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Biliary cirrhosis caused by *Campula* spp. in a dolphin and four porpoises

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ABSTRACT: Biliary cirrhosis produced by *Campula* spp. is described in 1 striped dolphin *Stenella coeruleoalba* and 4 harbour porpoises *Phocoena phocoena*. The hepatic lesions consisted of severe proliferation of fibrous connective tissue with loss of the lobular pattern, nodular regeneration of the hepatic tissue, bile duct hyperplasia and severe inflammatory infiltrate composed of eosinophils, macrophages, lymphocytes and plasma cells. These lesions were associated with severe infestation by *Campula* spp. Although inflammatory and degenerative hepatic lesions are frequently found in stranded dolphins, biliary cirrhosis has not been previously reported in cetaceans. Massive infestation by these parasites should be included as a cause of hepatic failure resulting in stranding of marine mammals.

KEY WORDS: Cetacean \cdot Stenella coeruleoalba \cdot Phocoena phocoena \cdot Hepatic lesions \cdot Biliary cirrhosis \cdot Trematode

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INTRODUCTION

Interest in parasitic diseases of marine mammals has increased greatly during the last few years because they have accounted for an important portion of the pathological changes found in these animals (Dailey & Ridgway 1976). One of the most frequent parasitic infestations observed in the digestive system of cetaceans is produced by *Campula* spp., trematodes that primarily inhabit the bile and pancreatic ducts of cetaceans (Zam et al. 1971). A high prevalence of these worms in different species of cetaceans has been reported (Zam et al. 1971, Sweeney & Ridgway 1975, Dailey & Stroud 1978, Jaber et al. 2004), producing extensive irritation of the ducts, fibrosis in the portal areas, biliary hyperplasia (Dailey & Stroud 1978) and inflammatory infiltration composed mainly of lymphocytes, plasma cells and eosinophils (Migaki et al. 1979).

In humans, chronic obstruction of the intrahepatic biliary duct may result in primary biliary cirrhosis (PBC; Speranzini et al. 1988, Crawford 1999), characterised by granulomatous inflammation and destruction of medium-sized intrahepatic biliary ducts, portal inflammation and subsequent fibrosis, resulting in cirrhosis and hepatic insufficiency (Gershwin et al. 2000). The hepatic dysfunction produced by this disease can result in a metabolic disorder of the central nervous system that is termed hepatic encephalopathy (Crawford 1999). In terrestrial mammals, biliary cirrhosis has been experimentally induced by surgery in dogs (Koblik et al. 1995) or by antifungal drugs in rats (Somchit et al. 2004), and has been associated in cattle with parasites (Vítovec 1974) or a mass in the biliary tract (Shimada et al. 1998). However, to date, this disease has not been described in marine mammals. The aim of this paper was to study the pathological and immunohistochemical findings associated with severe infestation by *Campula* spp. in 2 species of cetaceans stranded in different locations.

MATERIALS AND METHODS

A total of 5 animals were included in this study: 4 subadult harbour porpoises Phocoena phocoena (3 females, 1 male) stranded on the eastern coast of the USA and 1 mature male striped dolphin Stenella coeruleoalba stranded on the shore of Fuerteventura, Canary Islands, Spain. The latter showed hypothermia, disorientation and weakness. A full post mortem examination was carried out for all 5 animals. After opening the abdominal cavity, an increase in the abdominal fluid and moderate yellowish colouration of the serosal membranes were observed. Grossly, the liver was normal in size, but moderately hard in consistency. The liver of the striped dolphin showed the presence of numerous hard and whitish nodules ranging from 2 to 3 cm in diameter, affecting approximately half of the organ, and associated with marked dilatation of the hepatic ducts. In the other animals, these nodules affected one third of the organ. The common hepatic duct contained numerous adult trematodes, causing a partial obstruction of this duct. Samples from the central nervous system, lung, heart, liver, spleen, forestomach, fundic stomach, pyloric stomach, pancreas, adrenal gland, intestine, and prescapular, mediastinal and mesenteric lymph nodes were collected and fixed in 10% neutral buffered formalin. Formalin-fixed tissues were routinely processed, embedded in paraffin wax, sectioned at 5 μ m, and stained with haematoxylin and eosin (H&E) for histological examination.

For the ultrastructural study, liver tissue samples from the striped dolphin initially fixed in formalin were post-fixed in osmium tetroxide (pH 7.4) and embedded in epoxy resin. Semi-thin sections were cut and stained with toluidine blue for light microscopical examination. Ultrathin sections were cut from selected blocks, stained with lead citrate and uranyl acetate and examined using a Philips CM-10 transmission electron microscope. To study parasites found in the liver, samples were processed, goldsputtered, and examined using a Jeol JSM 6300 scanning electron microscope.

Selected sections were stained by an immunoperoxidase test for Morbillivirus antigen (Kennedy et al. 1991). The avidin-biotin-peroxidase method described elsewhere (Pérez et al. 2001, Jaber et al. 2013) was used to further characterise the inflammatory infiltrate. Details of the optimal antigen retrieval methods and dilutions of the specific primary antibodies are given in Table 1. Primary antibodies against CD3, lysozyme, MHC class II, S-100 protein and immunoglobin (IgG) have been shown to crossreact with cetacean tissues (Jaber et al. 2003). Crossreactivity between the primary antibodies anti-rat Foxp3 and anti-human iNOS in cetacean tissues was evaluated in the present study. Tissue sections in which the specific primary antibodies were substituted with PBS, rabbit or mouse non-immune sera were used as negative controls. Dolphin, human, bovine and mouse lymph node tissue sections were used as positive controls. Immunolabelled tissue sections were evaluated with a photomicroscope, and the density of immunolabelled cells in affected areas was scored by 2 pathologists in 10 fields of 0.2 mm² as follows: -, negative; ±/-, 0 to 5; +, 6 to 15; ++, 16 to 30; +++, > 30 cells per field.

RESULTS AND DISCUSSION

Histological examination revealed that the biliary ducts contained an elevated number of adult parasites and eggs. Morphologic features were consistent with *Campula* spp. (Zam et al. 1971, Dailey & Stroud 1978), characterised by the single pair of anterior diverticula of the intestinal caeca and the lobed testes. Scanning electron microscopy revealed a tegument that was covered with large and scale-like

Table 1. Primary antibodies used in this study. pAb: polyclonal antibody; mAb: monoclonal antibody. Treatments were as follows; Pronase: 0.01% pronase (Sigma Chemical) for 10 min at room temperature; Microwave: 0.01% citric acid in microwave at 100°C for 7 min; Autoclave: 0.01% citric acid in autoclave at 121°C for 10 min

Antibody	Origin	Treatment	Dilution	Manufacturer	
CD3 (pAb)	Human	Pronase	1:200	Dako	
Lysozyme (pAb)	Human	Pronase	1:100	Dako	
S-100 (pAb)	Bovine	None	1:200	Dako	
iNOS (pAb)	Mouse	Pronase	1:100	Neomarkers	
IgG (pAb)	Human	Pronase	1:1000	Dako	
MHC class II (mAb)	Human	Microwave	1:50	Dako	
Foxp3 (mAb)	Mouse	Autoclave	1:100	eBioscience	



Fig. 1. *Campula* sp. Detail of the suckers of the parasite and its tegument with large and scale-like spines (scanning electron microscopy) ×45

spines and the presence of 2 suckers, one observed ventrally (also called the acetabulum) and the other surrounding the mouth or anterior sucker (Fig. 1).

The presence of parasites in the bile ducts induced epithelial cell erosion, bile duct hyperplasia, portal fibrosis and severe inflammatory infiltrate (Fig. 2). Rupturing of the affected biliary ducts resulted in the presence of parasites and their eggs in the stroma, which was surrounded by eosinophils, lysozymepositive macrophages and multinucleate giant cells,

and peripherally by numerous CD3⁺ T lymphocytes and IgG⁺ plasma cells leading to granulomata formation (Table 2). Severe bridging fibrosis that extended from one portal tract to another was observed (Fig. 2). Within the fibrotic regions, there was some nodular regeneration of the hepatic tissue resulting in disorganisation of the hepatic architecture and neovascularisation, bile duct proliferation and infiltration of mononuclear cells composed of lysozyme-positive macrophages, CD3⁺ lymphocytes (Fig. 3) and IgG⁺ plasma cells within the fibrotic tissue. Around these portal areas, there was an abundant inflammatory infiltrate composed of IgG⁺ plasma cells and CD3⁺ lymphocytes, which were predominantly organised in lymphoid follicles, with prominent germinal centres and interfollicular diffuse lymphoid tissue.

Occasional cells with large cytoplasm identified as macrophages showed cytoplasmic immunostaining for iNOS. The low numbers of macrophages expressing iNOS in the inflammatory infiltrate of the present study agree with the low iNOS expression in chronic cholangitis caused by Fasciola hepatica in goats and sheep (authors' unpubl. results). The Foxp3 antibody showed nuclear and cytoplasmic immunoreactivity in some lymphocytes located in the paracortical areas of dolphin lymph nodes and also in a few lymphocytes in the hepatic inflammatory infiltrates, mainly in diffuse lymphoid infiltrate closely related to bile ducts where 5 to 8% of lymphocytes were foxp3⁺ (Fig. 4). In parasitic diseases of domestic animals, such as Psoroptes ovis in sheep, Foxp3⁺ T lymphocytes are recruited to the skin lesions (McNeilly et al. 2010). In contrast, no Foxp3⁺ cells were found in hepatic lesions caused by Schistosoma mansoni in mice (Dewals et al. 2010), and reduction of this cell type in chronic colonic granulomas compared to acute lesions was observed in mice infected with S. mansoni (Turner et al. 2011). In chronic hepatic lesions caused by F. hepatica in goats, a marked reduction of Foxp3+ cells was found (authors' unpubl. data). This is in agreement with the occasional Foxp3⁺ cells found in the hepatic inflammatory infiltrates of the cetaceans in the present study and also with recent studies reporting that helminths such as S. mansoni and Heligmosomoides polygyrus led to a reduced



Fig. 2. *Campula* sp. infecting *Phocoena phocoena*. Presence of the parasite in a liver bile duct (B) showing severe portal fibrosis (F) and inflammatory infiltration (I) in portal spaces and hepatic parenchyma (L), resulting in loss of the he patic lobular pattern (H&E). Scale bar = 100 µm

Table 2. Phocoena phocoena and Stenella coeruleoalba. Distribution of immunolabelled cells (-: 0; +/-: 0-5; +: 5-15; ++: 15-30; +++: >30 cells per 0.2 mm²) with the stated antibodies in the lymphonodular inflammatory infiltrates of the liver. LYSOZ: lysozyme

Case no.	CD3	LYSOZ	S-100	iNOS	IgG	MHC class II	Foxp3		
Harbour porpoise									
IFAW11-054Pp	+++	++	++	_	+++	+	+		
IFAW11-088Pp	++ F	+/-	+	_	+	+	_		
IFAW11-039Pp	+++	+	+	+/-	++/+++	+	+		
IFAW11-009Pp	+++	+/-	+	+/-	+	+/-	_		
Striped dolphin 61/00	+++	_	-	+/-	++	+/-	+/-		



Fig. 3. Phocoena phocoena. Bile duct proliferation (B) and infiltration of mononuclear cells composed mainly of CD3⁺ lymphocytes (arrows) within the fibrotic tissue. L: liver. (avidin-biotin-peroxidase method, Mayer's haema toxylin counterstain). Scale bar = 100 μm

expansion and maintenance of Foxp3 Treg cells (Redpath et al. 2013).

The lymphoproliferative nodules were associated with the parasitic cholangitis and were similar to those found in the cortex of control lymph nodes, including the presence of S100⁺ and MHC class II⁺ dendritic-like cells in lymphoid follicles and interfollicular areas. Similar cellular distribution has been reported in goats (Pérez et al. 1999) parasitised by *Fasciola hepatica* and in pigs parasitised by *Ascaris suum* (Pérez et al. 2001). The organisation of these lymphonodular inflammatory infiltrates may enhance antigen presentation and the local humoral and cellular immune responses against parasite or egg antigens. In addition, the high number of IgG⁺ plasma cells indicates a strong local humoral response against this parasite.

In the striped dolphin, some hepatocytes contained cytoplasmic hyaline inclusions, which were round to oval in shape, moderately eosinophilic and on occasion caused peripheral displacement of the nucleus. In these inclusions, a central or eccentric densely staining core was frequently apparent. During stranding, thoracic and abdominal vasculature compression compromises blood flow, leading to acute to subacute liver congestion and subsequent hepatocellular hypoxia. These hepatocytes then develop hyaline inclusions, which are similar to those seen in humans with congestive heart failure (Ponsold 1961). This finding suggests that this morphological change would be reversible, as in humans (Jaber et al. 2004)

In the central nervous system of the striped dolphin, numerous cell bodies of the neurons were shrunken and the cytoplasm stained lightly eosinophilic, whereas the nucleus of the neurons was reduced in size, and the nucleolus was not detectable. Accumulations of glial cells were found surrounding neuronal cell bodies and vessels. Diffuse vacuolation, particularly the white matter, was observed. This vacuolation showed a well defined pattern following a number of myelinated tracts in central nervous system white matter, in isolated fibres crossing grey matter in the

brain stem, and along grey and white matter borders in the cerebrum and spinal cord. These histological findings were suggestive of hepatic encephalopathy, which is a complication that develops in a small percentage of human patients with chronic liver disease (Durán-Ferreras et al. 2011). However, in the above mentioned case, it is difficult to distinguish whether the cause of the vacuolation was provoked by this process or related to agonal hypoxia.

The test for *Morbillivirus* antigens was negative in all of the selected sections studied. Histopathological examination of the remaining organs was not remarkable.

In humans, PBC is an organ-specific autoimmune disease that predominantly affects women and is characterised by chronic progressive destruction of



Fig. 4. *Stenella coeruleoalba.* Foxp3 antibody showing nuclear and cytoplasmic immunoreactivity in a few lymphocytes (arrows) located in the hepatic inflammatory infiltrates in the liver (avidin-biotin-peroxidase method, Mayer's haematoxylin counterstain). Scale bar = 50 μm

small intrahepatic bile ducts with portal inflammation and ultimately fibrosis (Gershwin et al. 2000). Biliary cirrhosis observed in the dolphins in the present study was associated with parasites and their eggs in the stroma of portal spaces. A parasitic origin of PBC has been reported in cows parasitised by Fasciola hepatica (Vítovec 1974, Pérez et al. 1999). This trematode has frequently been found in the liver and pancreas of cetaceans (Migaki et al. 1979, Jaber et al. 2004). Light infestation may be asymptomatic, but heavy infestations may cause changes similar to those observed in the present study, in which major severity of the lesions was observed when parasite eggs were found in the stroma of portal spaces, causing severe granulomatous lesions with destruction of the biliary ducts, similar to that described in humans (Crawford 1999, Gershwin et al. 2000). Thus, the histopathological features closely resemble those reported in humans (Crawford 1999), but the lymphocytic infiltrate in granulomatous lesions associated with parasite eggs was more prominent in this dolphin. Biliary cirrhosis caused by Fasciola spp. in ruminants (Vítovec 1974) and experimentally induced in dogs (Koblik et al. 1995) also showed similar histopathological features to those observed in our case.

To date, there have been no reports of primary biliary cirrhosis associated with *Campula* spp. in

cetaceans. While these lesions have been well documented in ruminants with fascioliasis and in humans with biliary trematodiasis, little has research has been conducted with the spectrum of lesions reported in cetaceans. It seems that the massive infestation by this parasite should be considered a differential diagnosis of hepatic failure resulting in stranding of marine mammals.

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