatigue during high-intensity endurance exercise ($\geq 90\% \dot{V}O_{2max}$). Romer et al. (117) used two cycling exercise sessions at identical power output and duration, during which EIAH was either allowed to develop (Fi_{O_2} 0.21; control) or prevented via small increases in Fi_{O_2} (0.24–0.30). By preventing EIAH, end-exercise peripheral locomotor muscle fatigue was reduced by more than one-half compared with the level of peripheral quadriceps fatigue measured after the control trial (comparison of pre- vs. postexercise force-frequency curves). These effects of EIAH on locomotor muscle fatigue are reflected in a significant limitation to exercise performance (5-km cycling time trial) (6). Ca_{O_2} , and thus O_2 delivery to the working limbs, was increased by $\sim 8\%$ when the exercise-induced fall in Sa_{O_2} was prevented during the time trial via increases in Fi_{O_2} . Preventing this drop in Sa_{O_2} during the race resulted in a 2–5% reduction in the time to completion and an up to 5% increase in mean power output. However, although ventilatory muscle work was not directly measured in the study by Romer et al. (117), minute ventilation was significantly reduced when EIAH was prevented via increased Fi_{O_2} , and this alleviation in respiratory muscle work might have attenuated respiratory muscle fatigue, which in turn might have resulted in a higher limb blood flow compared with the condition when EIAH developed. Therefore, by preventing EIAH, O_2 delivery to the working muscle might have been increased—and therefore locomotor muscle fatigue reduced—due to the higher Sa_{O_2} (i.e., Ca_{O_2}) and the higher limb blood flow.

More recently Amann et al. (7) were able to isolate the independent effects of the ventilatory muscle work (and by extension limb blood flow) vs. Ca_{O_2} during heavy-intensity bicycle exercise in acute hypoxia on locomotor muscle fatigue (see Fig. 2). By contrasting normoxic vs. hypoxic Ca_{O_2} (Fi_{O_2} 0.21 and 0.15; $Ca_{O_2} \sim 20$ ml O_2 /dl and ~ 17 ml O_2 /dl, respectively) at equal work rates and durations of exercise with very low levels of ventilatory work (via the use of a mechanical ventilator), they showed that low Ca_{O_2} —independent of any influence of work of breathing—exacerbated low-frequency quadriceps fatigue by over 50%.

Blood flow to working muscle. Increases in blood flow to the electrically stimulated in situ canine muscle per se can attenuate—and reductions exacerbate—the rate of fatigue during long-term contractions through a mechanism independent of alterations in O_2 and substrate delivery but probably related to alterations in intracellular environment, i.e., washout of local metabolites (12). However, changes in the rate of fatigue development due to alterations in blood flow are mainly via its effects on O_2 delivery rather than due to blood flow changes alone. This statement is based on the work by Hogan et al. (69, 70, 72), which is discussed in detail in the excellent review by Hepple (68). Briefly, the most compelling evidence stems from a study on electrically stimulated canine gastrocnemius muscle that yielded about 60% of $\dot{V}O_{2max}$ for this model (69). Under conditions of ischemia, muscle force rapidly fell; it recovered quickly upon restoration of normal blood flow and O_2 delivery, but not when normal blood flow alone was restored with deoxygenated blood (69). It is important to remark that upon reoxygenation the recovery of force was incomplete, remaining 24–33% below the control value (69).

Respiratory muscle work can cause skeletal muscle fatigue by reducing blood flow (see above), and thus O_2 delivery, to the working limb (62). The isolated effects of work of breathing-induced changes in limb blood flow on the development of

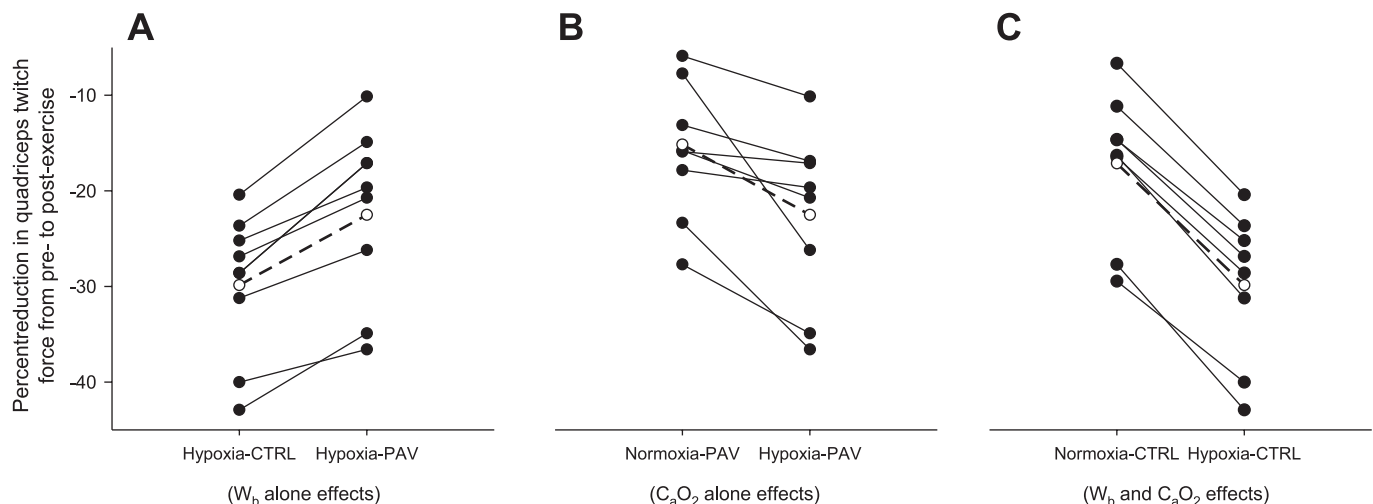


Fig. 2. Individual (●) and group mean (○) effects of constant workload bicycling exercise on the development of locomotor (quadriceps) muscle fatigue. All trials were conducted at identical work rate (273 ± 6 W) and identical duration (8.6 ± 0.2 min). A mechanical proportional assist ventilator (PAV) known to reduce inspiratory muscle work (W_b) by up to 70%, thereby increasing limb blood flow by more than 5% from control (62), and exposure to acute hypoxia (Fi_{O_2} 0.15, $Sa_{O_2} \sim 82\%$, $Ca_{O_2} \sim 17$ ml O_2 /dl) were used to isolate and separate the independent effects of the two determinants of limb O_2 delivery (limb blood flow \times Ca_{O_2}) on whole-body exercise-induced locomotor muscle fatigue. A: isolated effects of W_b (i.e., limb blood flow) in hypoxia on quadriceps fatigue. W_b was reduced by about 70%, whereas Ca_{O_2} was unchanged (~ 17 ml O_2 /dl) from Hypoxia-Control (-CTRL) to Hypoxia-PAV. B: isolated effects of Ca_{O_2} on quadriceps fatigue. Ca_{O_2} was reduced from ~ 20 to ~ 17 ml O_2 /dl, whereas W_b , and therefore limb blood flow, was unchanged from Normoxia-PAV to Hypoxia-PAV. C: exacerbating effects of reductions in Ca_{O_2} ($\sim 15\%$) combined with increases in W_b ($\sim 36\%$)—and therefore reductions in limb blood flow—on peripheral fatigue when exercising in hypoxia vs. normoxia. From Amann et al. (7).

peripheral locomotor muscle fatigue—independent of changes in CaO_2 —were recently quantified by means of two trials of strenuous ($>90\% \dot{V}_{\text{O}_{2\text{max}}}$) cycling exercise of identical work rate, duration, and CaO_2 (7, 118; see Fig. 2). When the exercise trial was performed with a mechanical ventilator capable of reducing force output of the inspiratory muscles by up to 70%, limb blood flow was increased by 5% or more and leg O_2 uptake (\dot{V}_{O_2}) by 3% compared with the control conditions without ventilatory assistance (62). Exercise-induced peripheral quadriceps fatigue (pre- vs. postexercise force-frequency curves) was reduced by $\sim 30\%$ compared with control when both trials were performed in normoxia (118) and by over 35% when both trials were performed in acute hypoxia (7). In contrast, enhancing inspiratory muscle work by about 80% greater than during the control trial (via resistive loading)—and thus reducing limb blood flow by over 10% from control—almost doubled the amount of end-exercise quadriceps fatigue (118). Finally, it is important to mention that the work of breathing-induced change in limb blood flow and the consequential effect on peripheral fatigue is only apparent at exercise intensities greater than 80% of $\dot{V}_{\text{O}_{2\text{max}}}$ (7), since it has been shown that those intensities, sustained to exhaustion, are necessary to elicit diaphragm fatigue, triggering the metaboreflex as described above (37). The documented effects of limb blood flow—as affected by normally occurring work of breathing—on the development of locomotor muscle fatigue during high-intensity whole body exercise are reflected in significant limitations to exercise performance (7, 64). The time to the limit of exhaustion was increased by $\sim 14\%$ when the normally occurring work of breathing was reduced by about one-half during strenuous constant workload cycling exercise in normoxia (64); when normally occurring work of breathing was reduced by about 70% during strenuous constant workload cycling exercise in acute hypoxia, time to exhaustion was increased by $\sim 16\%$ (7).

Central Fatigue

Central fatigue has been related to changes in extracellular neurotransmitter levels or exercise-induced alterations in the activity of different neurotransmitter systems (34, 95). However, under some experimental conditions it has been shown that central fatigue can also be elicited by low brain oxygenation, i.e., by insufficient O_2 delivery and/or low pressure gradient to drive the diffusion of O_2 from the capillaries to the mitochondria. The CNS is highly sensitive to reductions in oxygenation, and consequently cerebral oxygenation is strongly defended by several homeostatic mechanisms (98). Low brain oxygenation may cause a mismatch between brain O_2 demand and O_2 supply, leading to reduced interstitial (PtiO_2) and cellular Po_2 . The critical Po_2 levels in the brain capillaries and mitochondria remain unknown; there are no data on PtiO_2 in the cerebral white matter of healthy humans (83). In severely head-injured patients without intracranial hypertension or decreased cerebral perfusion, PtiO_2 ranges between 25 and 30 Torr (93). At a jugular venous oxygen saturation (SjvO_2) of 50% (considered clinically as the lower tolerable limit), brain PtiO_2 ranged from 3 to 12 Torr (the regression curve's best fit value being 8.5 Torr; see Ref. 83). An SjvO_2 of 30% was associated with a brain PtiO_2 close to 0 Torr, and an SjvO_2 of 70% corresponded to a brain PtiO_2 of 20

Torr (83). In one report on a series of 22 patients with severe head injuries, five of six patients with PtiO_2 values of 5 Torr or below died or stayed vegetative (132). In a later study these authors reported that even a local PtiO_2 value of less than 10 Torr for more than 10 min carried a statistically significant risk of death (131). Thus it is likely that if the PtiO_2 approaches or falls below 10 Torr during exercise, central fatigue will ensue due to impaired neuronal function, which will develop more easily the longer the low PtiO_2 is maintained.

Low PtiO_2 during exercise. Reduced PtiO_2 may be caused by reduced O_2 delivery, which may or may not be accompanied by hypoxemia (low PaO_2). Brain oxygen delivery is determined by the product of $\text{CaO}_2 \times$ cerebral blood flow (CBF), and both factors are tightly controlled. A reduction in CBF is compensated by increasing cerebral O_2 extraction up to a limit of 0.60, beyond this point lowering CBF results in a reduction in cerebral \dot{V}_{O_2} (101). CBF is mainly regulated depending on brain metabolism, arterial blood pressure, CaO_2 , Po_2 , and PCO_2 (90, 112). Although some controversy remains, CBF increases with exercise intensity up to $\sim 60\%$ of $\dot{V}_{\text{O}_{2\text{max}}}$, after which it decreases toward resting values and sometimes below resting values (67, 100). The reduction in CBF above 60% of $\dot{V}_{\text{O}_{2\text{max}}}$ has been attributed to the development of hyperventilation-induced hypocapnia (15, 115), an effect that is exacerbated during exercise with hyperthermia (103), in part due to enhanced cerebral CO_2 reactivity (115). During exercise at a constant intensity eliciting fatigue between 5 and 10 min, middle cerebral artery blood velocity increases to a value 20–30% higher than basal at 90 s of exercise, and thereafter it declines to baseline values (59). Brain oxygenation [measured with near-infrared spectroscopy (NIRS)] shows a continuous decline to reach a nadir at exhaustion, while fractional O_2 extraction [calculated as $(\text{CaO}_2 - \text{SjvO}_2)/\text{CaO}_2$] is maintained close to 0.38, and it increases to 0.47 at exhaustion (59). When the exercise is started with a 1°C higher core temperature, exercise time is reduced and brain deoxygenation is accelerated, but at exhaustion similar levels of deoxygenation and brain O_2 extraction are reached compared with control conditions (59). At first glance, it seems that during high-intensity exercise at a constant intensity, brain deoxygenation could play a role as a mechanism causing central fatigue. Nevertheless, in these experiments the limit of cerebral O_2 extraction was not reached, and the calculated cerebral \dot{V}_{O_2} if anything increased toward the end of the exercise (59). Thus it seems that the reduction in brain oxygenation was not low enough to impair exercise capacity in these experiments.

Exercise-induced elevations in CBF occur primarily in the most active areas of the brain (36, 110). Pott et al. (110) showed that during unilateral handgrip exercise, middle cerebral artery blood velocity increased in the contralateral artery more than in the ipsilateral side. Logically, the areas activated during exercise, which have a higher O_2 demand, should be more vulnerable to a reduction in oxygen delivery and/or PaO_2 during exercise. In agreement, it has been reported that brain oxygenation, measured with NIRS, declines at intensities above the respiratory compensation point, i.e., when arterial PCO_2 (PaCO_2) starts to decrease during progressive exercise to exhaustion (15). However, maneuvers that prevent a reduction in brain oxygen delivery during peak exercise at sea level, such as the prevention of hypocapnia (60) or the acute administration of a hyperoxic gas mixture (FiO_2 0.3–1.0) at exhaustion (9)

do not prolong exercise time, suggesting that during peak exercise in normoxia, the deficit in cerebral oxygenation per se does not reach a level low enough to cause termination of exercise. An exception to this behavior may be the elite athlete (102) and other humans who show arterial hypoxemia and desaturation during exercise at sea level. For example, elite rowers, who showed marked arterial hypoxemia and 17% cerebral venous O₂ desaturation during exercise at sea level, attained higher performance when the exercise was performed with mild hyperoxia (F_IO₂ 0.30), which was high enough to prevent arterial and cerebral desaturation (see Fig. 3) (102). However, since the whole exercise bout was performed with increased F_IO₂, this likely also delayed the development of peripheral fatigue (117). Moreover, the level of brain deoxygenation shown in these experiments is rather low compared with other studies (59), meaning that these subjects could likely tolerate even a higher degree of brain deoxygenation prior to exhaustion.

In conditions where the normal cardiac output response to exercise is blunted, such as in patients with atrial fibrillation (79), in subjects under β -blockade (77), or during exercise with hyperthermia (57, 58), the normal exercise-induced elevation of middle cerebral artery blood velocity is blunted (77, 79, 104). Although, theoretically, limited cerebral oxygenation could contribute to central fatigue during exercise with hyperthermia, other mechanisms are likely more important (31, 56, 130). In addition, it remains to be tested if preventing hypocapnia during exercise with hyperthermia results in enhanced cerebral perfusion and exercise capacity.

During exercise in severe acute hypoxia (F_IO₂ 0.105), peak exercise cardiac output is reduced—along with peak work rate—when the exercise is performed with a large muscle mass (24) but not when the exercise recruits only one leg. Due to the very low PaO₂ observed during whole-body exercise with severe acute hypoxia (~34 Torr), pulmonary ventilation is strongly stimulated, leading to very low PaCO₂ values (~25 Torr, i.e., 8 Torr less than during peak exercise in normoxia; see Ref. 24). Cerebral blood flow drops between 2 and 3% per each 1 Torr drop in PaCO₂ when Po₂ remains close to 100 Torr

(78), and even if the effect of low PaCO₂ on CBF may be attenuated by severe hypoxia (20, 111), the vasoconstricting effect of hypocapnia predominates over the vasodilatory action of hypoxemia (40, 111, 123). The reduced CBF combined with the reduction in CaO₂ causes brain deoxygenation, which is more accentuated during intense exercise (126). The situation is further complicated by the very low PaO₂, which may lead to PtiO₂ values close to or below 10 Torr in some areas of the brain. Strong support for the role of low brain oxygenation in fatigue during exercise in severe acute hypoxia has been provided by the finding that, at the point of task failure, acute reoxygenation allows for immediate resumption of exercise (see below; see Refs. 9, 24).

With altitude acclimatization, cerebral oxygenation at peak exercise is restored to sea level values due to higher cerebral blood flow and CaO₂ (20, 99). Thus it is unlikely that insufficient brain oxygenation contributes to cause central fatigue during exercise at moderate altitude in acclimatized humans. However, at extreme altitudes the low PaO₂ may cause a low PtiO₂ and by this mechanism limit O₂ diffusion in some regions of the CNS, even in the altitude-acclimatized human (76). The hypothesis that insufficient brain oxygenation contributes to fatigue during maximal exercise in chronic hypoxia is supported by the fact that in well-acclimatized humans at 5,000–5,260 m, acute reoxygenation at peak exercise enables the subjects to continue the exercise and even to increase workload (25, 82).

Is central command affected by hypoxia? The ability to generate maximal power as well as maximal force during brief efforts is preserved in severe acute (26) as well as chronic hypoxia (121). Similarly, various studies indicate the absence of a central mechanism limiting small muscle mass exercise in hypoxia (49, 54, 81, 107, 136). However, more recently it has been reported that during severe acute hypoxia (F_IO₂ 0.10) handgrip MVC force is reduced, while maximal finger-typing frequency is not affected (114).

It has been suggested that hypoxia could also blunt the normal activation of the vasomotor areas of the brain, establishing a lower ceiling for cardiac output and heart rate (24). In turn, the latter could also cause peripheral fatigue due to reduced O₂ delivery to the locomotor and respiratory muscles, enhancing inhibitory feedback on central command.

The effects of low brain oxygenation during submaximal, long-duration whole body exercise on central command remain unknown. However, it has been argued that sensory afferent feedback, originating in the fatiguing locomotor muscle, to the CNS is one of the key determinants of the conscious (and/or subconscious) regulation of central motor drive (i.e., exercise performance), suggesting a strong link between “peripheral” (biochemical changes *within* the muscle) and “central” (reductions in CNS motor drive to the working muscle) fatigue (4, 6). Amann and Dempsey (4) have proposed that the magnitude of this inhibitory neural feedback is proportional to the rate of development of peripheral locomotor muscle fatigue (i.e., fatigue-related metabolic byproducts), which in turn is highly sensitive to muscle O₂ delivery (Fig. 1; see Ref. 8). Consequently, the rate of peripheral fatigue development acts as a dose-dependent trigger of central fatigue. When 5-km cycling time trials (power output voluntarily adjustable) were performed at different levels of arterial oxygenation, central motor drive and muscle power output were upregulated with in-

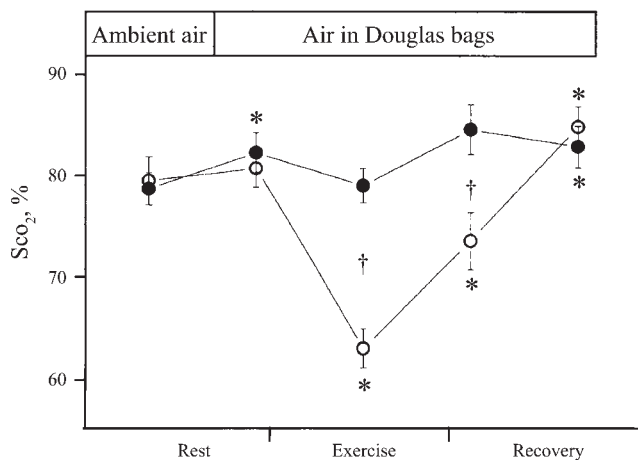


Fig. 3. Cerebral oxygenation (ScO₂) in ambient air, with an inspired O₂ fraction of 0.21 (●) or 0.30 (○) at rest, during 6 min of maximal rowing exercise, and at 2 and 4 min into recovery after exercise in elite rowers. Mean power output during the hyperoxic trial was 2.4% higher than during normoxia. **P* < 0.05 vs. control (rest); †*P* < 0.05 vs. trial with hyperoxia. From Nielsen et al. (102).

creased CaO_2 and downregulated with reduced CaO_2 ; however, the magnitude of peripheral muscle fatigue developed at end-exercise was identical (6; see Fig. 4). Since the rate of accumulation of peripheral fatigue (i.e., fatigue-causing metabolites) is enhanced with reduced CaO_2 and slowed with increased CaO_2 , the downregulation of central neural drive and consequently power output in the presence of reduced CaO_2 ensured that the rate of development of peripheral fatigue was slowed and prevented from exceeding a sensory tolerance limit (52). Hence, end-exercise peripheral locomotor muscle fatigue was identical between the time trials of various levels of CaO_2 and limited to a critical threshold (6). Based on this correlative evidence, peripheral locomotor muscle fatigue has been proposed as a sensed variable (6, 9, 29). However, how exactly the magnitude of fatigue and the associated metabolic milieu in the peripheral locomotor muscle is sensed and projected to higher brain areas where it might impose inhibitory effects on central motor drive remains to be solved (Fig. 5). These correlative data now need to be confirmed by a direct test of this hypothesis. This requires the determination of the true causal effect of specific interference with sensory input from working loco-

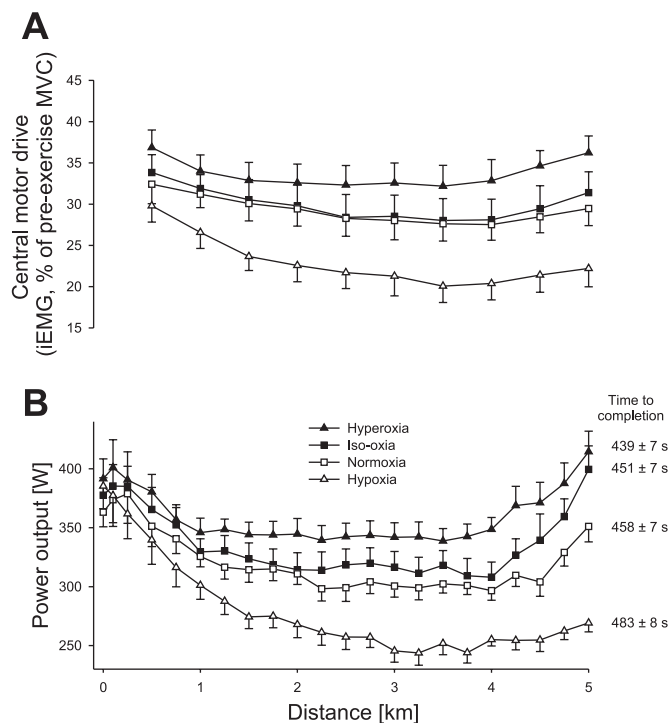


Fig. 4. Central motor drive and exercise performance during 5-km cycling time trials at various levels of CaO_2 . *A*: effects of various CaO_2 value on group mean central neural drive estimated via iEMG of vastus lateralis normalized to the iEMG obtained during pre-exercise maximal voluntary contractions (MVC). Each point represents mean iEMG of the preceding 0.5-km section. Mean iEMG during the time trial was significantly increased from hypoxia to hyperoxia ($P < 0.05$). *B*: group mean variations in power output during the 5-km time trial in four different conditions. End-exercise peripheral fatigue (estimated via changes in force output pre- vs. postexercise in response to supramaximal magnetic femoral nerve stimulation) was identical despite significant differences in exercise performance (i.e., time to completion and mean power output). Group mean power output was 356.5 ± 12.5 W, 331.0 ± 12.9 W, 313.8 ± 12.9 W, and 275.0 ± 9.7 W ($P < 0.05$) for Hyperoxia (FI_{O_2} 1.0, SaO_2 100%, CaO_2 24 ml/dl), Iso-oxia (FI_{O_2} 0.28, SaO_2 98%, CaO_2 23 ml/dl), Normoxia (FI_{O_2} 0.21, SaO_2 91%, CaO_2 21 ml/dl), and Hypoxia (FI_{O_2} 0.15, SaO_2 77%, CaO_2 18 ml/dl), respectively. $n = 8$. From Amann et al. (6).

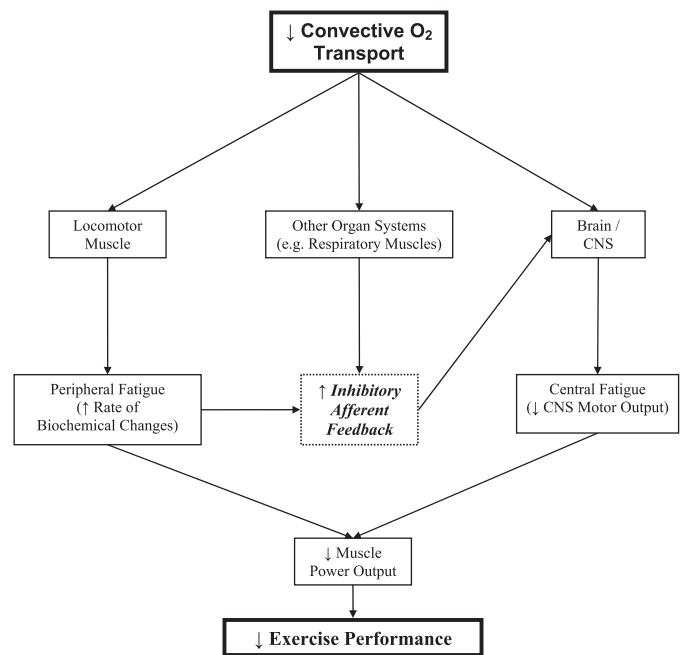


Fig. 5. Hypothetical scheme linking convective O_2 transport and exercise performance via its effects on fatigue. Alterations in convective O_2 transport occur throughout the organism and affect various organ systems. Consequently, various organs might affect exercise performance via inhibitory feedback mechanisms controlling central motor output. It has been proposed that peripheral muscle fatigue is carefully regulated via modulations of central nervous system (CNS) motor output to ensure muscle homeostasis during exercise from hyperoxia to moderate hypoxia (6). The relative importance of peripheral fatigue appears to diminish at more severe levels of hypoxia, and cerebral hypoxia might gain in relative influence regarding the termination of exercise (9). From Amann et al. (5).

tor muscle by means of blocking somatosensory afferent feedback.

Exceptions to this hypothesis have been shown to occur under conditions of critically low levels of brain oxygenation, i.e., exposure to extreme altitude or hyperventilation (see *Central Fatigue*). The O_2 -dependent rate of development of peripheral fatigue and associated inhibitory effects of somatosensory afferent feedback on central motor drive has been shown to be crucial up to an acutely challenged level of SaO_2 of $>70\%$ (6, 116), whereas below this level of SaO_2 , O_2 -sensitive sources of inhibition of central motor drive within the CNS appear to dominate the regulation of muscular performance (9, 24).

Summary

In many exercise models and environmental conditions, fatigue shows dependency on convective O_2 transport (Fig. 5). Reductions in O_2 supply exacerbate and increases attenuate the rate of fatigue development. Insufficient peripheral oxygenation elicits fatigue by facilitating the accumulation of metabolic byproducts, which interfere with excitation-contraction coupling within the myocyte. O_2 -dependent central fatigue is mediated via 1) the O_2 -sensitive balance of ATP demand and supply in brain areas involved in the generation of central motor command, and/or 2) inhibitory neural feedback to the CNS whose magnitude is proportional to the O_2 -dependent accumulation of metabolic byproducts in the working locomotor muscles.

REFERENCES

1. Adams RP, Welch HG. Oxygen uptake, acid-base status, and performance with varied inspired oxygen fractions. *J Appl Physiol* 49: 863–868, 1980.
2. Allen DG, Lannergren J, Westerblad H. Muscle cell function during prolonged activity: cellular mechanisms of fatigue. *Exp Physiol* 80: 497–527, 1995.
3. Allen DG and Westerblad H. The effects of caffeine on intracellular calcium, force and the rate of relaxation of mouse skeletal muscle. *J Physiol* 487: 331–342, 1995.
4. Amann M, Dempsey JA. Locomotor muscle fatigue modifies central motor drive in healthy humans and imposes a limitation to exercise performance. *J Physiol*, first published online October 25, 2007.
5. Amann M, Dempsey JA. Peripheral muscle fatigue from hyperoxia to moderate hypoxia – a carefully regulated variable? *Physiol News* 66: 28–29, 2007.
6. Amann M, Eldridge MW, Lovering AT, Stickland MK, Pegelow DF, Dempsey JA. Arterial oxygenation influences central motor output and exercise performance via effects on peripheral locomotor muscle fatigue. *J Physiol* 575: 937–952, 2006.
7. Amann M, Pegelow DF, Jacques AJ, Dempsey JA. Inspiratory muscle work in acute hypoxia influences locomotor muscle fatigue and exercise performance of healthy humans. *Am J Physiol Regul Integr Comp Physiol* 293: R2036–R2045, 2007.
8. Amann M, Romer LM, Pegelow DF, Jacques AJ, Hess CJ, Dempsey JA. Effects of arterial oxygen content on peripheral locomotor muscle fatigue. *J Appl Physiol* 101: 119–127, 2006.
9. Amann M, Romer LM, Subudhi AW, Pegelow DF, Dempsey JA. Severity of arterial hypoxaemia affects the relative contributions of peripheral muscle fatigue to exercise performance in healthy humans. *J Physiol* 581: 389–403, 2007.
10. Arbogast S, Vassilakopoulos T, Darques JL, Duvauchelle JB, Jammes Y. Influence of oxygen supply on activation of group IV muscle afferents after low-frequency muscle stimulation. *Muscle Nerve* 23: 1187–1193, 2000.
11. Arendt-Nielsen L, Mills KR. The relationship between mean power frequency of the EMG spectrum and muscle fibre conduction velocity. *Electroencephalogr Clin Neurophysiol* 60: 130–134, 1985.
12. Barclay JK. A delivery-independent blood flow effect on skeletal muscle fatigue. *J Appl Physiol* 61: 1084–1090, 1986.
13. Basmajian J, De Luca CJ. *Muscles Alive: Their Functions Revealed by Electromyography*. Baltimore, MD: Williams & Wilkins, 1985.
14. Bendahan D, Badier M, Jammes Y, Confort-Gouny S, Salvain AM, Guillot C, Cozzone PJ. Metabolic and myoelectrical effects of acute hypoxaemia during isometric contraction of forearm muscles in humans: a combined ³¹P-magnetic resonance spectroscopy-surface electromyogram (MRS-SEMG) study. *Clin Sci (Lond)* 94: 279–286, 1998.
15. Bhambhani Y, Malik R, Mookerjee S. Cerebral oxygenation declines at exercise intensities above the respiratory compensation threshold. *Respir Physiol Neurobiol* 156: 196–202, 2007.
16. Bigland-Ritchie B, Johansson R, Lippold OC, Woods JJ. Contractile speed and EMG changes during fatigue of sustained maximal voluntary contractions. *J Neurophysiol* 50: 313–324, 1983.
17. Bigland-Ritchie BR, Dawson NJ, Johansson RS, Lippold OC. Reflex origin for the slowing of motoneurone firing rates in fatigue of human voluntary contractions. *J Physiol* 379: 451–459, 1986.
18. Boushel R, Calbet JA, Radegran G, Sondergaard H, Wagner PD, Saltin B. Parasympathetic neural activity accounts for the lowering of exercise heart rate at high altitude. *Circulation* 104: 1785–1791, 2001.
19. Bowie W, Cumming GR. Sustained handgrip-reproducibility: effects of hypoxia. *Med Sci Sports* 3: 24–31, 1971.
20. Brugniaux JV, Hodges AN, Hanly PJ, Poulin MJ. Cerebrovascular responses to altitude. *Respir Physiol Neurobiol* 158: 221–223, 2007.
21. Bruton JD, Lannergren J, Westerblad H. Effects of CO₂-induced acidification on the fatigue resistance of single mouse muscle fibers at 28°C. *J Appl Physiol* 85: 478–483, 1998.
22. Burse RL, Cymerman A, Young AJ. Respiratory response and muscle function during isometric handgrip exercise at high altitude. *Aviat Space Environ Med* 58: 39–46, 1987.
23. Cairns SP, Knicker AJ, Thompson MW, Sjogaard G. Evaluation of models used to study neuromuscular fatigue. *Exerc Sport Sci Rev* 33: 9–16, 2005.
24. Calbet JA, Boushel R, Radegran G, Sondergaard H, Wagner PD, Saltin B. Determinants of maximal oxygen uptake in severe acute hypoxia. *Am J Physiol Regul Integr Comp Physiol* 284: R291–R303, 2003.
25. Calbet JA, Boushel R, Radegran G, Sondergaard H, Wagner PD, Saltin B. Why is $\dot{V}O_{2max}$ after altitude acclimatization still reduced despite normalization of arterial O₂ content? *Am J Physiol Regul Integr Comp Physiol* 284: R304–R316, 2003.
26. Calbet JA, De Paz JA, Garatachea N, Cabeza De Vaca S, Chavarren J. Anaerobic energy provision does not limit Wingate exercise performance in endurance-trained cyclists. *J Appl Physiol* 94: 668–676, 2003.
27. Calbet JA, Lundby C, Koskolou M, Boushel R. Importance of hemoglobin concentration to exercise: Acute manipulations. *Respir Physiol Neurobiol* 151: 132–140, 2006.
28. Calbet JA, Radegran G, Boushel R, Sondergaard H, Saltin B, Wagner PD. Effect of blood haemoglobin concentration on $\dot{V}(O_{2,max})$ and cardiovascular function in lowlanders acclimated to 5260 m. *J Physiol* 545: 715–728, 2002.
29. Calbet JAL. The rate of fatigue accumulation as a sensed variable. *J Physiol* 575: 688–689, 2006.
30. Calbet JAL, De Paz JA, Garatachea N, Cabeza de Vaca S, Chavarren J. Anaerobic energy provision does not limit Wingate exercise performance in endurance-trained cyclists. *J Appl Physiol* 94: 668–676, 2003.
31. Cheung SS. Hyperthermia and voluntary exhaustion: integrating models and future challenges. *Appl Physiol Nutr Metab* 32: 808–817, 2007.
32. Cymerman A, Reeves JT, Sutton JR, Rock PB, Groves BM, Malcomian MK, Young PM, Wagner PD, Houston CS. Operation Everest II: maximal oxygen uptake at extreme altitude. *J Appl Physiol* 66: 2446–2453, 1989.
33. Dahlstedt AJ, Katz A, Westerblad H. Role of myoplasmic phosphate in contractile function of skeletal muscle: studies on creatine kinase-deficient mice. *J Physiol* 533: 379–388, 2001.
34. Davis JM, Bailey SP. Possible mechanisms of central nervous system fatigue during exercise. *Med Sci Sports Exerc* 29: 45–57, 1997.
35. Degens H, Sanchez Horneros JM, Hopman MT. Acute hypoxia limits endurance but does not affect muscle contractile properties. *Muscle Nerve* 33: 532–537, 2006.
36. Delp MD, Armstrong RB, Godfrey DA, Laughlin MH, Ross CD, Wilkerson MK. Exercise increases blood flow to locomotor, vestibular, cardiorespiratory and visual regions of the brain in miniature swine. *J Physiol* 533: 849–859, 2001.
37. Dempsey JA, Romer L, Rodman J, Miller J, Smith C. Consequences of exercise-induced respiratory muscle work. *Respir Physiol Neurobiol* 151: 242–250, 2006.
38. Dempsey JA, Wagner PD. Exercise-induced arterial hypoxemia. *J Appl Physiol* 87: 1997–2006, 1999.
39. Duhamel TA, Green HJ, Sandiford SD, Perco JG, Ouyang J. Effects of progressive exercise and hypoxia on human muscle sarcoplasmic reticulum function. *J Appl Physiol* 97: 188–196, 2004.
40. Duong TQ. Cerebral blood flow and BOLD fMRI responses to hypoxia in awake and anesthetized rats. *Brain Res* 1135: 186–194, 2007.
41. Dutka TL, Cole L, Lamb GD. Calcium phosphate precipitation in the sarcoplasmic reticulum reduces action potential-mediated Ca²⁺ release in mammalian skeletal muscle. *Am J Physiol Cell Physiol* 289: C1502–C1512, 2005.
42. Edwards R. Human muscle function and fatigue. In: *Human Muscle Fatigue Physiological Mechanisms*, edited by Porter R and Whelan J. London: Pitman, 1981, p. 1–8.
43. Eiken O, Tesch PA. Effects of hyperoxia and hypoxia on dynamic and sustained static performance of the human quadriceps muscle. *Acta Physiol Scand* 122: 629–633, 1984.
44. Enoka RM, Stuart DG. Neurobiology of muscle fatigue. *J Appl Physiol* 72: 1631–1648, 1992.
45. Fabiato A, Fabiato F. Effects of pH on the myofilaments and the sarcoplasmic reticulum of skinned cells from cardiac and skeletal muscles. *J Physiol* 276: 233–255, 1978.
46. Fitts RH. Cellular mechanisms of muscle fatigue. *Physiol Rev* 74: 49–94, 1994.
47. Fitts RH, Holloszy JO. Lactate and contractile force in frog muscle during development of fatigue and recovery. *Am J Physiol* 231: 430–433, 1976.

48. Fryer MW, Owen VJ, Lamb GD, Stephenson DG. Effects of creatine phosphate and P(i) on Ca²⁺ movements and tension development in rat skinned skeletal muscle fibres. *J Physiol* 482: 123–140, 1995.
49. Fulco CS, Cymerman A, Muza SR, Rock PB, Pandolf KB, Lewis SF. Adductor pollicis muscle fatigue during acute and chronic altitude exposure and return to sea level. *J Appl Physiol* 77: 179–183, 1994.
50. Fulco CS, Lewis SF, Frykman PN, Boushel R, Smith S, Harman EA, Cymerman A, Pandolf KB. Muscle fatigue and exhaustion during dynamic leg exercise in normoxia and hypobaric hypoxia. *J Appl Physiol* 81: 1891–1900, 1996.
51. Fulco CS, Rock PB, Cymerman A. Maximal and submaximal exercise performance at altitude. *Aviat Space Environ Med* 69: 793–801, 1998.
52. Gandevia SC. Spinal and supraspinal factors in human muscle fatigue. *Physiol Rev* 81: 1725–1789, 2001.
53. Gandevia SC, Enoka RM, McComas AJ, Stuart DG, Thomas CK. *Fatigue: Neural and Muscular Mechanisms*. New York: Plenum Press, 1995.
54. Garner SH, Sutton JR, Burse RL, McComas AJ, Cymerman A, Houston CS. Operation Everest II: neuromuscular performance under conditions of extreme simulated altitude. *J Appl Physiol* 68: 1167–1172, 1990.
55. Godt RE, Nosek TM. Changes of intracellular milieu with fatigue or hypoxia depress contraction of skinned rabbit skeletal and cardiac muscle. *J Physiol* 412: 155–180, 1989.
56. Gonzalez-Alonso J. Hyperthermia impairs brain, heart and muscle function in exercising humans. *Sports Med* 37: 371–373, 2007.
57. Gonzalez-Alonso J, Calbet JA. Reductions in systemic and skeletal muscle blood flow and oxygen delivery limit maximal aerobic capacity in humans. *Circulation* 107: 824–830, 2003.
58. Gonzalez-Alonso J, Calbet JA, Nielsen B. Muscle blood flow is reduced with dehydration during prolonged exercise in humans. *J Physiol* 513: 895–905, 1998.
59. Gonzalez-Alonso J, Dalsgaard MK, Osada T, Volianitis S, Dawson EA, Yoshiga CC, Secher NH. Brain and central haemodynamics and oxygenation during maximal exercise in humans. *J Physiol* 557: 331–342, 2004.
60. Graham T, Wilson BA, Sample M, Van Dijk J, Bonen A. The effects of hypercapnia on metabolic responses to progressive exhaustive work. *Med Sci Sports Exerc* 12: 278–284, 1980.
61. Green HJ. Mechanisms of muscle fatigue in intense exercise. *J Sports Sci* 15: 247–256, 1997.
62. Harms CA, Babcock MA, McClaran SR, Pegelow DF, Nickle GA, Nelson WB, Dempsey JA. Respiratory muscle work compromises leg blood flow during maximal exercise. *J Appl Physiol* 82: 1573–1583, 1997.
63. Harms CS, McClaran SR, Nickle GA, Pegelow DF, Nelson WB, Dempsey JA. Effect of exercise-induced arterial O₂ desaturation on VO₂max in women. *Med Sci Sports Exerc* 32: 1101–1108, 2000.
64. Harms CS, Wetter TJ, St Croix CM, Pegelow DF, Dempsey JA. Effects of respiratory muscle work on exercise performance. *J Appl Physiol* 89: 131–138, 2000.
65. Haseler LJ, Hogan MC, Richardson RS. Skeletal muscle phosphocreatine recovery in exercise-trained humans is dependent on O₂ availability. *J Appl Physiol* 86: 2013–2018, 1999.
66. Haseler LJ, Richardson RS, Videen JS, Hogan MC. Phosphocreatine hydrolysis during submaximal exercise: the effect of F_{IO₂}. *J Appl Physiol* 85: 1457–1463, 1998.
67. Hellstrom G, Fischer-Colbrie W, Wahlgren NG, Jogestrand T. Carotid artery blood flow and middle cerebral artery blood flow velocity during physical exercise. *J Appl Physiol* 81: 413–418, 1996.
68. Hepple RT. The role of O₂ supply in muscle fatigue. *Can J Appl Physiol* 27: 56–69, 2002.
69. Hogan MC, Kohin S, Stary CM, Hepple RT. Rapid force recovery in contracting skeletal muscle after brief ischemia is dependent on O₂ availability. *J Appl Physiol* 87: 2225–2229, 1999.
70. Hogan MC, Nioka S, Brechue WF, Chance B. A ³¹P-NMR study of tissue respiration in working dog muscle during reduced O₂ delivery conditions. *J Appl Physiol* 73: 1662–1670, 1992.
71. Hogan MC, Richardson RS, Haseler LJ. Human muscle performance and PCR hydrolysis with varied inspired oxygen fractions: a ³¹P-MRS study. *J Appl Physiol* 86: 1367–1373, 1999.
72. Hogan MC, Richardson RS, Kurdak SS. Initial fall in skeletal muscle force development during ischemia is related to oxygen availability. *J Appl Physiol* 77: 2380–2384, 1994.
73. Hogan MC, Roca J, Wagner PD, West JB. Limitation of maximal O₂ uptake and performance by acute hypoxia in dog muscle in situ. *J Appl Physiol* 65: 815–821, 1988.
74. Hogan MC, Welch HG. Effect of altered arterial O₂ tensions on muscle metabolism in dog skeletal muscle during fatiguing work. *Am J Physiol Cell Physiol* 251: C216–C222, 1986.
75. Hogan MC, Welch HG. Effect of varied lactate levels on bicycle ergometer performance. *J Appl Physiol* 57: 507–513, 1984.
76. Hornbein TF, Townes BD, Schoene RB, Sutton JR, Houston CS. The cost to the central nervous system of climbing to extremely high altitude. *N Engl J Med* 321: 1714–1719, 1989.
77. Ide K, Boushel R, Sorensen HM, Fernandes A, Cai Y, Pott F, Secher NH. Middle cerebral artery blood velocity during exercise with beta-1 adrenergic and unilateral stellate ganglion blockade in humans. *Acta Physiol Scand* 170: 33–38, 2000.
78. Ide K, Eliasziw M, Poulin MJ. Relationship between middle cerebral artery blood velocity and end-tidal PCO₂ in the hypocapnic-hypercapnic range in humans. *J Appl Physiol* 95: 129–137, 2003.
79. Ide K, Gulløv AL, Pott F, Van Lieshout JJ, Koefoed BG, Petersen P, Secher NH. Middle cerebral artery blood velocity during exercise in patients with atrial fibrillation. *Clin Physiol* 19: 284–289, 1999.
80. Kanstrup IL, Ekblom B. Blood volume and hemoglobin concentration as determinants of maximal aerobic power. *Med Sci Sports Exerc* 16: 256–262, 1984.
81. Katayama K, Amann M, Pegelow DF, Jacques AJ, Dempsey JA. Effect of arterial oxygenation on quadriceps fatigability during isolated muscle exercise. *Am J Physiol Regul Integr Comp Physiol* 292: R1279–R1286, 2007.
82. Kayser B, Narici M, Binzoni T, Grassi B, Cerretelli P. Fatigue and exhaustion in chronic hypobaric hypoxia: influence of exercising muscle mass. *J Appl Physiol* 76: 634–640, 1994.
83. Kiening KL, Unterberg AW, Bardt TF, Schneider GH, Lanksch WR. Monitoring of cerebral oxygenation in patients with severe head injuries: brain tissue PO₂ versus jugular vein oxygen saturation. *J Neurosurg* 85: 751–757, 1996.
84. Kjaer M, Hanel B, Worm L, Perko G, Lewis SF, Sahlin K, Galbo H, Secher NH. Cardiovascular and neuroendocrine responses to exercise in hypoxia during impaired neural feedback from muscle. *Am J Physiol Regul Integr Comp Physiol* 277: R76–R85, 1999.
85. Knight DR, Schaffartzik W, Poole DC, Hogan MC, Bebout DE, Wagner PD. Effects of hyperoxia on maximal leg O₂ supply and utilization in men. *J Appl Physiol* 75: 2586–2594, 1993.
86. Koskolou MD, Calbet JA, Radegran G, Roach RC. Hypoxia and the cardiovascular response to dynamic knee-extensor exercise. *Am J Physiol Heart Circ Physiol* 272: H2655–H2663, 1997.
87. Koskolou MD, Roach RC, Calbet JA, Radegran G, Saltin B. Cardiovascular responses to dynamic exercise with acute anemia in humans. *Am J Physiol Heart Circ Physiol* 273: H1787–H1793, 1997.
88. Lamb GD, Stephenson DG, Bangsbo J, Juel C. Lactic acid accumulation is an advantage/disadvantage during muscle activity. *J Appl Physiol* 100: 1410–1412, 2006.
89. Lannergren J, Westerblad H. Force decline due to fatigue and intracellular acidification in isolated fibres from mouse skeletal muscle. *J Physiol* 434: 307–322, 1991.
90. Lassen NA. Control of cerebral circulation in health and disease. *Circ Res* 34: 749–760, 1974.
91. Lee JA, Westerblad H, Allen DG. Changes in tetanic and resting [Ca²⁺]_i during fatigue and recovery of single muscle fibres from *Xenopus laevis*. *J Physiol* 433: 307–326, 1991.
92. Lundby C, Damsgaard R. Exercise performance in hypoxia after novel erythropoiesis stimulating protein treatment. *Scand J Med Sci Sports* 16: 35–40, 2006.
93. Maas AI, Fleckenstein W, de Jong DA, van Santbrink H. Monitoring cerebral oxygenation: experimental studies and preliminary clinical results of continuous monitoring of cerebrospinal fluid and brain tissue oxygen tension. *Acta Neurochir Suppl (Wien)* 59: 50–57, 1993.
94. Maher JT, Jones LG, Hartley LH. Effects of high-altitude exposure on submaximal endurance capacity of men. *J Appl Physiol* 37: 895–898, 1974.
95. Meeusen R, Watson P, Hasegawa H, Roelands B, Piacentini MF. Central fatigue: the serotonin hypothesis and beyond. *Sports Med* 36: 881–909, 2006.

96. Merletti R, Knaflitz M, De Luca CJ. Myoelectric manifestations of fatigue in voluntary and electrically elicited contractions. *J Appl Physiol* 69: 1810–1820, 1990.
97. Merton PA. Voluntary strength and fatigue. *J Physiol* 123: 553–564, 1954.
98. Miyamoto O, Auer RN. Hypoxia, hyperoxia, ischemia, and brain necrosis. *Neurology* 54: 362–371, 2000.
99. Moller K, Paulson OB, Hornbein TF, Colier WN, Paulson AS, Roach RC, Holm S, Knudsen GM. Unchanged cerebral blood flow and oxidative metabolism after acclimatization to high altitude. *J Cereb Blood Flow Metab* 22: 118–126, 2002.
100. Moraine JJ, Lamotte M, Berre J, Niset G, Leduc A, Naeije R. Relationship of middle cerebral artery blood flow velocity to intensity during dynamic exercise in normal subjects. *Eur J Appl Physiol* 67: 35–38, 1993.
101. Nemoto EM, Yonas H, Kuwabara H, Pindzola RR, Sashin D, Meltzer C, Price JC, Chang Y. Detection of stage II compromised cerebrovascular reserve by xenon-CT cerebral blood flow with acetazolamide and oxygen extraction fraction by positron emission tomography. In: *Brain Imaging Using PET*, edited by Senda M, Kimura Y and Herscovitch P. San Diego, CA: Academic, 2002, p. 259–267.
102. Nielsen HB, Boushel R, Madsen P, Secher NH. Cerebral desaturation during exercise reversed by O₂ supplementation. *Am J Physiol Heart Circ Physiol* 277: H1045–H1052, 1999.
103. Nybo L, Moller K, Volianitis S, Nielsen B, Secher NH. Effects of hyperthermia on cerebral blood flow and metabolism during prolonged exercise in humans. *J Appl Physiol* 93: 58–64, 2002.
104. Nybo L, Nielsen B. Middle cerebral artery blood velocity is reduced with hyperthermia during prolonged exercise in humans. *J Physiol* 534: 279–286, 2001.
105. Nybo L, Rasmussen P. Inadequate cerebral oxygen delivery and central fatigue during strenuous exercise. *Exerc Sport Sci Rev* 35: 110–118, 2007.
106. Nybo L, Secher NH. Cerebral perturbations provoked by prolonged exercise. *Prog Neurobiol* 72: 223–261, 2004.
107. Orizio C, Esposito F, Vecesteinas A. Effect of acclimatization to high altitude (5,050 m) on motor unit activation pattern and muscle performance. *J Appl Physiol* 77: 2840–2844, 1994.
108. Pate E, Bhimani M, Franks-Skiba K, Cooke R. Reduced effect of pH on skinned rabbit psoas muscle mechanics at high temperatures: implications for fatigue. *J Physiol* 486: 689–694, 1995.
109. Pedersen TH, Nielsen OB, Lamb GD, Stephenson DG. Intracellular acidosis enhances the excitability of working muscle. *Science* 305: 1144–1147, 2004.
110. Pott F, Jensen K, Hansen H, Christensen NJ, Lassen NA, Secher NH. Middle cerebral artery blood velocity and plasma catecholamines during exercise. *Acta Physiol Scand* 158: 349–356, 1996.
111. Poulin MJ, Fatemian M, Tansley JG, O'Connor DF, Robbins PA. Changes in cerebral blood flow during and after 48 h of both isocapnic and poikilocapnic hypoxia in humans. *Exp Physiol* 87: 633–642, 2002.
112. Querido JS, Sheel AW. Regulation of cerebral blood flow during exercise. *Sports Med* 37: 765–782, 2007.
113. Raichle ME, Hornbein TF. The high-altitude brain. In: *High Altitude: an Exploration of Human Adaptation*, edited by Hornbein TF and Schoene RB. New York: Marcel Dekker, 2001, p. 377–423.
114. Rasmussen P, Dawson EA, Nybo L, van Lieshout JJ, Secher NH, Gjedde A. Capillary-oxygenation-level-dependent near-infrared spectrometry in frontal lobe of humans. *J Cereb Blood Flow Metab* 27: 1082–1093, 2007.
115. Rasmussen P, Stie H, Nielsen B, Nybo L. Enhanced cerebral CO₂ reactivity during strenuous exercise in man. *Eur J Appl Physiol* 96: 299–304, 2006.
116. Romer LM, Haverkamp HC, Amann M, Lovering AT, Pegelow DF, Dempsey JA. Effect of acute severe hypoxia on peripheral fatigue and endurance capacity in healthy humans. *Am J Physiol Regul Integr Comp Physiol* 292: R598–R606, 2007.
117. Romer LM, Haverkamp HC, Lovering AT, Pegelow DF, Dempsey JA. Effect of exercise-induced arterial hypoxemia on quadriceps muscle fatigue in healthy humans. *Am J Physiol Regul Integr Comp Physiol* 290: R365–R375, 2006.
118. Romer LM, Lovering AT, Haverkamp HC, Pegelow DF, Dempsey JA. Effect of inspiratory muscle work on peripheral fatigue of locomotor muscles in healthy humans. *J Physiol* 571: 425–439, 2006.
119. Sandiford SD, Green HJ, Duhamel TA, Perco JG, Schertzer JD, Ouyang J. Inactivation of human muscle Na⁺-K⁺-ATPase in vitro during prolonged exercise is increased with hypoxia. *J Appl Physiol* 96: 1767–1775, 2004.
120. Sandiford SD, Green HJ, Duhamel TA, Schertzer JD, Perco JD, Ouyang J. Muscle Na-K-pump and fatigue responses to progressive exercise in normoxia and hypoxia. *Am J Physiol Regul Integr Comp Physiol* 289: R441–R449, 2005.
121. Savard GK, Areskog NH, Saltin B. Maximal muscle activation is not limited by pulmonary ventilation in chronic hypoxia. *Acta Physiol Scand* 157: 187–190, 1996.
122. Schuler B, Thomsen JJ, Gassmann M, Lundby C. Timing the arrival at 2340 m altitude for aerobic performance. *Scand J Med Sci Sports* 17: 588–594, 2007.
123. Stary CM, Hogan MC. Impairment of Ca²⁺ release in single *Xenopus* muscle fibers fatigued at varied extracellular PO₂. *J Appl Physiol* 88: 1743–1748, 2000.
124. Stephens JA, Taylor A. Fatigue of maintained voluntary muscle contraction in man. *J Physiol* 220: 1–18, 1972.
125. Stienen GJ, Roosemalen MC, Wilson MG, Elzinga G. Depression of force by phosphate in skinned skeletal muscle fibers of the frog. *Am J Physiol Cell Physiol* 259: C349–C357, 1990.
126. Subudhi AW, Dimmen AC, Roach RC. Effects of acute hypoxia on cerebral and muscle oxygenation during incremental exercise. *J Appl Physiol* 103: 177–183, 2007.
127. Swallow EB, Gosker HR, Ward KA, Moore AJ, Dayer MJ, Hopkinson NS, Schols AMWJ, Moxham J, Polkey MI. A novel technique for non-volitional assessment of quadriceps muscle endurance in man. *J Appl Physiol*. doi:10.1152/jappphysiol.00025.2007.
128. Taylor AD, Bronks R, Smith P, Humphries B. Myoelectric evidence of peripheral muscle fatigue during exercise in severe hypoxia: some references to m. vastus lateralis myosin heavy chain composition. *Eur J Appl Physiol* 75: 151–159, 1997.
129. Thomsen JJ, Rentsch RL, Robach P, Calbet JA, Boushel R, Rasmussen P, Juel C, Lundby C. Prolonged administration of recombinant human erythropoietin increases submaximal performance more than maximal aerobic capacity. *Eur J Appl Physiol*, 2007.
130. Todd G, Butler JE, Taylor JL, Gandevia SC. Hyperthermia: a failure of the motor cortex and the muscle. *J Physiol*, 2004.
131. van den Brink WA, van Santbrink H, Steyerberg EW, Avezaat CJ, Suazo JA, Hogesteeger C, Jansen WJ, Kloos LM, Vermeulen J, Maas AI. Brain oxygen tension in severe head injury. *Neurosurgery* 46: 868–876; discussion 876–868, 2000.
132. van Santbrink H, Maas AI, Avezaat CJ. Continuous monitoring of partial pressure of brain tissue oxygen in patients with severe head injury. *Neurosurgery* 38: 21–31, 1996.
133. Vollestad NK. Measurement of human muscle fatigue. *J Neurosci Methods* 74: 219–227, 1997.
134. Westerblad H, Allen DG, Lannergren J. Muscle fatigue: lactic acid or inorganic phosphate the major cause? *News Physiol Sci* 17: 17–21, 2002.
135. Westerblad H, Lannergren J, Allen DG. Slowed relaxation in fatigued skeletal muscle fibers of *Xenopus* and Mouse. Contribution of [Ca²⁺]_i and cross-bridges. *J Gen Physiol* 109: 385–399, 1997.
136. Young A, Wright J, Knapik J, Cymerman A. Skeletal muscle strength during exposure to hypobaric hypoxia. *Med Sci Sports Exerc* 12: 330–335, 1980.