






Positive species interactions structure rhodolith bed communities at a global scale

Fabio Bulleri^{1,*} , Nadine Schubert² , Jason M. Hall-Spencer^{3,4} , Daniela Basso⁵, Heidi L. Burdett^{6,7}, Ronaldo B. Francini-Filho⁸, Jacques Grall⁹, Paulo A. Horta¹⁰, Nicholas A. Kamenos^{6,7}, Sophie Martin¹¹, Matteo Nannini¹² , Pedro Neves^{2,13}, Irene Olivé¹² , Viviana Peña¹⁴, Federica Ragazzola^{15,16}, Cláudia Ribeiro^{2,17}, Eli Rinde¹⁸, Marina Sissini¹⁹, Fernando Tuya²⁰ and João Silva²

¹*Dipartimento di Biologia, Università di Pisa, Via Derna 1, Pisa 56126, Italy*

²*Centre of Marine Sciences (CCMAR/CIMAR LA), Campus de Gambelas, Universidade do Algarve, Faro 8005-139, Portugal*

³*Shimoda Marine Research Center, University of Tsukuba, Shizuoka, Japan*

⁴*School of Biological and Marine Sciences, University of Plymouth, Plymouth, UK*

⁵*Department of Earth and Environmental Sciences, University of Milano–Bicocca, CoNISMa Research Unit of Milano–Bicocca, Milan, Italy*

⁶*Umeå Marine Sciences Centre, Umeå University, Norrbyn, Sweden*

⁷*Department of Ecology and Environmental Sciences, Umeå University, Umeå, Sweden*

⁸*Laboratório de Biodiversidade e Conservação Marinha, Centro de Biologia Marinha (CEBIMar), Universidade de São Paulo (USP), São Sebastião, Brazil*

⁹*UAR 3113 OSU, Institut Universitaire Européen de la Mer, Univ Brest, Plouzané, France*

¹⁰*Laboratório de Ficologia, Departamento de Botânica, Centro de Ciências Biológicas, Universidade Federal de Santa Catarina, Florianópolis, Brazil*

¹¹*UMR 7144 Adaptation et Diversité en Milieu Marin, CNRS, Sorbonne Université, Station Biologique de Roscoff, Roscoff, France*

¹²*Department of Integrative Marine Ecology, Stazione Zoologica Anton Dohrn, Villa Comunale, Naples, NA 80121, Italy*

¹³*Observatório Oceânico da Madeira, Agência Regional para o Desenvolvimento da Investigação, Tecnologia e Inovação (OOM/ARDITI), Funchal, Madeira, Portugal*

¹⁴*BioCost Research Group, Faculty of Sciences, University of A Coruña, rúa da Fraga 10, A Coruña 15008, Spain*

¹⁵*Department of Integrative Marine Ecology, Genoa Marine Centre, Stazione Zoologica Anton Dohrn, 9 Villa del Principe, Piazza del Principe 4, Genoa 16126, Italy*

¹⁶*NBFC, National Biodiversity Future Center, Palermo 90133, Italy*

¹⁷*IFCN—Instituto das Florestas e Conservação da Natureza, IP-RAM, Funchal, Madeira, Portugal*

¹⁸*Norwegian Institute for Water Research, Oslo, Norway*

¹⁹*Department of Marine Biology, Federal Fluminense University, Niterói, Rio de Janeiro, Brazil*

²⁰*Grupo en Biodiversidad y Conservación (IU-ECOQUA), Universidad de Las Palmas de Gran Canaria, Telde, Spain*

ABSTRACT

Rhodolith beds are diverse and globally distributed habitats. Nonetheless, the role of rhodoliths in structuring the associated species community through a hierarchy of positive interactions is yet to be recognised. In this review, we provide evidence that rhodoliths can function as foundation species of multi-level facilitation cascades and, hence, are fundamental for the persistence of hierarchically structured communities within coastal oceans. Rhodoliths generate facilitation cascades by buffering physical stress, reducing consumer pressure and enhancing resource availability. Due to large variations in their shape, size and density, a single rhodolith bed can support multiple taxonomically distant and architecturally distinct habitat-forming species, such as primary producers, sponges or bivalves, thus encompassing a broad range of functional traits and providing a wealth of secondary microhabitat and food resources. In addition, rhodoliths are often mobile, and thus can redistribute associated species, potentially expanding the distribution of species with

* Author for correspondence (Tel.: +39 050 2211 448; E-mail: fabio.bulleri@unipi.it).

short-distance dispersal abilities. Key knowledge gaps we have identified include: the experimental assessment of the role of rhodoliths as basal facilitators; the length and temporal stability of facilitation cascades; variations in species interactions within cascades across environmental gradients; and the role of rhodolith beds as climate refugia. Addressing these research priorities will allow the development of evidence-based policy decisions and elevate rhodolith beds within marine conservation strategies.

Key words: rhodoliths, encrusting coralline algae, foundation species, maerl beds, benthic habitats, facilitation cascades, marine biodiversity.

CONTENTS

I. Introduction	2
II. Rhodolith and maerl beds	3
III. Variation in diversity and life traits of rhodoliths across environmental gradients	4
IV. Potential of rhodoliths to modify biotic and abiotic conditions	4
(1) Amelioration of environmental conditions	5
(2) Reduction of predation and competition pressure	5
(3) Enhancement of resource availability	5
V. Multiple facilitation cascades in rhodolith beds	6
(1) Seaweeds	7
(2) Sponges	9
(3) Bivalves	10
VI. Research needs and future directions in the context of facilitation cascades	10
(1) Experimental evaluation of the role of rhodoliths as basal facilitators	10
(2) The length of facilitation cascades and negative feedbacks on rhodoliths	10
(3) Variability across environmental gradients	12
(4) The temporal stability of cascades	12
(5) Rhodolith beds as climate refugia	13
VII. Conclusions	13
VIII. Acknowledgements	13
IX. References	13

I. INTRODUCTION

Advances in our understanding of the mechanisms that underpin patterns of species distribution and abundance show that positive species interactions are a driving force in the organisation of natural communities (Bertness & Callaway, 1994; Stachowicz, 2001; Bruno, Stachowicz & Bertness, 2003; Michalet *et al.*, 2006; Brooker *et al.*, 2008; Gross, 2008; Bulleri *et al.*, 2016). Foundation species, defined as those often making up most of the biomass in an ecosystem and located at or near the base of directional networks of mutualistic or non-trophic interactions (*sensu* Ellison, 2019), including trees, corals, seagrasses, oysters, mussels, salt-marsh plants and seaweeds, can sustain highly biodiverse habitats (Altieri, Silliman & Bertness, 2007; Silliman *et al.*, 2011). The physical structure of these foundation species enhances the establishment and persistence of other species *via* three mechanisms: (i) amelioration of environmental stress; (ii) reduction of consumer or competition pressure; and (iii) increased resource availability. These mechanisms support the more generic notion of foundation species as habitat-formers (Gribben *et al.*, 2019).

Many marine communities are structured by a hierarchy of positive interactions triggered by the presence of

foundation species (Bruno & Bertness, 2001). For example, on intertidal cobble beaches, shading and substratum stabilisation by the cordgrass, *Spartina alterniflora*, promotes the presence of other species, such as mussels, snails and seaweeds (Altieri *et al.*, 2007). Likewise, cockles can expand seaweed distribution onto sandy or muddy bottoms by providing hard substrata for attachment (Gribben *et al.*, 2009). Species directly sustained by foundation species can, in turn, act as secondary facilitators, generating a facilitation cascade (*sensu* Altieri *et al.*, 2007). The presence of the basal or primary facilitator is a prerequisite for the presence of further habitat-formers (secondary or upper-level facilitators). This establishes the hierarchical structure of the whole community, ultimately enhancing the availability of microenvironments and/or resources (Thomsen *et al.*, 2010, 2018). Facilitation cascades are often size-structured (*sensu* Thomsen *et al.*, 2016), whereby body size decreases progressively when moving from the basal to upper-level facilitators. For example, mangrove trees, seagrass or salt-marsh plants are generally larger than the bivalves or macroalgae they support and that function as secondary facilitators for other invertebrates (Edgar & Robertson, 1992; Altieri *et al.*, 2007; Bishop *et al.*, 2012). However, in marine environments, facilitation cascades in which relatively small-bodied foundation species,

such as mussels, cockles or tubeworms, support larger secondary facilitators, often macroalgae, have been broadly documented (Witman 1987; Bulleri & Airoldi, 2005; Thomsen *et al.*, 2016, 2022; Bracken, 2018; Ape *et al.*, 2018). Independently from the size-structure of the cascade, this chain or web of positive interactions often culminates with species that do not form habitat, but may be relevant as key elements for ecosystem functioning.

Facilitation cascades have been documented in a variety of terrestrial and marine systems globally, including coral reefs, temperate and tropical forests, salt marshes, soft-bottoms, seagrass meadows, mangroves and kelp forests (Crain & Bertness, 2006; Thomsen *et al.*, 2010, 2018, 2022; Gribben *et al.*, 2019). For instance, on tidal mudflats, aerial mangrove roots provide substrata for oyster recruitment and entangle drifting algae, which, in turn, generate suitable habitat for a variety of gastropods (Bishop *et al.*, 2012). Similarly, epiphytes growing on trees, seagrasses or freshwater plants can host diverse assemblages of invertebrates (Angelini & Silliman, 2014; Thomsen *et al.*, 2022). So far, cascades of up to five levels of co-occurring habitat-formers have been documented (Thomsen *et al.*, 2016).

Several aspects of marine facilitation cascades, including underlying mechanisms, spatial configuration (embedded *versus* adjacent), trait- and density-mediated effects, variation across environmental gradients and the relationship between cascade length and temporal stability, have been reviewed recently (Gribben *et al.*, 2019). However, out of 100 papers reviewed, none dealt with habitats formed by free-living red coralline algae (Gribben *et al.*, 2019). These so-called rhodolith or maerl beds are globally distributed and can be highly biodiverse (Fig. 1). This would indicate that the role of

rhodoliths as basal habitat-formers has been overlooked by the ecological facilitation scientific community (but see Otero-Ferrer *et al.*, 2019). Despite rhodolith beds being 2.5–30 times more extensive than habitats formed by widely recognised coastal foundation species (e.g. mangroves, seagrasses and kelps), the number of studies on rhodolith beds is disproportionately small (Rendina *et al.*, 2022; Tuya *et al.*, 2023), leaving a significant gap in our understanding of their ecological structure, community assembly rules and ecosystem functioning.

Herein, we aim to (i) summarise patterns of species and functional diversity of rhodolith beds and their associated communities, (ii) identify the mechanisms underpinning rhodolith modification of biotic and abiotic conditions and resource availability, (iii) review evidence for the role of rhodoliths as basal species and, hence, as promoters of hierarchical organisation of associated communities, (iv) identify their role in sustaining biodiversity and ecosystem functioning, and (v) highlight priority research needs and future directions to assess facilitation cascades in rhodolith beds.

II. RHODOLITH AND MAERL BEDS

The term rhodolith (*rhodo* = red + *lith* = stone) was formally coined by Bosellini & Ginsburg (1971) as “rhodolite”, later corrected to the current spelling to avoid confusion with a garnet. In their original definition, the name referred to unattached nodules formed by calcareous red algae and their branched growths, as part of a continuous spectrum of forms and structures, with size spanning from 2 to 250 mm mean

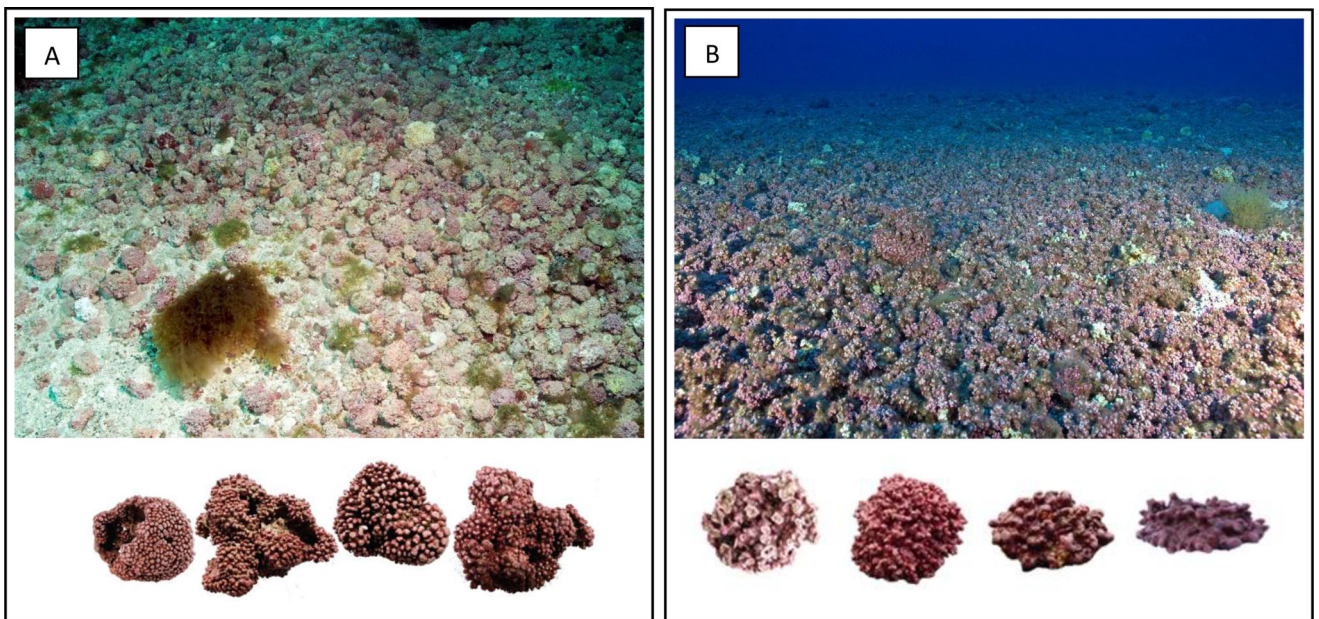


Fig. 1. (A) Rhodolith beds from the Fernando de Noronha Archipelago, Brazil, ~ 20 m depth (photograph credit: Ronaldo Francini-Filho). (B) Rhodolith beds from the Madeira Archipelago, Portugal (photograph credit: Pedro Neves). Bottom images illustrate maerl and rhodolith nodules of different shapes (photograph credits: Eli Rinde and João Silva).

diameter. The term rhodolith parallels the names of other types of unattached nodules composed by different organisms – such as bryolith for unattached bryozoan nodules (James, Foster & O’Sullivan, 2006), corallith for unattached coral nodules (Glynn, 1974) – using nomenclature based on the nodule builder – and includes both nucleated and non-nucleated nodules of calcareous red algae.

The term “maerl” comes from a Breton word, referring to an area of calcareous land or marine deposits of calcified algae (Grall & Hall-Spencer, 2003). In northeastern Atlantic countries, the term maerl has been used for centuries to indicate mostly twig-like, frequently intertwined, unattached coralline algae forming thick accumulations on the shallow seabed. Based on these definitions, the term “rhodolith beds” includes beds made of nucleated and non-nucleated nodules (i.e. maerl beds) and calcareous *Peyssonnelia* beds (Steller *et al.*, 2003; Steller & Foster, 1995; Foster *et al.*, 2013). Herein, we will use “rhodoliths” to include both rhodolith and maerl forms. It is worth noting that there is no clear definition of a rhodolith bed in terms of coverage and proportion of live *versus* dead nodules. Rhodolith beds have been variously defined as areas with a cover of living coralline thalli >10% (Steller *et al.*, 2003) or >30% (OSPAR, 2008), within an area of at least 100 m² according to Rinde *et al.* (2022).

III. VARIATION IN DIVERSITY AND LIFE TRAITS OF RHODOLITHS ACROSS ENVIRONMENTAL GRADIENTS

The potential of rhodoliths to act as foundation species and to enable facilitation is regulated by their size, shape and abundance – attributes that can vary within and among species. The spatial extent and degree of patchiness of rhodolith beds also influence their ability to initiate a facilitation cascade, as well as the number of levels in a cascade, due to different habitat requirements of the associated species (Gribben *et al.*, 2019). Thus, understanding the factors shaping variations in the structure of rhodolith beds is crucial for assessing their role as basal species in facilitation cascades.

Variability in rhodolith distribution, shape and morphology has long been observed (Bosellini & Ginsburg, 1971; Bosence, 1983; Basso, 1998). Rhodolith nodules can be composed of one single morphotype (e.g. maerl beds composed of variably shaped unattached branches; Bosence, 1983; Basso *et al.*, 2016), by mixtures of coralline morphotypes (Bracchi *et al.*, 2022; Vale *et al.*, 2022) or by coralline and *Peyssonnelia* nodules. There are examples of highly diverse rhodolith beds, such as that around Punta de la Mona (western Mediterranean), which is formed by 25 morphospecies belonging to six genera (*Lithophyllum*, *Spongites*, *Neogoniolithon*, *Lithothamnion*, *Mesophyllum* and *Phymatolithon*) (Del Rio *et al.*, 2022).

The heterogeneity of the physical structure of rhodolith beds can be described with a ternary diagram that considers the three main morphotypes of nodules [pralines, unattached branches or boxwork (Basso, 1998; Basso,

Nalin & Nelson, 2009)], allowing a visual description of the rhodolith bed (Basso *et al.*, 2016; Bracchi *et al.*, 2022; Caronni *et al.*, 2023). The shape of rhodoliths is often measured following the criteria of Sneed & Folk (1958), which requires measuring the longest, intermediate, and shortest (axial) diameters (Sciberras *et al.*, 2009) and using these measures to classify nodules into four classes – spheroidal, discoidal, ellipsoidal or bladed (Bosence, 1976; Gagnon, Matheson & Stapleton, 2012; Carro *et al.*, 2014; Villas-Bôas *et al.*, 2014; Otero-Ferrer *et al.*, 2020; Neves *et al.*, 2021) – as well as to calculate sphericity (Voerman *et al.*, 2022).

Light, temperature, hydrodynamics and sedimentation are among the main factors regulating the characteristics of rhodolith assemblages (Basso, 1998; Carvalho *et al.*, 2020; Otero-Ferrer *et al.*, 2020; Sissini *et al.*, 2022). Rhodolith size and shape are mostly regulated by hydrodynamics, whilst light limitation or burial under sediments can stunt growth (Villas-Bôas *et al.*, 2014; Bracchi *et al.*, 2019; Omachi *et al.*, 2019). Features of coastlines (e.g. presence of sheltered bays or inlets) and the extension of the continental shelf influence these physical factors and, hence, may play a role in regulating the size, shape and density of rhodolith nodules. For example, along bathymetric gradients, the largest rhodoliths have been found in the shallowest areas at some sites (Bahia *et al.*, 2010; Voerman *et al.*, 2022), while they are located more often in intermediate or deepest areas sampled at other sites (Sañe *et al.*, 2016; Del Rio *et al.*, 2022; Perez-Peris *et al.*, 2023). This suggests that location-specific environmental conditions can be more important than factors covarying with depth (e.g. light, temperature, sedimentation and hydrodynamic forces) in determining rhodolith size and/or abundance. However, in the northeast Atlantic, the shape of rhodoliths and complexity of the bed they formed were weakly influenced by bottom currents and wind exposure, and driven instead by underlying sediment composition (Jardim *et al.*, 2022). In addition, the shape of rhodoliths is influenced by biotic factors. For example, boring bivalves can produce cavities in nodules, providing further niche space for both vertebrates and invertebrates (Gagnon *et al.*, 2012; Teichert, 2014).

IV. POTENTIAL OF RHODOLITHS TO MODIFY BIOTIC AND ABIOTIC CONDITIONS

Rhodoliths are widely recognised as ecosystem engineers (Nelson, 2009; Teichert & Freiwald, 2014; Qui-Minet *et al.*, 2018; Otero-Ferrer *et al.*, 2019; Voerman *et al.*, 2022) and there is compelling evidence that they support higher invertebrate species diversity compared to surrounding soft sediments (Steller *et al.*, 2003; Teichert, 2014; Boye *et al.*, 2019; Neves & Costa, 2022) and equivalent fish species diversity compared to adjacent coral reefs (Moura *et al.*, 2021; Anderson *et al.*, 2023). Nonetheless, few studies have formally identified the mechanisms underpinning facilitation, which requires experimental manipulation of biotic (e.g. manipulation of consumer or competition

pressure) and/or abiotic conditions (e.g. environmental stressors) (Bulleri, 2009; Thomsen *et al.*, 2018, 2022; Gribben *et al.*, 2019). Instead, the positive effects of rhodoliths on other species have been widely explained through the general effect of (micro)habitat creation, since nodule and bed complexity generate multiple niches for associated organisms. Below, we provide some examples of rhodolith beds enhancing the settlement, growth and survival of benthic species for each of the three facilitation mechanisms. Note that this is not meant to be a comprehensive review of the literature and here we distinguish among the different facilitation mechanisms for clarity, although they are likely to co-occur due to simultaneous changes in both biotic and abiotic conditions induced by rhodoliths.

(1) Amelioration of environmental conditions

Although the amelioration of environmental conditions is not a predominant mechanism of species facilitation in subtidal environments (Bulleri, 2009), intense hydrodynamic forces and high sediment load can generate adverse conditions for benthic species (Witman, 1987; Bulleri *et al.*, 2011). Rhodolith beds can thrive in wave- and/or current-swept areas, yet their rigid calcareous thalli can reduce hydrodynamic forces and trap finer sediments, providing shelter and trophic resources to other species (Hall-Spencer, 1998; Gabara, 2020). Experiments showed that irregularly shaped rhodoliths can enhance substratum stability in wave-swept environments (Joshi, Duffy & Brown, 2017), regulate sediment grain size distribution (de Queiroz *et al.*, 2016) and create interstices that favour oxygen penetration to deeper layers (Hall-Spencer & Atkinson, 1999), ultimately favouring colonisation by both epibenthic and infauna species (Caronni *et al.*, 2023). Rhodoliths have also been documented on intertidal flats, but reported densities are too low ($<1\text{ m}^2$) for the formation of habitat (Perry, 2005), and, hence, their potential to facilitate species through the buffering of heat and desiccation stress remains uncertain.

Coralline algae, including rhodoliths, alter seawater chemistry through their metabolic activity (i.e. photosynthesis, respiration and calcification) by producing O_2 , and modifying pH, HCO_3^- and CO_3^{2-} concentrations at their surface with respect to the surrounding water (Hofmann, Schoenrock & de Beer, 2018; McNicholl, Koch & Hofmann, 2019; Schubert *et al.*, 2021). It is likely that these changes in seawater chemistry provide more suitable conditions for other non-calcifying species, at least during daytime. This feature is likely to become even more relevant under projected levels of ocean acidification and warming (Cornwall *et al.*, 2022).

(2) Reduction of predation and competition pressure

By providing a three-dimensional structure, rhodolith beds act as nursery areas for both vertebrate and invertebrate species, including those of commercial importance; this is likely

due to the reduction of competition pressure they provide *via* refuge creation (Kamenos, Moore & Hall-Spencer, 2004a). For example, juvenile cod (*Gadus morhua*), saithe (*Pollachius virens*) and pollack (*Pollachius pollachius*) were more abundant in rhodolith habitats than adjacent heavily vegetated rocky and gravel substrata (Kamenos, Moore & Hall-Spencer, 2004c). Rhodolith beds have a high holding capacity for juvenile gadoids, likely by virtue of their food and refuge provisioning, and are thus an important part of the inshore nursery system (Kamenos, Moore & Hall-Spencer, 2004d). The sand tilefish *Malacanthus plumieri* is almost exclusively found on rhodolith beds, building rhodolith mounds that are used as a refuge during foraging movements, shelter against predation and may also play a role in social organisation (Pereira *et al.*, 2015). The mounds themselves are used as microhabitats for several other fish and invertebrate species (Pereira *et al.*, 2015; Francini *et al.*, 2018). Similarly, hollow rhodoliths, formed by boring bivalves, are known to function as nesting sites for fish and for shelter from predation for several species of ophiurids (Gagnon *et al.*, 2012; Teichert, 2014). The positive effects of rhodoliths through the reduction of consumer pressure are not limited to the formation of interstices that can be used by prey as refuges from predators. Leemans *et al.* (2020) demonstrated experimentally that spiky rhodoliths promoted the recovery and persistence of the seagrass *Thalassia testudinum* by directly reducing the access of marine turtles to plants. Likewise, juvenile bivalves of many species have been found at higher densities in rhodoliths compared to surrounding substrata (Kamenos *et al.*, 2004c; Steller & Caceres-Martinez, 2009). This is thought to be due to attraction to the living coralline algal surface and also the presence of a rugose three-dimensional structure (Kamenos, Moore & Hall-Spencer, 2004b).

There is evidence that the presence of the living algal veneer acts as an attractant to juvenile scallops, signalling refuge presence and endowing lower stress responses in the presence of predatory starfish (Kamenos, Calosi & Moore, 2006). The attractant effect may be facilitated by the high production of dimethylsulphide (DMS) and its secondary metabolite precursor dimethylsulphoniopropionate (DMSP) by rhodoliths, especially at high and low latitudes (Burdett, Hatton & Kamenos, 2015; Burdett, 2017). Both compounds are important chemical cues for a range of ecological processes, including herbivorous grazing and vertebrate larval settlement (Lyons, Scheibling & Van Alstyne, 2010; Foretich *et al.*, 2017).

(3) Enhancement of resource availability

Biogenic substrata, primarily formed by rhodoliths and often consolidated by binding species, such as sponges or mussels, may extend the distribution of hard-bottom-dwelling species to soft sediments, which can play an important ecological role in places with reduced availability of natural rocky bottoms (Ávila, Riosmena-Rodriguez & Hinojosa-Arango, 2013; Pereira *et al.*, 2015). This phenomenon has been recorded in estuaries, deep reefs in oceanic islands with reduced shelf

and on seamounts (Steller *et al.*, 2003; Pereira *et al.*, 2012; Otero-Ferrer *et al.*, 2019). For example, in the Madeira Archipelago, rhodolith beds considerably increase the availability of subtidal hard substrata around the islands (Neves *et al.*, 2021). The provisioning of consolidated, biogenic substrata could be highly relevant also on hard bottoms since, despite the availability of hard surfaces, the settlement and growth of benthic species can be facilitated or, indeed, restricted to living coral-line algal beds (Kamenos *et al.*, 2004a; Steller & Caceres-Martinez, 2009). Vertical expansions of species distribution (i.e. bathymetric) fostered by rhodoliths might be particularly relevant in the context of global warming (see Section VI).

Rhodolith beds increase food availability for associated organisms by entraining organic matter and promoting local small-scale primary production from microphytobenthos and soft red seaweeds, thus increasing secondary production (Grall *et al.*, 2006; Gabara, 2020; Neto, Bernardino & Netto, 2021; Teper, Parrish & Gagnon, 2022). This is consistent with analyses of the trophic groups associated with rhodolith beds, revealing a dominance of deposit feeders (Grall *et al.*, 2006; Sciberras *et al.*, 2009; Teper *et al.*, 2022). The higher resource availability and diversity of food sources in rhodolith beds may ensure a more constant resource supply than in bare habitats. Indeed, stable isotope analyses have suggested that Arctic rhodolith beds may function as benthic–pelagic hotspots, at least seasonally (Teper *et al.*, 2022). Finally, the high functional richness and redundancy of associated communities (Boye *et al.*, 2019) may promote temporal community stability through compensatory dynamics and asynchronous species temporal fluctuations within functional groups (Magurran & Henderson, 2018).

V. MULTIPLE FACILITATION CASCADES IN RHODOLITH BEDS

Many of the species or groups of species that are dependent upon rhodoliths for colonisation of soft sediment areas can act as secondary facilitators. According to the Foundation Species-Biodiversity (FSB) model (Angelini & Silliman, 2014), the difference in morphological traits between the basal and the secondary facilitator determines the strength of the effects on species diversity. The larger the difference, the greater the positive effect of the association between the basal species and secondary facilitators on species diversity at upper cascade levels. By contrast, the addition of a secondary facilitator that does not differ from the basal species in terms of morphological complexity would simply increase habitat availability, enhancing the abundance, but not the diversity, of species within the assemblage associated with the basal species. The effects of secondary facilitators on species abundance and diversity can be expected to be very high in rhodolith beds since they support a broad variety of secondary facilitators. Rhodolith beds are often composed of different rhodolith morphospecies (up to 25 in a single bed; Del Rio *et al.*, 2022), with

subsequent large variations in functional traits (e.g. size, shape, rugosity, branching) and thereby, high microhabitat diversity. Indeed, although unexplored, some rhodolith morphospecies could facilitate others. For example, highly branched forms could enhance the retention of simpler nodules, by reducing their drag by currents. This means that, conceptually, the first two levels of a facilitation cascade (i.e. the basal species and the secondary facilitator) could both be represented by rhodoliths. High within-bed rhodolith diversity should therefore broaden the pool of species they can support at the seascape scale and, hence, increase the likelihood of secondary facilitator establishment. In addition, a single rhodolith bed can support multiple taxonomically distant and architecturally distinct species (e.g. Porifera, Cnidaria, Mollusca, Ochrophyta, Chlorophyta, other Rhodophyta), with a broad range of functional traits and hence, a wealth of secondary microhabitat and food resources provision. Rhodoliths, by supporting many secondary facilitators that differ morphologically, are expected to amplify positive effects on species diversity.

Two features that distinguish rhodoliths from most foundation species are their small size and mobility. Many of the widely acknowledged marine foundation species, including seagrasses, mangroves, kelps and corals, are generally conspicuous in size and larger than the species they support through their facilitative effects. Thus, in most cases, there is a clear decreasing pattern in body size when moving from the basal species to the upper levels of facilitation cascades. This does not necessarily apply to rhodoliths, as they are often smaller than the species they support that act as secondary facilitators, such as many seaweeds, sponges and bivalves. Thus, despite being dominant in abundance, and at the base of a network of positive, non-trophic species interactions (in accordance with the definition by Ellison, 2019), rhodoliths markedly deviate from the common view of foundation species. Although unexplored, the smaller size of rhodoliths – in comparison to that of the species they support – could increase the likelihood of negative feedbacks. For example, epibiota (e.g. sponges, oysters) and epiphytes (macroalgae) on the roots of mangroves (Bishop *et al.*, 2012; Gribben *et al.*, 2019) are unlikely to have negative effects on the trees, while they could completely overgrow rhodoliths, hampering their movement and impairing their photosynthetic efficiency.

Rhodoliths can be transported by waves and currents (Lavenère-Wanderley *et al.*, 2021). The only other known example of a mobile foundation species is that of the pencil sea urchins in the Galapagos (Altieri & Witman, 2014). The mobility of rhodoliths implies that they can redistribute associated species, potentially expanding the distribution of species with short-distance dispersal abilities. Widespread rolling and movement of rhodoliths occurs sporadically because of storms, but also periodically, due to currents and wind-propagated waves [up to several centimetres per day (Steller & Foster, 1995; Harris *et al.*, 1996; Marrack, 1999)]. Overgrowth by large erect species, such as macroalgae and sea fans, is likely to increase the distance over which rhodoliths can be dragged by currents and waves. Moreover,

associated fauna can contribute to rhodolith displacement through bioturbation, for example movement of sea urchins that use rhodoliths as covering material (Foster *et al.*, 1997; Marrack, 1999) and the activity of the sand tilefish that moves rhodoliths to build mounds, likely playing an important role in the spatial extension of rhodolith beds (Pereira *et al.*, 2015). Displacement by biotic or abiotic factors does not transport rhodoliths exclusively horizontally, but can cause their spilling down from shallow-water environments to greater depths, such as in the case of steep slopes of seamounts, where rhodoliths have been found at a depth of up to 290 m (Littler, Littler & Hanisak, 1991). On the other hand, some epifauna (e.g. sponges, tunicates, anemones) can also bind rhodoliths together, decreasing their movement (Marrack, 1999).

Although rhodolith survival is dependent upon light availability, their calcified skeleton means that even dead nodules maintain their shape and can continue to support a high diversity of associated species (Kamenos *et al.*, 2004a,b). As acknowledged for other foundation species (Saldaña *et al.*, 2024), rhodoliths are therefore likely to maintain their foundation species effect beyond their lifetime and in areas unsuitable for their survival.

To the best of our knowledge, no study has, to date, experimentally demonstrated facilitation cascades (i.e. the co-occurrence of the basal species, i.e. the primary facilitator, and a secondary facilitator or a focal species that does not form habitat) in rhodolith beds. Nonetheless, based on the available information and to illustrate the potential of rhodoliths to start facilitation cascades, we here focus on three taxa, seaweeds, sponges and bivalves (Fig. 2), that are commonly associated with rhodoliths and are broadly known to function as primary or secondary facilitators (Dayton, 1972; Steneck *et al.*, 2002; Gribben *et al.*, 2009; Bishop *et al.*, 2012; MacDonald & Weis, 2013; van de Koppel *et al.*, 2015; Thomsen *et al.*, 2018, 2022; Ravaglioli *et al.*, 2021). This does not exclude the potential for other taxa to be part of rhodolith facilitation cascades. Moreover, our examples are focused on facilitation cascades in which the rhodoliths and the secondary benefactors are embedded within the same patch (i.e. nested facilitation, *sensu* Angelini *et al.*, 2011). However, as shown for other facilitative interactions, such as those involving mangroves, seagrasses, corals, oysters and marsh plants (van de Koppel *et al.*, 2015; Gribben *et al.*, 2019), positive effects of rhodoliths could expand beyond the margins of the bed they form to influence other habitats (i.e. adjacent or landscape facilitation, *sensu* Angelini *et al.*, 2011). For instance, some of the biomass generated by invertebrates and macroalgae preferentially recruiting into rhodolith beds might move outside, either actively or passively, fuelling adjacent bare-sand trophic webs. Likewise, rhodolith-driven changes in water chemistry (i.e. pH and CO₃²⁻) or bottom currents could affect adjacent habitats, such as coralligenous or coral reefs and seagrass beds.

(1) Seaweeds

Seaweed assemblages associated with rhodolith beds are known to be extremely diverse. For instance, northeast

Atlantic maerl beds host at least 349 associated macroalgal species, including 232 Rhodophyta, 72 Heterokontophyta and 45 Chlorophyta, making up about 30% of the total macroalgal diversity in the region (Peña *et al.*, 2014). Likewise, Helias & Burel (2023) recorded 170 macroalgal species, belonging to Rhodophyta, Ochrophyta and Chlorophyta in rhodolith beds from the Bay of Brest, 108 of which were found growing exclusively on rhodoliths, while 14 occurred only as epiphytes on other species, indicating two and three levels of co-occurring habitat-formers. In Brazil, local estimates (across areas of tens of km²) of seaweed diversity in rhodolith beds range from 44 to 67 species, with regional estimates (i.e. across hundreds of km² in the Abrolhos Bank) of 146 species (Riul *et al.*, 2009; Pascelli *et al.*, 2013; Brasileiro *et al.*, 2016).

Rhodolith beds also host kelp species (Fig. 2A), which are themselves foundation species (Steneck *et al.*, 2002; Fernandez, 2011; Bracken, 2018). Association of kelp species (e.g. *Saccharina latissima*, *Saccorhiza polyschides*, *Laminaria hyperborea*, *L. abyssalis*, *L. ochroleuca*, *L. rodriguezii*, *Ecklonia radiata*) with rhodoliths has been documented in both tropical and temperate basins, including the Mediterranean, and the coasts of France, New Zealand, Brazil and the Madeira islands (Amado *et al.*, 2007; Peña & Bárbara, 2008; Nelson, 2009; Amado-Filho & Pereira-Filho, 2012; Barberá *et al.*, 2012; Marins *et al.*, 2014; Peña *et al.*, 2014; Braga-Henriques *et al.*, 2022). In some cases, such as that of *L. rodriguezii* in the Menorca Channel (western Mediterranean), kelps can comprise most of the macroalgal biomass associated with rhodolith beds. While large kelp individuals, and the rhodoliths to which their haptera are attached, can be dislodged from rhodolith beds in shallow, wave-swept areas, kelp populations on deeper beds are likely to be more stable. For example, deep (45–120 m) rhodolith beds of the Abrolhos Bank (Brazil) host permanent populations of *L. abyssalis* (Amado-Filho & Pereira-Filho, 2012; Foster *et al.*, 2013). Thus, rhodolith beds can extend the distribution of this marine foundation species to areas that would otherwise lack suitable hard surfaces (Foster *et al.*, 2013), creating a facilitation cascade that enhances local and regional species diversity (Fig. 3A).

Epiphytic macroalgae supported by rhodoliths, including filamentous and fleshy morphotypes (Fig. 2B), can also provide shelter against predators and resources (space and food) for fish (Chaves, Pereira & Feitosa, 2013; Fulton *et al.*, 2020) and for sessile and mobile invertebrates, as widely demonstrated for epiphytes on seagrasses and macroalgae (Thomsen *et al.*, 2018; Ravaglioli *et al.*, 2021; El-Khaled *et al.*, 2022). Thus, although not considered as foundation or habitat-forming species, epiphytic macroalgae can still function as secondary or tertiary facilitators (Fig. 3). In addition, some macroalgal species, such as *Codium bursa* and *C. cf. effusum*, were exclusively found free-living (i.e. unattached to the bottom) within rhodolith beds in the Bay of Brest, France (Helias & Buriel, 2023) and in Sardinia (D. Basso, personal observations). This suggests that facilitation can occur at scales varying from that of the single

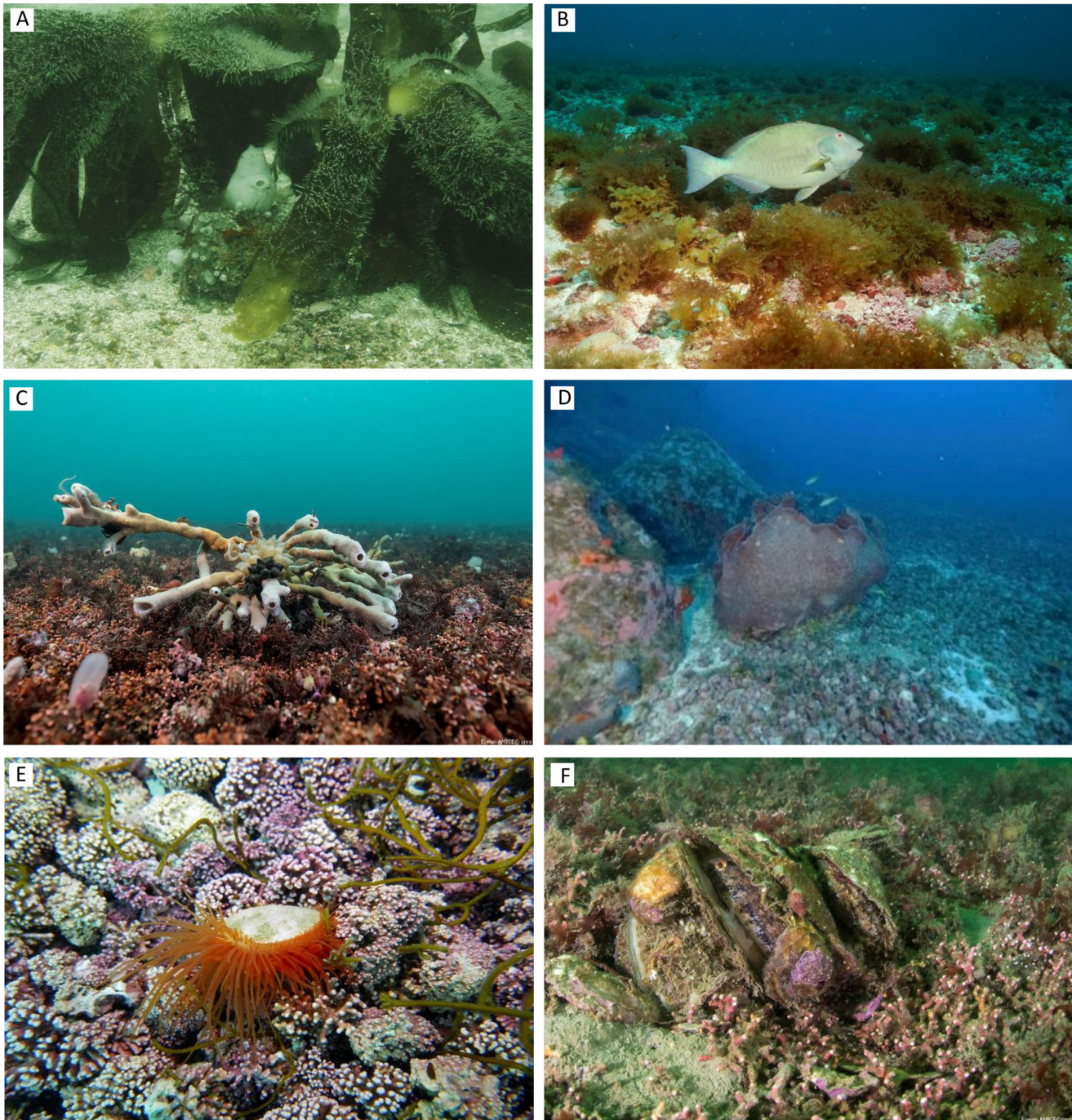


Fig. 2. Habitat-forming species in rhodolith/maerl beds. (A) *Laminaria ochroleuca* from Galicia, Spain, depth 11 m (photograph credit: Ignacio Bárbara). (B) Diverse macroalgal assemblages supported by rhodoliths at the Fernando de Noronha Archipelago, Brazil, depth 40 m (photograph credit: Zaira Matheus). (C) The tubular sponge, *Haliclona simulans*, supporting ophiurans, gastropods and hosting cuttlefish eggs in Brittany, France, depth 7 m (photograph credit: Erwan Amice). (D) Unidentified concave sponge from the Fernando de Noronha Archipelago, Brazil, depth 40 m (photograph credit: Zaira Matheus). (E) The bivalve *Limaria hians* in the north of Norway, depth 15 m (photograph credit: Jason Hall-Spencer). (F) The flat oyster *Ostrea edulis* supporting fish spawning, hydroids, seaweeds, encrusting sponges, ascidians and galatheidae in a maerl bed of Brittany, France, depth 3 m (photograph credit: Erwan Amice).

rhodolith (by providing a surface for attachment) to that of the whole bed, which, by virtue of its complex topography, acts as a passive collector of macroalgae, comparable to

mangrove pneumatophores in tidal flats (Bishop *et al.*, 2012; Bastos *et al.*, 2013). This may be important in sustaining a large biomass of fleshy organic carbon which can then be

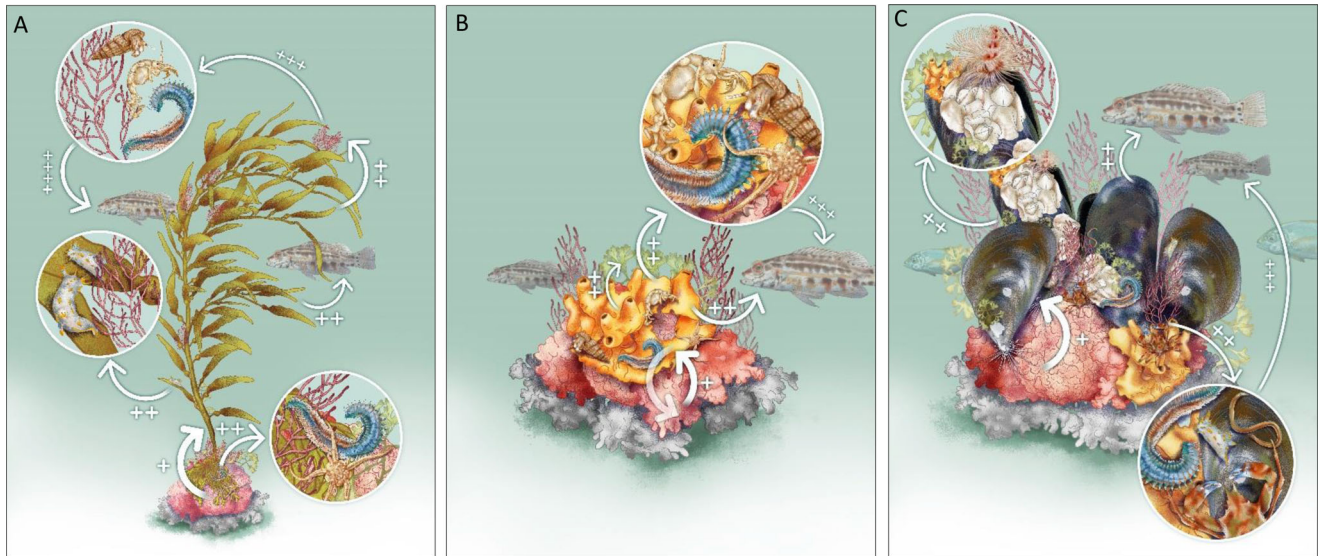


Fig. 3. Positive interactions within facilitation cascades in rhodolith beds. (A) Rhodoliths (the basal species) promote the establishment of kelp on sedimentary bottoms through the provisioning of hard substrates for attachment. The holdfast, stipe and blades of kelp (secondary facilitator) can provide habitat and/or food to fish and invertebrates, directly or indirectly by supporting epibiota (e.g. macroalgae) that act as tertiary facilitators. (B) Rhodoliths facilitate the establishment of sponges which, in turn, can stabilise beds through the binding of nodules. Sponges (secondary facilitators) can provide habitat for a variety of invertebrates and sustain the growth of macroalgae by recycling nutrients. (C) Bivalves growing on rhodoliths can facilitate invertebrates and macroalgae by providing interstitial or attachment space. In all cases, species at the second or upper level within the facilitation cascade can be used as food by consumers (i.e. herbivores or predators). Arrows show the positive effects of one species on another, with the number of plus signs indicating the level of the facilitative interaction within the cascade. Some of the functions performed by rhodoliths, such as the creation of microhabitat and provisioning of attachment surface, can continue when they are dead (represented in grey); for convenience, dead rhodoliths are illustrated below the living layer, although, in real beds, the surficial layer is often composed of a variable proportion of live and dead nodules. Magnifying lenses provide details of macroalgae and invertebrates supported by secondary facilitators (Illustration by ©Lúcia Antunes, www.luciaantunes.com).

buried locally in the rhodolith bed, thus acting as a blue carbon repository (James *et al.*, 2024; Mao *et al.*, 2020). Finally, rhodolith beds may also function as a bank for microscopic algal stages, thus representing a reservoir for several macroalgae (Hoffmann & Santelices, 1991; Fredericq *et al.*, 2019).

(2) Sponges

As described above for seaweeds, rhodolith beds host highly diverse sponge assemblages (Fig. 2C, D), including boring (infauna) and epifaunal species, across the world's oceans (Solórzano & Urgorri, 1991; Aguilar *et al.*, 2009; Ávila *et al.*, 2013; Pereira-Filho *et al.*, 2015; Longo *et al.*, 2020). For example, in two rhodolith beds off the Island of Ustica (Southern Tyrrhenian Sea), Longo *et al.* (2020) documented the presence of 25 sponge taxa. While mostly belonging to the class of Demospongiae, these sponges were characterised by a variety of growth forms (i.e. massive, encrusting, insinuating and excavating). Similarly, Solórzano & Urgorri (1991) reported 39 species of sponges associated with a single Galician maerl bed, while Santín *et al.* (2024) documented eight sponge species in rhodolith beds within the Madeira archipelago, including a new species (*Hemimycale funchalensis*). Recent surveys in mesophotic rhodolith beds off the Amazon

River mouth in Brazil have recorded highly diverse sponge assemblages, including new species (Moura *et al.*, 2016; Sandes *et al.*, 2021). Such “sponge gardens” over rhodolith beds in north Brazil, in turn, serve as habitat for a diverse fish assemblage and may act as stepping stones for reef biota between the Caribbean and Brazilian Provinces (Rocha, Rosa & Feitoza, 2000; Rocha, 2003).

When at high density and biomass (Lopez-Acosta *et al.*, 2022), sponges can form structurally complex structures that provide nursery and rearing areas for other organisms, often augmenting the diversity of invertebrate and fish assemblages (Fig. 3B) (Rocha *et al.*, 2000; Kazanidis *et al.*, 2016; Hawkes *et al.*, 2019; Campanino *et al.*, 2023). Sponges are known to be secondary habitat-formers in mangrove forest facilitation cascades (Gribben *et al.*, 2019). For example, the species richness and abundance of fish and mobile invertebrate assemblages in Caribbean mangrove forests was positively correlated with the abundance of sponges growing on prop-roots (MacDonald & Weis, 2013; Stewart *et al.*, 2022). Sponges with a tubular, convoluted or massive growing form could provide shelter from predation for smaller fish and, at the same time, represent a food source for other fish or invertebrate species. In oligotrophic systems, sponges can also exert positive effects on primary producers

through nutrient supply (Archer *et al.*, 2021) – providing a growth-promoting feedback loop to the rhodoliths and other associated algae.

In addition, sponge–rhodolith assemblages play an important role in substrate construction and stabilisation, due to overgrowth and binding (Ávila *et al.*, 2013; Pereira-Filho *et al.*, 2015). For example, on the continental shelf of the Fernando de Noronha Archipelago, rhodolith mounds formed by the sand tilefish can then be bound by the sponges *Xestospongia muta* and *Agelas clathrodes*, stabilising the rhodolith accumulation and facilitating the establishment of coral colonies (Pereira *et al.*, 2015; Pereira-Filho *et al.*, 2015). Within this context, rhodoliths directly enable at least two distinct cascades – one initiated by the sand tilefish and the other by sponges, with cascading effects across trophic levels.

(3) Bivalves

Bivalves are common inhabitants of rhodolith beds (Fig. 2E, F), both as epifauna and endofauna (Kamenos *et al.*, 2004c; Hall-Spencer *et al.*, 2003; Steller *et al.*, 2003) and can also act as secondary facilitators (Fig. 3C). Several aggregating bivalve species, such as flat oysters or mytilid species, may reach densities high enough to enhance further the three-dimensional complexity of rhodolith beds (Fig. 3C), promoting local biodiversity through the provisioning of refugia against predators or by trapping organic matter (Norling & Kautsky, 2008; Bishop *et al.*, 2012; Gribben *et al.*, 2019), as also shown for oysters growing on mangrove prop-roots (Bishop *et al.*, 2012; Stewart *et al.*, 2022). Rhodolith beds are able to support bivalve species in areas where they are otherwise locally rare, such as enhanced abundance of *Gregar-iella semigranata* and *Leisonelus aristatu* in the oligotrophic waters of the Madeira Archipelago (AMACO, 2022). Similarly, in the northeast Atlantic, *Limaria hians*, a bivalve species of high conservation value, uses rhodolith beds for nesting, again stabilising the substratum and facilitating further colonisation by sessile species (Hall-Spencer & Moore, 2000). Further species diversity is facilitated in rhodolith beds *via* support of larger bivalve species such as fan mussels, horse mussels and scallops (Steller & Caceres-Martinez, 2009; Kersting & García-March, 2017), which host additional distinctive species assemblages on their shells, including macroalgae, hydroids, sponges, molluscs, bryozoans, and crustaceans (Corriero & Pronzato, 1987; Cummings *et al.*, 1998; Giacobbe, 2002; Cerrano *et al.*, 2001; Farren & Donovan, 2007) (Fig. 3C).

VI. RESEARCH NEEDS AND FUTURE DIRECTIONS IN THE CONTEXT OF FACILITATION CASCADES

Below, we provide a synthetic account of strategic research required to fill in gaps in our understanding of the role of

positive species interactions in shaping the biodiversity and functioning of rhodolith beds.

(1) Experimental evaluation of the role of rhodoliths as basal facilitators

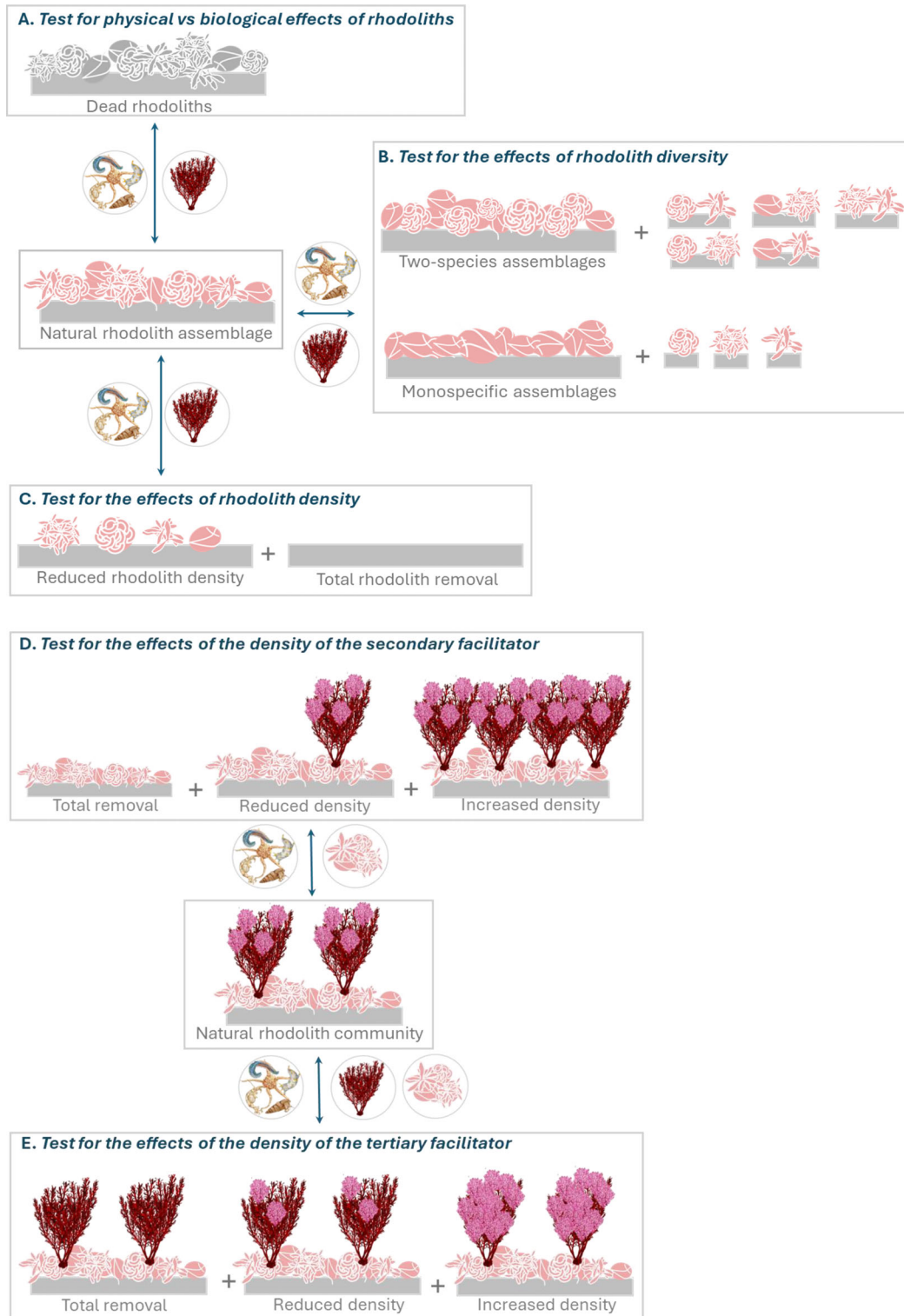
Due to the hierarchical nature of the organisation of rhodolith communities, assessing the effect of the foundation species (i.e. the rhodoliths) on species at upper levels of the facilitation cascade should be considered a priority. To the best of our knowledge, no study has assessed whether the facilitative effects of rhodoliths are biological and/or physical (Fig. 4A). Only one study (Otero-Ferrer *et al.*, 2019) has experimentally investigated the effects of variations in the size and heterogeneity of nodules on macrofaunal assemblages and the role of rhodolith species composition in regulating the structure of associated invertebrate and macroalgal communities (Fig. 4B) remains virtually unexplored. The removal of rhodolith nodules from some areas, or transplanting them onto bare sediments, to generate different densities (Fig. 4C), would allow assessments of their net effect on the diversity of the associated community and to identify which species are reliant on their presence (i.e. obligate associations). In addition, the experimental manipulation of rhodolith species richness and abundance could provide insights into the ecological mechanisms sustaining the functioning of the associated community (Tilman, 1999; Lehman & Tilman, 2000; Loreau, 2000). According to the Biodiversity–Ecosystem Function theory (Hooper *et al.*, 2012; Naeem, Duffy & Zavaleta, 2012), the number and relative abundance of facilitated species would be expected to increase with the number of rhodolith species or morphologies composing a bed. Due to their mobile nature and reduced size, the experimental manipulation of rhodoliths is more feasible than those of other coastal foundation species that are generally large sized, sessile and, in some cases, have developed root systems (e.g. seagrass and mangroves). Rhodolith manipulation, among areas within beds or between beds and nearby bare habitats, over relatively small areas would be sufficient to assess the effects on associated flora and fauna without causing significant damage to beds. On the other hand, experimental manipulations by scuba divers could be limited by depth in some regions. For example, in the Mediterranean Sea, many rhodolith beds occur at depths greater than 50 m (Basso *et al.*, 2017; Illa-López *et al.*, 2023). However, rapid technological advancements in underwater remote operating vehicles may provide novel opportunities for experimental research on deeper habitats in the near future.

(2) The length of facilitation cascades and negative feedbacks on rhodoliths

We currently have no empirical data on the number of levels within rhodolith bed facilitation cascades. Experimental removal approaches will be needed to assess the role of secondary facilitators on upper levels of the facilitation cascade

(Thomsen *et al.*, 2016). The analysis of features of secondary and upper-level facilitators, such as their abundance (Fig. 4D, E) and the degree of morphological complexity,

could enhance our understanding of their roles within the cascade (Thomsen *et al.*, 2022). In particular, assessing whether there are density thresholds for the formation and



(Figure 4 legend continues on next page.)

maintenance of facilitation cascades should be a priority (Fig. 4D, E). Secondary facilitators may be ineffective at sustaining other species when at very low abundance, or may trigger negative feedbacks on the foundation species (i.e. exert negative effects on rhodoliths) when at very high abundance. For example, in deep waters, high covers of facilitated macroalgae will likely have a detrimental effect on rhodolith nodules through shading and competition for light. In shallow waters, high epiphyte loads may favour the persistence of rhodoliths by mitigating excessive light intensity (Ravaglioli *et al.*, 2021), but also increase drag forces due to wave action, potentially leading to onshore stranding (J. Grall, personal observations). Also, the permanent stabilisation of rhodolith beds by bivalves or sponges may be detrimental to underlying rhodolith nodules (Ávila *et al.*, 2013), with anecdotal concerns that flame shell *Limaria hians* nests are beginning to smother rhodolith habitat in northwest Scotland. Similarly, the invasive gastropod *Crepidula fornicata* has been reported to overgrow living rhodoliths, leading to smothering and ultimately their death (Grall & Hall-Spencer, 2003). Likewise, boring species of sponges and bivalves, while generating further microhabitats, can cause nodule fragmentation when at high densities. Thus, as shown for other systems, density-dependent switches from facilitation to parasitism or competitive exclusion can be expected (Bulleri *et al.*, 2011; Schob *et al.*, 2014), resulting in a low density of living rhodolith thalli and a decline or loss of the rhodolith bed. Nonetheless, potential negative effects of facilitated species on rhodoliths remain unexplored.

(3) Variability across environmental gradients

The factors that shape rhodolith composition may also directly influence the associated species community and the interactions among species within and across cascade levels. The presence of marked gradients of environmental conditions or resource availability could open avenues for research framed within well-established theoretical frameworks, such as the Stress Gradient Hypothesis (SGH; Bertness & Callaway, 1994). Assessing changes in the sign and strength

of interactions among species within a facilitation cascade under varying environmental conditions, such as wave exposure, light intensity, water chemistry, sedimentation rate or consumer pressure, would provide insights into the facilitation pathways underpinning the hierarchical organisation of rhodolith beds. According to the SGH, the intensity of positive effects of rhodoliths on sessile and mobile species through the provisioning of more stable substrates or the trapping of organic matter are predicted to decrease when moving from wave-swept to sheltered environments. Likewise, the relevance of rhodoliths as refuges against predation would weaken in areas where consumer pressure is low. For example, positive effects of rhodoliths on the seagrass *Thalassia testudinum* should decrease in intensity and eventually shift to negative (i.e. competition), in areas where the abundance of sea turtles and, hence, grazing pressure, is low.

(4) The temporal stability of cascades

Facilitation cascade stability is predicted to decrease with growing numbers of facilitation links and increase with functional redundancy (Yakovis & Artemieva, 2017; Gribben *et al.*, 2019). Due to the high numbers of species supported, functional redundancy within and across facilitation cascade levels could be expected to be high, enhancing its stability. For example, according to the biodiversity–stability theory (Tilman & Downing, 1994), rhodolith beds composed of multiple rhodolith species could absorb external perturbations more efficiently than monospecific beds. Thus, facilitation cascades in multispecies-rhodolith beds might be more stable than those found in seagrass beds, mangroves and kelp forests, which are often formed by single species. This potential resilience (Allison, 2004) should be addressed experimentally, testing multi-stressor scenarios encompassing ocean warming, acidification and their interactions with regional and local stressors (Gissi *et al.*, 2021). Assessing the temporal stability of rhodolith facilitation cascades requires, however, a mix of experimental and observational research. On the one hand, the selective removal/addition of species within the same facilitation level would allow evaluation of the role

(Figure legend continued from previous page.)

Fig. 4. (A–C) Schematic representation of potential experiments assessing the effects of rhodolith bed traits on associated species. (A) A comparison between live and dead rhodoliths. (B) Comparisons among rhodolith assemblages composed of a different number of species or morphospecies (e.g. natural assemblage *versus* two-species assemblages *versus* monospecific assemblages); in this example, natural rhodolith assemblages are composed of four species across a branching gradient. (C) Comparisons between assemblages composed of the same rhodolith species, but differing in their density (natural *versus* reduced *versus* total removal). (D, E) Schematic representation of potential experiments assessing the effects of upper-level facilitators on rhodoliths and associated assemblages. (D) Different densities of the secondary facilitator (natural *versus* total removal *versus* reduced density *versus* increased density). (E) Different densities of the tertiary facilitator (natural *versus* total removal *versus* reduced density *versus* increased density); in these examples, the secondary and tertiary facilitators are a canopy-forming and an epiphytic macroalga, respectively. For each of the illustrated experimental tests, double-headed arrows indicate comparisons with the natural controls and lateral close-ups represent the response variable/s (the rhodoliths, the canopy-forming macroalga and the invertebrate assemblage) that can be potentially taken into account to assess positive/negative effects of each of the three levels included in the cascade. The illustrated densities of the basal, secondary and tertiary facilitators are simple examples of possible manipulations which can be extended to the upper levels of a cascade. A detailed account of experimental designs for disentangling the effects of species richness from those of species identity and density can be found in Benedetti-Cecchi (2004).

of functional redundancy in sustaining stability. This seems particularly relevant in rhodolith beds since they often host multiple species potentially acting as secondary or tertiary facilitators (Fig. 3). On the other hand, temporal series of data encompassing generation turn-overs of involved species are necessary to calculate key metrics, such as the coefficient of variation and the synchrony of species fluctuations (Lehman & Tilman, 2000; Loreau & de Mazancourt, 2008).

(5) Rhodolith beds as climate refugia

While there has been considerable interest in assessing the vulnerability of rhodolith beds to climate changes (Noiset *et al.*, 2013; Rindi *et al.*, 2019; Koerich *et al.*, 2021; Costa *et al.*, 2023), field and laboratory experiments remain needed to evaluate their role as climatic refugia (Voerman *et al.*, 2022). Depth has been hypothesised to provide refugia against warming (Graham *et al.*, 2007; Liberman *et al.*, 2022) and the provision of consolidated substrata might extend the bathymetric distribution of hard-bottom dwelling species to depths less influenced by extreme warming events. In addition, rhodoliths support high diversity and biomass of epiphytic macroalgae, which can modify seawater chemistry through their biological activities. Epiphytic macroalgae, due to photosynthesis, could locally increase seawater pH (Burdett *et al.*, 2018) and provide spatial and temporal refugia for benthic calcifying species, including rhodoliths facing ocean acidification. In future envisaged high- $p\text{CO}_2$ conditions, epiphytic macroalgae on rhodoliths are expected to benefit from higher CO_2 concentrations and primary production to be enhanced in future rhodolith beds (Martin & Hall-Spencer, 2017). Thus, these habitats could provide suitable environmental (chemical) conditions for their associated calcifying species in future high- $p\text{CO}_2$ conditions. The wide distribution of rhodolith beds from tropical to polar latitudes (Fragkopoulou *et al.*, 2021), is broader than that of many other marine and coastal foundation species. Thus, assessing how the diversity of rhodolith species or morphospecies and that of associated habitat-formers varies across such broad latitudinal gradients would provide insights into their potential to act as climate refugia.

VII. CONCLUSIONS

- (1) Despite being distributed throughout the photic zone across the global oceans (from tropical to polar regions), and being recognised as biodiversity hotspots, rhodolith beds are yet to be fully appreciated as key marine habitats at the level of seagrass meadows, mangrove and macroalgal forests or coral reefs.
- (2) Here, we show that rhodoliths can function as foundation species of multi-level facilitation cascades, and hence are fundamental for the persistence of hierarchically structured communities within coastal oceans around the world.

- (3) Research priorities should now seek to understand better the processes underpinning these community assemblies, the pathways of ecological facilitation and the effects of human and environmental perturbations.
- (4) Addressing these priorities will allow the development of evidence-based policy decisions and elevate rhodolith beds within marine conservation and coastal management strategies.

VIII. ACKNOWLEDGEMENTS

The preparation of this review was supported by Euro-Marine through the Foresight Workshop RHODOCARB (<https://euromarinenetwork.eu/activities/rhodocarb/>). F.B. was supported by the European Union's Horizon 2020 research and innovation programme under grant agreement No. 869300 (FutureMARES). N.S. acknowledges funding by Portuguese National Funds from FCT-Fundação para a Ciência e a Tecnologia through an Assistant researcher grant (<https://doi.org/10.54499/2020.01282.CEECIND/CP1597/CT0003>), by a FCT/CAPES project (2019.00067.CBM), and through FCT-funded projects UIDB/04326/2020, UIDP/04326/2020, and LA/P/0101/2020. F.T. and V.P. acknowledge funding by the Spanish Science and Innovation Ministry (project "POPCORN", PID2021-124257OB-I00). F.R. acknowledges NBFC support to the Stazione Zoologica Anton Dohrn (SZN), funded under the National Recovery and Resilience Plan (NRRP) Mission 4, Component 2, Investment 1.4 -Call for tender No. 3138 of 16 December 2021, rectified by Decree n 3175 of 18 December 2021 of Italian Ministry of University and Research funded by the European Union -Next Generation EU. Project code, CN00000033 Concession Decree No.1034, of 17 June 2022 adopted by the Italian Ministry of University and Research, CUP: C63C22000520001 Project title "National Biodiversity future Center-NBFC". M.N. was supported by a PhD fellowship funded by the SZN (Open University – SZN PhD Program). R.B.F.-F. is grateful for a research productivity scholarship provided by CNPq (#309651/2021-2). H.L.B. and N.A.K. acknowledge funding from the Kempe Foundation (JCSMK24-0033). Finally, we sincerely thank two anonymous reviewers for providing insightful comments and constructive criticism.

IX. REFERENCES

- AGUILAR, R., PASTOR, X., TORRIENTE, A. & GARCIA, S. (2009). Deep-sea coralligenous beds observed with ROV on four seamounts in the western Mediterranean. In *Proceedings of the 1st Mediterranean Symposium on the Conservation of the Coralligenous and Others Calcareous Bio-Concretions*, pp. 147–149. UNEP-MAP-RAC/SPA. CAR/ASP Publishing, Tabarka, Tunisia.
- ALLISON, G. (2004). The influence of species diversity and stress intensity on community resistance and resilience. *Ecological Monographs* **74**, 117–134.
- ALTIERI, A. H., SILLIMAN, B. R. & BERTNESS, M. D. (2007). Hierarchical organization via a facilitation cascade in intertidal cordgrass bed communities. *American Naturalist* **169**, 195–206.

- ALTIERI, A. H. & WITMAN, J. D. (2014). Modular mobile foundation species as reservoirs of biodiversity. *Ecosphere* **5**, 1–11.
- AMACO (2022). Madeira Maerl Mapping & Conservation Project. Final Report. Life4Best 2020-M-44.
- AMADO, G. M., MANEVELDT, G., MANSO, R. C. C., MARINS-ROSA, B. V., PACHECO, M. R. & GUIMARAES, S. (2007). Structure of rhodolith beds from 4 to 55 meters deep along the southern coast of Espírito Santo state, Brazil. *Ciencias Marinas* **33**, 399–410.
- AMADO-FILHO, G. M. & PEREIRA-FILHO, G. H. (2012). Rhodolith beds in Brazil: a new potential habitat for marine bioprospection. *Brazilian Journal of Pharmacognosy* **22**, 782–788.
- ANDERSON, A. B., PINHEIRO, H. T., BATISTA, M. B., FRANCINI, R. B., GOMES, L. E. O., BERNARDINO, A. F., HORTA, P. & JOYEUX, J. C. (2023). Biogeographic patterns of marine fishes associated with rhodolith beds in the southwestern Atlantic reveal an ecotone of biodiversity. *Biodiversity and Conservation* **32**, 821–837.
- ANGELINI, C., ALTIERI, A. H., SILLIMAN, B. R. & BERTNESS, M. D. (2011). Interactions among foundation species and their consequences for community organization, biodiversity, and conservation. *Bioscience* **61**, 782–789.
- ANGELINI, C. & SILLIMAN, B. R. (2014). Secondary foundation species as drivers of trophic and functional diversity: evidence from a tree epiphyte system. *Ecology* **95**, 185–196.
- APE, F., GRISTINA, M., CHEMELLO, R., SARÀ, G. & MIRTO, S. (2018). Meiofauna associated with vermetid reefs: the role of macroalgae in increasing habitat size and complexity. *Coral Reefs* **37**, 875–889.
- ARCHER, S. K., ENGLISH, P. A., CAMPANINO, F. M. & LAYMAN, C. A. (2021). Sponges facilitate primary producers in a Bahamas seagrass system. *Marine Biology* **168**, 162.
- ÁVILA, E., RIOSMENA-RODRIGUEZ, R. & HINOJOSA-ARANGO, G. (2013). Sponge-rhodolith interactions in a subtropical estuarine system. *Helgoland Marine Research* **67**, 349–357.
- BAHIA, R. G., ABRANTES, D. P., BRASILEIRO, P. S., PEREIRA, G. H. & AMADO, G. M. (2010). Rhodolith bed structure along a depth gradient on the northern coast of Bahia state, Brazil. *Brazilian Journal of Oceanography* **58**, 323–337.
- BARBERÁ, C., MORANTA, J., ORDINES, F., RAMÓN, M., DE MESA, A., DÍAZ-VALDÉS, M., GRAU, A. M. & MASSUTÍ, E. (2012). Biodiversity and habitat mapping of Menorca Channel (western Mediterranean): implications for conservation. *Biodiversity and Conservation* **21**, 701–728.
- BASSO, D. (1998). Deep rhodolith distribution in the Pontian Islands, Italy: a model for the paleoecology of a temperate sea. *Palaeogeography Palaeoclimatology Palaeoecology* **137**, 173–187.
- BASSO, D., BABBINI, L., KALEB, S., BRACCHI, V. A. & FALACE, A. (2016). Monitoring deep Mediterranean rhodolith beds. *Aquatic Conservation-Marine and Freshwater Ecosystems* **26**, 549–561.
- BASSO, D., BABBINI, L., RAMOS-ESPLÁ, A. A. & SALOMIDI, M. (2017). Mediterranean Rhodolith Beds. In *Rhodolith/Maerl Beds: A Global Perspective*. Coastal Research Library (Volume 15, eds R. RIOSMENA-RODRIGUEZ, W. NELSON and J. AGUIRRE). Springer, Cham. https://doi.org/10.1007/978-3-319-29315-8_11.
- BASSO, D., NALIN, R. & NELSON, C. S. (2009). Shallow-water *Sporolithon* rhodoliths from North Island (New Zealand). *PALAIOS* **24**, 92–103.
- BASTOS, A. C., MOURA, R. L., AMADO, G. M., D'AGOSTINI, D. P., SECCHIN, N. A., FRANCINI, R. B., GUTH, A. Z., SUMIDA, P. Y. G., MAHIQUES, M. M. & THOMPSON, F. L. (2013). Buracas: novel and unusual sinkhole-like features in the Abrolhos Bank. *Continental Shelf Research* **70**, 118–125.
- BENEDETTI-CECCHI, L. (2004). Increasing accuracy of causal inference in experimental analyses of biodiversity. *Functional Ecology* **18**, 761–768.
- BERTNESS, M. D. & CALLAWAY, R. (1994). Positive interactions in communities. *Trends in Ecology & Evolution* **9**, 191–193.
- BISHOP, M. J., BYERS, J. E., MARCEK, B. J. & GRIBBEN, P. E. (2012). Density-dependent facilitation cascades determine epifaunal community structure in temperate Australian mangroves. *Ecology* **93**, 1388–1401.
- BOSELLINI, A. & GINSBURG, R. N. (1971). Form and internal structure of recent algal nodules (rhodolites) from Bermuda. *Journal of Geology* **79**, 669–682.
- BOSENCE, D. W. J. (1976). Ecological studies on two carbonate sediment producing coralline algae from western Ireland. *Palaeontology* **19**, 365–395.
- BOSENCE, D. W. J. (1983). The occurrence and ecology of recent rhodoliths – a review. In *Coated Grains* (ed. T. M. PERYT), pp. 217–224. Springer Verlag, Berlin.
- BOYE, A., THIEBAUT, E., GRALL, J., LEGENDRE, P., BROUDIN, C., HOUBIN, C., LE GARREC, V., MAGUER, M., DROUAL, G. & GAUTHIER, O. (2019). Trait-based approach to monitoring marine benthic data along 500 km of coastline. *Diversity and Distributions* **25**, 1879–1896.
- BRACCHI, V., ANGELETTI, L., MARCHESE, F., TAVIANI, M., CARDONE, F., IRKA, H., GRANDE, V., PRAMPOLINI, M., CARAGNANO, A., CORSELLI, C. & BASSO, D. (2019). A resilient deep-water rhodolith bed off the Egadi archipelago (Mediterranean Sea) and its actuopalaeontological significance. *Alpine and Mediterranean Quaternary* **32**, 131–150.
- BRACCHI, V. A., CARONNI, S., MERONI, A. N., BURGUETT, E. G., ATZORI, F., CADONI, N., MARCHESE, F. & BASSO, D. (2022). Morphostructural characterization of the heterogeneous rhodolith bed at the marine protected area “capo carbonara” (Italy) and hydrodynamics. *Diversity-Basel* **14**, 51.
- BRACCHI, M. E. S. (2018). When one foundation species supports another: tubeworms facilitate an extensive kelp bed in a soft-sediment habitat. *Ecosphere* **9**, e02429.
- BRAGA-HENRIQUES, A., BUHL-MORTENSEN, P., TOKAT, E., MARTINS, A., SILVA, T., JAKOBSEN, J., CANNING-CLODE, J., JAKOBSEN, K., DELGADO, J., VOIRAND, T. & BISCOITTO, M. (2022). Benthic community zonation from mesophotic to deep sea: description of first deep-water kelp forest and coral gardens in the Madeira archipelago (central NE Atlantic). *Frontiers in Marine Science* **9**, 973364.
- BRASILEIRO, P. S., PEREIRA, G. H., BAHIA, R. G., ABRANTES, D. P., GUIMARAES, S., MOURA, R. L., FRANCINI, R. B., BASTOS, A. C. & AMADO, G. M. (2016). Macroalgal composition and community structure of the largest rhodolith beds in the world. *Marine Biodiversity* **46**, 407–420.
- BROOKER, R. W., MAESTRE, F. T., CALLAWAY, R. M., LORTIE, C. L., CAVIERES, L. A., KUNSTLER, G., LIANCOURT, P., TIELBORGER, K., TRAVIS, J. M. J., ANTHELME, F., ARMAS, C., COLL, L., CORCKET, E., DELZON, S., FOREY, E., ET AL. (2008). Facilitation in plant communities: the past, the present, and the future. *Journal of Ecology* **96**, 18–34.
- BRUNO, J. F. & BERTNESS, M. D. (2001). Habitat modification and facilitation in benthic marine communities. In *Marine Community Ecology* (eds M. D. BERTNESS, S. GAINES and M. E. HAY), pp. 201–216. Sinauer, Sunderland, MA.
- BRUNO, J. F., STACHOWICZ, J. J. & BERTNESS, M. D. (2003). Inclusion of facilitation into ecological theory. *Trends in Ecology & Evolution* **18**, 119–125.
- BULLERI, F. (2009). Facilitation research in marine systems: state of the art, emerging patterns and insights for future developments. *Journal of Ecology* **97**, 1121–1130.
- BULLERI, F. & AIROLDI, L. (2005). Artificial marine structures facilitate the spread of a nonindigenous green alga, *Codium fragile* ssp. *tomentosoides*, in the North Adriatic Sea. *Journal of Applied Ecology* **42**, 1063–1072.
- BULLERI, F., BRUNO, J. F., SILLIMAN, B. R. & STACHOWICZ, J. J. (2016). Facilitation and the niche: implications for coexistence, range shifts and ecosystem functioning. *Functional Ecology* **30**, 70–78.
- BULLERI, F., CRISTAUDDO, C., ALESTRA, T. & BENEDETTI-CECCHI, L. (2011). Crossing gradients of consumer pressure and physical stress on shallow rocky reefs: a test of the stress-gradient hypothesis. *Journal of Ecology* **99**, 335–344.
- BURDETT, H. L. (2017). Exchange dynamics reveal significant accumulation of dimethylated sulfur by mediterranean benthic communities. *Frontiers in Marine Science* **4**, 431.
- BURDETT, H. L., HATTON, A. D. & KAMENOS, N. A. (2015). Coralline algae as a globally significant pool of marine dimethylated sulfur. *Global Biogeochemical Cycles* **29**, 1845–1853.
- BURDETT, H. L., PERNA, G., MCKAY, L., BROOMHEAD, G. & KAMENOS, N. A. (2018). Community-level sensitivity of a calcifying ecosystem to acute in situ CO₂ enrichment. *Marine Ecology Progress Series* **587**, 73–80.
- CAMPANINO, F. M., ENGLISH, P. A., LAYMAN, C. A. & ARCHER, S. K. (2023). Sponge presence increases the diversity and abundance of fish and invertebrates in a subtropical seagrass bed. *Estuaries and Coasts* **46**, 1009–1020.
- CARONNI, S., BRACCHI, V. A., ATZORI, F., CITTERIO, S., CADONI, N., GENTILI, R., MONTAGNANI, C., QUAGLINI, L. A. & BASSO, D. (2023). *Caulerpa cylindracea* spread on deep rhodolith beds can be influenced by the morphostructural composition of the bed. *Diversity-Basel* **15**, 349.
- CARRO, B., LOPEZ, L., PEÑA, V., BÁRBARA, I. & BARREIRO, R. (2014). DNA barcoding allows the accurate assessment of European maerl diversity: a proof-of-concept study. *Phytotaxa* **190**, 176–189.
- CARVALHO, V. F., ASSIS, J., SERRAO, E. A., NUNES, J. M., ANDERSON, A. B., BATISTA, M. B., BARUFI, J. B., SILVA, J., PEREIRA, S. M. B. & HORTA, P. A. (2020). Environmental drivers of rhodolith beds and epiphytes community along the South Western Atlantic coast. *Marine Environmental Research* **154**, 104827.
- CERRANO, C., PUCE, S., CHIANTORE, M., BAVESTRELLO, G. & CATTANEO-VIETTI, R. (2001). The influence of the epizoic hydroid *Hydractinia angusta* on the recruitment of the Antarctic scallop *Adamussium colbecki*. *Polar Biology* **24**, 577–581.
- CHAVES, L. T. C., PEREIRA, P. H. C. & FEITOSA, J. L. L. (2013). Coral reef fish association with macroalgal beds on a tropical reef system in North-eastern Brazil. *Marine and Freshwater Research* **64**, 1101–1111.
- CORNWALL, C. E., HARVEY, B. P., COMEAU, S., CORNWALL, D. L., HALL-SPENCER, J. M., PEÑA, V., WADA, S. & PORZIO, L. (2022). Understanding coralline algal responses to ocean acidification: meta-analysis and synthesis. *Global Change Biology* **28**, 362–374.
- CORRIERO, G. & PRONZATO, R. (1987). Epibiotic sponges on the bivalve *Pinna nobilis*. *Marine Ecology Progress Series* **35**, 75–82.
- COSTA, D. D., DOLBETH, M., CHRISTOFFERSEN, M. L., ZUNIGA-UPEGUI, P. T., VENANCIO, M. & DE LUCENA, R. F. P. (2023). An overview of rhodoliths: ecological importance and conservation emergency. *Life-Basel* **13**, 1556.

- CRAIN, C. M. & BERTNESS, M. D. (2006). Ecosystem engineering across environmental gradients: implications for conservation and management. *Bioscience* **56**, 211–218.
- CUMMINGS, V. J., THRUSH, S. F., HEWITT, J. E. & TURNER, S. J. (1998). The influence of the pinnid bivalve *Atrina zelandica* (Gray) on benthic macroinvertebrate communities in soft-sediment habitats. *Journal of Experimental Marine Biology and Ecology* **228**, 227–240.
- DAYTON, P. K. (1972). Toward an understanding of community resilience and the potential effects of enrichments to the benthos at McMurdo Sound, Antarctica. In *Proceedings of the Colloquium on Conservation Problems in Antarctica* (ed. B. C. PARKER), pp. 81–95. Allen Press, Lawrence, KS.
- DE QUEIROZ, E. V., ARAUJO, P. V. D., HAMMILL, E. & DO AMARAL, R. F. (2016). Morphological characteristics of rhodolith and correlations with associated sediment in a sandstone reef: Northeast Brazil. *Regional Studies in Marine Science* **8**, 133–140.
- DEL RIO, J., RAMOS, D. A., SANCHEZ-TOCINO, L., PEÑAS, J. & BRAGA, J. C. (2022). The Punta de la Mona rhodolith bed: shallow-water Mediterranean rhodoliths (Almuncar, Granada, southern Spain). *Frontiers in Earth Science* **10**, 884685.
- EDGAR, G. J. & ROBERTSON, A. I. (1992). The influence of seagrass structure on the distribution and abundance of mobile epifauna: pattern and processes in a Western Australian *Amphibolis* bed. *Journal of Experimental Marine Biology and Ecology* **160**, 13–31.
- EL-KHALED, Y. C., DARAGHMEH, N., TILSTRA, A., ROTH, F., HUETTEL, M., ROSSBACH, F. I., CASOLI, E., KOESTER, A., BECK, M., MEYER, R., PLEWKA, J., SCHMIDT, N., WINKELGRUND, L., MERK, B. & WILD, C. (2022). Fleshy red algae mats act as temporary reservoirs for sessile invertebrate biodiversity. *Communications Biology* **5**, 579.
- ELLISON, A. M. (2019). Foundation species, non-trophic interactions, and the value of being common. *iScience* **29**, 254–268.
- FARREN, H. M. & DONOVAN, D. A. (2007). Effects of sponge and barnacle encrustation on survival of the scallop *Chlamys hastata*. *Hydrobiologia* **592**, 225–234.
- FERNANDEZ, C. (2011). The retreat of large brown seaweeds on the north coast of Spain: the case of *Saccorhiza polyschides*. *European Journal of Phycology* **46**, 352–360.
- FORETICH, M. A., PARIS, C. B., GROSELL, M., STIEGLITZ, J. D. & BENETTI, D. D. (2017). Dimethyl sulfide is a chemical attractant for reef fish larvae. *Scientific Reports* **7**, 2498.
- FOSTER, M. S., AMADO FILHO, G. M., KAMENOS, N. A., RIOSMENA-RODRÍGUEZ, R. & STELLER, D. L. (2013). Rhodoliths and rhodolith beds research and discoveries: the revolution of science through scuba. In *Smithsonian Contributions to the Marine Sciences* (Volume 39, eds M. A. LANG, R. L. MARINELLI, S. J. ROBERTS and P. R. TAYLOR), pp. 143–155. Smithsonian Institution Scholarly Press, Washington, DC.
- FOSTER, M. S., RIOSMENA-RODRÍGUEZ, R., STELLER, D. L. & WOELKERLING, W. J. (1997). Living rhodolith beds in the Gulf of California and their implications. Pliocene carbonates and related facies flanking the Gulf of California, Baja California, Mexico. *Geological Society of America* **318**, 127–139.
- FRAGKOPOULOU, E., SERRAO, E. A., HORTA, P. A., KOERICH, G. & ASSIS, J. (2021). Bottom trawling threatens future climate refugia of rhodoliths globally. *Frontiers in Marine Science* **7**, 1246.
- FRANCINI, R. B., ASP, N. E., SIEGLE, E., HOCEVAR, J., LOWYCK, K., D'AVILA, N., VASCONCELOS, A. A., BAITTELO, R., REZENDE, C. E., OMACHI, C. Y., THOMPSON, C. C. & THOMPSON, F. L. (2018). Perspectives on the Great Amazon Reef: extension, biodiversity, and threats. *Frontiers in Marine Science* **5**, 142.
- FREDERICQ, S., KRAYESKY-SELF, S., SAUVAGE, T., RICHARDS, J., KITTLE, R., ARAKAKI, N., HICKERSON, E. & SCHMIDT, W. E. (2019). The critical importance of rhodoliths in the life cycle completion of both macro- and microalgae, and as holobionts for the establishment and maintenance of marine biodiversity. *Frontiers in Marine Science* **5**, 502.
- FULTON, C. J., BERKSTROM, C. T., WILSON, S. K., ABESAMIS, R. A., BRADLEY, M., AKERLUND, C., BARRETT, L. T., BUCOL, A. A., CHACIN, D. H., CHONG-SENG, K. M., COKER, D. J., DEPCZYNSKI, M., EGGERTSEN, L., EGGERTSEN, M., ELLIS, D., ET AL. (2020). Macroalgal meadow habitats support fish and fisheries in diverse tropical seascapes. *Fish and Fisheries* **21**, 700–717.
- GABARA, S. S. (2020). Trophic structure and potential carbon and nitrogen flow of a rhodolith bed at Santa Catalina Island inferred from stable isotopes. *Marine Biology* **167**, 30.
- GAGNON, P., MATHESON, K. & STAPLETON, M. (2012). Variation in rhodolith morphology and biogenic potential of newly discovered rhodolith beds in Newfoundland and Labrador (Canada). *Botanica Marina* **55**, 85–99.
- GIACOBBE, S. (2002). Epibiontic mollusc communities on *Pinna nobilis* L. (Bivalvia, Mollusca). *Journal of Natural History* **36**, 1385–1396.
- GISSI, E., MANEA, E., MAZARIS, A. D., FRASCHETTI, S., ALMPANIDOU, V., BEVILACQUA, S., COLL, M., GUARNIERI, G., LLORET-LLORET, E., PASQUAL, M., PETZA, D., RILOV, G., SCHONWALD, M., STELZENMÜLLER, V. & KATSANEVAKIS, S. (2021). A review of the combined effects of climate change and other local human stressors on the marine environment. *Science of the Total Environment* **755**, 142564.
- GLYNN, P. W. (1974). Rolling stones amongst the Scleractinia: mobile coralloliths in the Gulf of Panama. *Proceeding 2nd International Coral Reef Symposium* **2**, 183–198.
- GRAHAM, M. H., KINLAN, B. P., DRUEHL, L. D., GARSKE, L. E. & BANKS, S. (2007). Deep-water kelp refugia as potential hotspots of tropical marine diversity and productivity. *Proceedings of the National Academy of Sciences of the United States of America* **104**, 16576–16580.
- GRALL, J. & HALL-SPENCER, J. M. (2003). Problems facing maerl conservation in Brittany. *Aquatic Conservation-Marine and Freshwater Ecosystems* **13**, S55–S64.
- GRALL, J., LE LOC'H, F., GUYONNET, B. & RIERA, P. (2006). Community structure and food web based on stable isotopes (delta N-15 and delta C-13) analysis of a North Eastern Atlantic maerl bed. *Journal of Experimental Marine Biology and Ecology* **338**, 1–15.
- GRIFFIN, P. E., ANGELINI, C., ALTIERI, A. H., BISHOP, M. J., THOMSEN, M. S. & BULLERI, F. (2019). Facilitation cascades in marine ecosystems: a synthesis and future directions. *Oceanography and Marine Biology: An Annual Review* **57**, 127–168.
- GRIFFIN, P. E., BYERS, J. E., CLEMENTS, M., MCKENZIE, L. A., STEINBERG, P. D. & WRIGHT, J. T. (2009). Behavioural interactions between ecosystem engineers control community species richness. *Ecology Letters* **12**, 1127–1136.
- GROSS, K. (2008). Positive interactions among competitors can produce species-rich communities. *Ecology Letters* **11**, 929–936.
- HALL-SPENCER, J. (1998). Conservation issues relating to maerl beds as habitats for molluscs. *Journal of Conchology, S.I.2*, 271–285.
- HALL-SPENCER, J. M. & ATKINSON, R. J. A. (1999). *Upogebia deltaura* (Crustacea: Thalassinidea) in Clyde Sea maerl beds, Scotland. *Journal of the Marine Biological Association of the United Kingdom* **79**, 871–880.
- HALL-SPENCER, J. M., GRALL, J., MOORE, P. G. & ATKINSON, R. J. A. (2003). Bivalve fishing and maerl-bed conservation in France and the UK - retrospect and prospect. *Aquatic Conservation-Marine and Freshwater Ecosystems* **13**, S33–S41.
- HALL-SPENCER, J. M. & MOORE, P. G. (2000). *Limaria hians* (Mollusca: Limacea): a neglected reef-forming keystone species. *Aquatic Conservation-Marine and Freshwater Ecosystems* **10**, 267–277.
- HARRIS, P. T., TSUJI, Y., MARSHALL, J. F., DAVIES, P. J., HONDA, N. & MATSUDA, H. (1996). Sand and rhodolith-gravel entrainment on the mid-to outer-shelf under a western boundary current: Fraser Island continental shelf, eastern Australia. *Marine Geology* **129**(3–4), 313–330.
- HAWKES, N., KORABIK, M., BEAZLEY, L., RAPP, H. T., XAVIER, J. R. & KENCHINGTON, E. (2019). Glass sponge grounds on the Scotian Shelf and their associated biodiversity. *Marine Ecology Progress Series* **622**, 109–231.
- HELIAS, M. & BUREL, T. (2023). Maerl-associated macroalgae in the bay of Brest (Brittany, France). *Marine Biodiversity* **53**, 14.
- HOFFMANN, A. J. & SANTELICES, B. (1991). Banks of algal microscopic forms – hypotheses on their functioning and comparisons with seeds banks. *Marine Ecology Progress Series* **79**, 185–194.
- HOFMANN, L. C., SCHOENROCK, K. & DE BEER, D. (2018). Arctic coralline algae elevate surface pH and carbonate in the dark. *Frontiers in Plant Science* **9**, 1416.
- HOOPER, D. U., ADAIR, E. C., CARDINALE, B. J., BYRNES, J. E. K., HUNGATE, B. A., MATULICH, K. L., GONZALEZ, A., DUFFY, J. E., GAMFELDT, L. & O'CONNOR, M. I. (2012). A global synthesis reveals biodiversity loss as a major driver of ecosystem change. *Nature* **486**, 105–U129.
- ILLA-LÓPEZ, L., CABRITO, A., DE JUAN, S., MAYNOU, F. & DEMESTRE, M. (2023). Distribution of rhodolith beds and their functional biodiversity characterisation using ROV images in the western Mediterranean Sea. *Science of the Total Environment* **905**, 167270.
- JAMES, D. W., FOSTER, M. S. & O'SULLIVAN, J. (2006). Bryoliths (Bryozoa) in the Gulf of California. *Pacific Science* **60**, 117–124.
- JAMES, K., MACREADIE, P. I., BURDETT, H. L., DAVIES, I. & KAMENOS, N. A. (2024). It's time to broaden what we consider a 'blue carbon ecosystem'. *Global Change Biology* **30**, e17261.
- JARDIM, V. L., GAUTHIER, O., TOUMI, C. & GRALL, J. (2022). Quantifying maerl (rhodolith) habitat complexity along an environmental gradient at regional scale in the Northeast Atlantic. *Marine Environmental Research* **181**, 105768.
- JOSHI, S., DUFFY, G. P. & BROWN, C. (2017). Critical bed shear stress and threshold of motion of maerl biogenic gravel. *Estuarine Coastal and Shelf Science* **194**, 128–142.
- KAMENOS, N. A., CALOSI, P. & MOORE, P. G. (2006). Substratum-mediated heart rate responses of an invertebrate to predation threat. *Animal Behaviour* **71**, 809–813.
- KAMENOS, N. A., MOORE, P. G. & HALL-SPENCER, J. M. (2004a). Attachment of the juvenile queen scallop (*Aequipecten opercularis* L.) to maerl in mesocosm conditions; juvenile habitat selection. *Journal of Experimental Marine Biology and Ecology* **306**, 139–155.
- KAMENOS, N. A., MOORE, P. G. & HALL-SPENCER, J. M. (2004b). Maerl grounds provide both refuge and high growth potential for juvenile queen scallops (*Aequipecten opercularis* L.). *Journal of Experimental Marine Biology and Ecology* **313**, 241–254.
- KAMENOS, N. A., MOORE, P. G. & HALL-SPENCER, J. M. (2004c). Nursery-area function of maerl grounds for juvenile queen scallops *Aequipecten opercularis* and other invertebrates. *Marine Ecology Progress Series* **274**, 183–189.

- KAMENOS, N. A., MOORE, P. G. & HALL-SPENCER, J. M. (2004d). Small-scale distribution of juvenile gadoids in shallow inshore waters; what role does maerl play? *ICES Journal of Marine Science* **61**(3), 422–429.
- KAZANIDIS, G., HENRY, L. A., ROBERTS, J. M. & WITTE, U. F. M. (2016). Biodiversity of *Spongosities coralliophaga* (Stephens, 1915) on coral rubble at two contrasting cold-water coral reef settings. *Coral Reefs* **35**, 193–208.
- KERSTING, D. K. & GARCÍA-MARCH, J. R. (2017). Long-term assessment of recruitment, early stages and population dynamics of the endangered Mediterranean fan mussel *Pinna nobilis* in the Columbretes Islands (NW Mediterranean). *Marine Environmental Research* **130**, 282–292.
- KOERICH, G., COSTA, G. B., SISSINI, M. N., ORTIZ, C. L., CANEVER, B. F., OLIVEIRA, W., TONKIN, J. D. & HORTA, P. A. (2021). Physiology, niche characteristics and extreme events: current and future habitat suitability of a rhodolith-forming species in the Southwestern Atlantic. *Marine Environmental Research* **169**, 105394.
- LAVENÈRE-WANDERLEY, A. A., EDVIN ASP, N., THOMPSON, F. L. & SIEGLE, E. (2021). Rhodolith mobility potential from seasonal and extreme waves. *Continental Shelf Research* **228**, 104527.
- LEEMANS, L., MARTINEZ, I., VAN DER HEIDE, T., VAN KATWIJK, M. M. & TUSSENBOEK, B. I. (2020). A mutualism between unattached coralline algae and seagrasses prevents overgrazing by sea turtles. *Ecosystems* **23**, 1631–1642.
- LEHMAN, C. L. & TILMAN, D. (2000). Biodiversity, stability, and productivity in competitive communities. *American Naturalist* **156**, 534–552.
- LIBERMAN, R., SHLESINGER, T., LOYA, Y. & BENAYAHU, Y. (2022). Soft coral reproductive phenology along a depth gradient: can “going deeper” provide a viable refuge? *Ecology* **103**, e3760.
- LITTLER, M. M., LITTLER, D. S. & HANISAK, M. D. (1991). Deep-water rhodolith distribution, productivity, and growth history at sites of formation and subsequent degradation. *Journal of Experimental Marine Biology and Ecology* **150**, 163–182.
- LONGO, C., CORRIERO, G., CARDONE, F., MERCURIO, M., PIERRI, C. & MARZANO, C. N. (2020). Sponges from rhodolith beds surrounding Ustica Island marine protected area (southern Tyrrhenian Sea), with a comprehensive inventory of the Island sponge fauna. *Scientia Marina* **84**, 297–308.
- LOPEZ-ACOSTA, M., MALDONADO, M., GRALL, J., EHRHOLD, A., SITJA, C., GALOBBART, C., PEREZ, F. F. & LEYNAERT, A. (2022). Sponge contribution to the silicon cycle of a diatom-rich shallow bay. *Limnology and Oceanography* **67**, 2431–2447.
- LOREAU, M. (2000). Biodiversity and ecosystem functioning: recent theoretical advances. *Oikos* **91**(1), 3–17.
- LOREAU, M. & DE MAZANCOURT, C. (2008). Species synchrony and its drivers: neutral and nonneutral community dynamics in fluctuating environments. *American Naturalist* **172**, E48–E66.
- LYONS, D. A., SCHEIBLING, R. E. & VAN ALSTYNE, K. L. (2010). Spatial and temporal variation in DMSP content in the invasive seaweed *Codium fragile* ssp. *fragile*: effects of temperature, light and grazing. *Marine Ecology Progress Series* **417**, 51–61.
- MACDONALD, J. A. & WEIS, J. S. (2013). Fish community features correlate with prop root epibionts in Caribbean mangroves. *Journal of Experimental Marine Biology and Ecology* **441**, 90–98.
- MAGURRAN, A. E. & HENDERSON, P. A. (2018). More than the sum of the parts: annual partitioning within spatial guilds underpins community regulation. *Proceedings of the Royal Society B-Biological Sciences* **285**, 20180659.
- MAO, J., BURDETT, H. L., MCGILL, R. A. R., NEWTON, J., GULLIVER, P. & KAMENOS, N. A. (2020). Carbon burial over the last four millennia is regulated by both climatic and land use change. *Global Change Biology* **26**, 2496–2504.
- MARINS, B. V., AMADO, G. M., BARBARINO, E., PEREIRA, G. H. & LONGO, L. L. (2014). Seasonal changes in population structure of the tropical deep-water kelp *Laminaria abyssalis*. *Phycological Research* **62**(1), 55–62.
- MARRACK, E. C. (1999). The relationship between water motion and living rhodolith beds in the Southwestern Gulf of California, Mexico. *PALAIOS* **14**, 159–171.
- MARTIN, S. & HALL-SPENCER, J. M. (2017). Effects of ocean warming and acidification on rhodolith/maerl beds. In *Rhodolith/Maerl Beds: A Global Perspective* (eds R. RIOSMENA-RODRIGUEZ, W. NELSON and J. AGUIRRE), pp. 55–85. Springer, Cham.
- MCNICHOLL, C., KOCH, M. S. & HOFMANN, L. C. (2019). Photosynthesis and light-dependent proton pumps increase boundary layer pH in tropical macroalgae: a proposed mechanism to sustain calcification under ocean acidification. *Journal of Experimental Marine Biology and Ecology* **521**, 151208.
- MICHALET, R., BROOKER, R. W., CAVIERES, L. A., KIKVIDZE, Z., LORTIE, C. J., PUGNAIRE, F. I., VALIENTE-BANUET, A. & CALLAWAY, R. M. (2006). Do biotic interactions shape both sides of the humped-back model of species richness in plant communities? *Ecology Letters* **9**, 767–773.
- MOURA, R. L., ABIERI, M. L., CASTRO, G. M., CARLOS, L. A., CHIROQUE-SOLANO, P. M., FERNANDES, N. C., TEIXEIRA, C. D., RIBEIRO, F. V., SALOMON, P. S., FREITAS, M. O., GONCALVES, J. T., NEVES, L. M., HACKRADT, C. W., FELIX-HACKRADT, F., ROLIM, F. A., ET AL. (2021). Tropical rhodolith beds are a major and belittled reef fish habitat. *Scientific Reports* **11**, 794.
- MOURA, R. L., AMADO, G. M., MORAES, F. C., BRASILEIRO, P. S., SALOMON, P. S., MAHIQUES, M. M., BASTOS, A. C., ALMEIDA, M. G., SILVA, J. M., ARAUJO, B. F., BRITO, F. P., RANGEL, T. P., OLIVEIRA, B. C. V., BAHIA, R. G., PARANHOS, R. P., ET AL. (2016). An extensive reef system at the Amazon River mouth. *Science Advances* **2**(2), e15012.
- NAEEM, S., DUFFY, J. E. & ZAVALA, E. (2012). The functions of biological diversity in an age of extinction. *Science* **336**, 1401–1406.
- NELSON, W. A. (2009). Calcified macroalgae – critical to coastal ecosystems and vulnerable to change: a review. *Marine and Freshwater Research* **60**, 787–801.
- NETO, J. M., BERNARDINO, A. F. & NETTO, S. A. (2021). Rhodolith density influences sedimentary organic matter quantity and biochemical composition, and nematode diversity. *Marine Environmental Research* **171**, 105470.
- NEVES, P., SILVA, J., PEÑA, V. & RIBEIRO, C. (2021). “Pink round stones” – rhodolith beds: an overlooked habitat in Madeira Archipelago. *Biodiversity and Conservation* **30**, 3359–3383.
- NEVES, S. B. & COSTA, K. G. (2022). Diversity of benthic fauna of rhodoliths and sediments deposited on sandstone reefs in Southeast Brazil. *Ocean and Coastal Research* **70**, e22010.
- NOISETTE, F., DUONG, G., SIX, C., DAVOULT, D. & MARTIN, S. (2013). Effects of elevated CO₂ on the metabolism of a temperate rhodolith *Lithothamnion coralloides* grown under different temperatures. *Journal of Phycology* **49**, 746–757.
- NORLING, P. & KAUTSKY, N. (2008). Patches of the mussel *Mytilus* sp. are islands of high biodiversity in subtidal sediment habitats in the Baltic Sea. *Aquatic Botany* **4**, 75–87.
- OMACHI, C. Y., ASP, N. E., SIEGLE, E., COUCEIRO, M. A. A., FRANCI, R. B. & THOMPSON, F. L. (2019). Light availability for reef-building organisms in a plume-influenced shelf. *Continental Shelf Research* **181**, 25–33.
- OSPAR (2008). <https://www.ospar.org/work-areas/bdc/species-habitats/list-of-threatened-declining-species-habitats/habitats/maerl-beds>
- OTERO-FERRER, F., COSME, M., TUYA, F., ESPINO, F. & HAROUN, R. (2020). Effect of depth and seasonality on the functioning of rhodolith seabeds. *Estuarine Coastal and Shelf Science* **235**, 106579.
- OTERO-FERRER, F., MANNARA, E., COSME, M., FALACE, A., MONTIEL-NELSON, J. A., ESPINO, F., HAROUN, R. & TUYA, F. (2019). Early-faunal colonization patterns of discrete habitat units: a case study with rhodolith-associated vagile macrofauna. *Estuarine Coastal and Shelf Science* **218**, 9–22.
- PASCELLI, C., RIUL, P., RIOSMENA-RODRIGUEZ, R., SCHERNER, F., NUNES, M., HALL-SPENCER, J. M., DE OLIVEIRA, E. C. & HORTA, P. (2013). Seasonal and depth-driven changes in rhodolith bed structure and associated macroalgae off Arvoredo Island (Southeastern Brazil). *Aquatic Botany* **111**, 62–65.
- PEÑA, V. & BÁRBARA, I. (2008). Biological importance of an Atlantic European maerl bed off Benecia Island (Northwest Iberian Peninsula). *Botanica Marina* **51**, 493–505.
- PEÑA, V., BÁRBARA, I., GRALL, J., MAGGS, C. A. & HALL-SPENCER, J. M. (2014). The diversity of seaweeds on maerl in the NE Atlantic. *Marine Biodiversity* **44**, 533–551.
- PEREIRA, G. H., AMADO, G. M., DE MOURA, R. L., BASTOS, A. C., GUIMARAES, S., SALGADO, L. T., FRANCI, R. B., BAHIA, R. G., ABRANTES, D. P., GUTH, A. Z. & BRASILEIRO, P. S. (2012). Extensive rhodolith beds cover the summits of Southwestern Atlantic ocean seamounts. *Journal of Coastal Research* **28**, 261–269.
- PEREIRA, G. H., VERAS, P. D., FRANCI, R. B., DE MOURA, R. L., PINHEIRO, H. T., GIBRAN, F. Z., MATHEUS, Z., NEVES, L. M. & AMADO, G. M. (2015). Effects of the sand tilefish *Malacanthus plumieri* on the structure and dynamics of a rhodolith bed in the Fernando de Noronha Archipelago, tropical West Atlantic. *Marine Ecology Progress Series* **541**, 65–73.
- PEREIRA-FILHO, G. H., FRANCI-FILHO, R. B., PIEROZZI-JR, I., PINHEIRO, H. T., BASTOS, A. C., DE MOURA, R. L., MORAES, F. C., MATHEUS, Z., BAHIA, R. D. G. & AMADO-FILHO, G. M. (2015). Sponges and fish facilitate succession from rhodolith beds to reefs. *Bulletin of Marine Science* **91**(1), 45–46.
- PEREZ-PERIS, I., NAVARRO-MAYORAL, S., DE ESTEBAN, M. C., TUYA, F., PEÑA, V., BÁRBARA, I., NEVES, P., RIBEIRO, C., ABREU, A., GRALL, J., ESPINO, F., BOSCH, N. E., HAROUN, R. & OTERO-FERRER, F. (2023). Effect of depth across a latitudinal gradient in the structure of rhodolith seabeds and associated biota across the eastern Atlantic Ocean. *Diversity-Basel* **15**, 103.
- PERRY, C. T. (2005). Morphology and occurrence of rhodoliths in siliciclastic, intertidal environments from a high latitude reef setting, southern Mozambique. *Coral Reefs* **24**, 201–207.
- QUI-MINET, Z. N., DELAUNAY, C., GRALL, J., SIX, C., CARIU, T., BOHNER, O., LEGRAND, E., DAVOULT, D. & MARTIN, S. (2018). The role of local environmental changes on maerl and its associated non-calcareous epiphytic flora in the Bay of Brest. *Estuarine Coastal and Shelf Science* **208**, 140–152.
- RAVAGLIOLI, C., LANGENECK, J., CAPOCCHI, A., CASTELLI, A., FONTANINI, D., GRIBBEN, P. E. & BULLERI, F. (2021). Positive cascading effects of epiphytes enhance the persistence of a habitat-forming macroalga and the biodiversity of the associated invertebrate community under increasing stress. *Journal of Ecology* **109**, 1078–1093.
- RENDINA, F., BUONOCORE, E., DI MONTANARA, A. C. & RUSSO, G. F. (2022). The scientific research on rhodolith beds: a review through bibliometric network analysis. *Ecological Informatics* **70**, 101738.
- RINDE, E., MOY, S. R., TVEITEN, L. A., KVILE, K. Ø., WALDAY, M. G., CHRISTIE, H., BRKJACIC, M. S., KILE, M. R., BEKKBY, T., GITMARK, J. K.,

- MJELDE, M., FAGERLI, C. W., OUG, E. & ANGLÉS D'AURIAC, M. (2022). Feltbasert kunnskap, metodikk og kriterier for økologisk kvalitet til et utvalg av marine naturtyper. NIVA rapport 7691–2022 (summary in English).
- RINDI, F., BRAGA, J. C., MARTIN, S., PEÑA, V., LE GALL, L., CARAGNANO, A. & AGUIRRE, J. (2019). Coralline algae in a changing Mediterranean Sea: how can we predict their future, if we do not know their present? *Frontiers in Marine Science* **6**, 723.
- RIUL, P., LAGOUTH, P., PAGLIOSA, P. R., CHRISTOFFERSEN, M. L. & HORTA, P. A. (2009). Rhodolith beds at the easternmost extreme of South America: community structure of an endangered environment. *Aquatic Botany* **90**, 315–320.
- ROCHA, L. A. (2003). Patterns of distribution and processes of speciation in Brazilian reef fishes. *Journal of Biogeography* **30**, 1161–1171.
- ROCHA, L. A., ROSA, I. L. & FEITOZA, B. M. (2000). Sponge-dwelling fishes of Northeastern Brazil. *Environmental Biology of Fishes* **59**, 453–458.
- SALDAÑA, P. H., ANGELINI, C., BERTNESS, M. D. & ALTIERI, A. H. (2024). Dead foundation species drive ecosystem dynamics. *Trends in Ecology & Evolution* **39**, 294–305.
- SANDES, J., MORAES, F., PINHEIRO, U. & MURICY, G. (2021). Taxonomy and distribution of *Didiscus* and *Myrmeioderma* (Demospongiae: Axinellida) off the mouths of the two largest rivers in Brazil, with description of four new species. *Marine Biodiversity* **51**, 27.
- SAÑE, E., CHIOCCI, F. L., BASSO, D. & MARTORELLI, E. (2016). Environmental factors controlling the distribution of rhodoliths: an integrated study based on seafloor sampling, ROV and side scan sonar data, offshore the W-pontine archipelago. *Continental Shelf Research* **129**, 10–22.
- SANTÍN, A., WIRTZ, P., NEVES, P. & RIBEIRO, C. (2024). Filling gaps in the Webbsia marine diversity: the Madeiran sponge fauna. *Continental Shelf Research* **277**, 105230.
- SCHOB, C., MICHALET, R., CAVIERES, L. A., PUGNAIRE, F. I., BROOKER, R. W., BUTTERFIELD, B. J., COOK, B. J., KIKVIDZE, Z., LORTIE, C. J., XIAO, S., AL HAYEK, P., ANTHELME, F., CRANSTON, B. H., GARCIA, M. C., ET AL. (2014). A global analysis of bidirectional interactions in alpine plant communities shows facilitators experiencing strong reciprocal fitness costs. *New Phytologist* **202**, 95–105.
- SCHUBERT, N., HOFMANN, L. C., SAÁ, A. C. A., MOREIRA, A. C., ARENHART, R. G., FERNANDES, C. P., DE BEER, D., HORTA, P. A. & SILVA, J. (2021). Calcification in free-living coralline algae is strongly influenced by morphology: implications for susceptibility to ocean acidification. *Scientific Reports* **11**, 11232.
- SCIBERRAS, M., RIZZO, M., MIFUD, J. R., CAMILLERI, K., BORG, J. A., LANFRANCO, E. & SCHEMBRI, P. J. (2009). Habitat structure and biological characteristics of a maerl bed off the northeastern coast of the Maltese islands (central Mediterranean). *Marine Biodiversity* **39**, 251–264.
- SILLIMAN, B. R., BERTNESS, M. D., ALTIERI, A. H., GRIFFIN, J. N., BAZTERRICA, M. C., HIDALGO, F. J., CRAIN, C. M. & REYNA, M. V. (2011). Whole-community facilitation regulates biodiversity on Patagonian rocky shores. *PLoS One* **6**, e24502.
- SISSINI, M. N., KOERICH, G., DE BARROS-BARRETO, M. B., COUTINHO, L. M., GOMES, F. P., OLIVEIRA, W., COSTA, I. O., NUNES, J. M. D., HENRIQUES, M. C., VIEIRA-PINTO, T., TORRANO-SILVA, B. N., OLIVEIRA, M. C., LE GALL, L. & HORTA, P. A. (2022). Diversity, distribution, and environmental drivers of coralline red algae: the major reef builders in the Southwestern Atlantic. *Coral Reefs* **41**, 711–725.
- SNEED, E. D. & FOLK, R. L. (1958). Pebbles in the lower Colorado river, Texas. A study in particle morphogenesis. *The Journal of Geology* **66**, 114–150.
- SOLÓRZANO, M. R. & URGORRI, V. (1991). Nuevas señalizaciones de demosponjas en el litoral Gallego (No. de la Península Ibérica). *Thalassas* **9**, 79–86.
- STACHOWICZ, J. J. (2001). Mutualism, facilitation, and the structure of ecological communities. *Bioscience* **51**, 235–246.
- STELLER, D. L. & CACERES-MARTINEZ, C. (2009). Coralline algal rhodoliths enhance larval settlement and early growth of the Pacific calico scallop *Argopecten ventricosus*. *Marine Ecology Progress Series* **396**, 49–60.
- STELLER, D. L. & FOSTER, M. S. (1995). Environmental factors influencing distribution and morphology of rhodoliths in Bahía Concepción, BCS, Mexico. *Journal of Experimental Marine Biology and Ecology* **194**, 201–212.
- STELLER, D. L., RIOSMENA-RODRIGUEZ, R., FOSTER, M. S. & ROBERTS, C. A. (2003). Rhodolith bed diversity in the Gulf of California: the importance of rhodolith structure and consequences of disturbance. *Aquatic Conservation-Marine and Freshwater Ecosystems* **13**, S5–S20.
- STENECK, R. S., GRAHAM, M. H., BOURQUE, B. J., CORBETT, D., ERLANDSON, J. M., ESTES, J. A. & TEGNER, M. J. (2002). Kelp forest ecosystems: biodiversity, stability, resilience and future. *Environmental Conservation* **29**, 436–459.
- STEWART, H. A., JANIÁK, D. S., WRIGHT, J. L., HUNT, D., CORTES, A. C., POWELL, K. T., CHAPMAN, L. J. & ALTIERI, A. H. (2022). Epibiont community composition of red mangroves *Rhizophora mangle* are contingent on root characteristics. *Marine Ecology Progress Series* **686**, 15–35.
- TEICHERT, S. (2014). Hollow rhodoliths increase Svalbard's shelf biodiversity. *Scientific Reports* **4**, 6972.
- TEICHERT, S. & FREIWALD, A. (2014). Polar coralline algal CaCO₃ production rates correspond to intensity and duration of the solar radiation. *Biogeosciences* **11**, 833–842.
- TEPER, S. H., PARRISH, C. C. & GAGNON, P. (2022). Multiple trophic tracer analyses of subarctic rhodolith (*Lithothamnion glaciale*) bed trophodynamics uncover bottom-up forcing and benthic-pelagic coupling. *Frontiers in Marine Science* **9**, 899812.
- THOMSEN, M. S., ALTIERI, A. H., ANGELINI, C., BISHOP, M. J., BULLERI, F., FARHAN, R., FRUHLING, V. M. M., GRIBBEN, P. E., HARRISON, S. B., HE, Q., KLINGHARDT, M., LANGENECK, J., LANHAM, B. S., MONDARDINI, L., MULDER, Y., ET AL. (2022). Heterogeneity within and among co-occurring foundation species increases biodiversity. *Nature Communications* **13**, 581.
- THOMSEN, M. S., ALTIERI, A. H., ANGELINI, C., BISHOP, M. J., GRIBBEN, P. E., LEAR, G., HE, Q., SCHIEL, D. R., SILLIMAN, B. R., SOUTH, P. M., WATSON, D. M., WERNBERG, T. & ZOTZ, G. (2018). Secondary foundation species enhance biodiversity. *Nature Ecology & Evolution* **2**, 634–639.
- THOMSEN, M. S., HILDEBRAND, T., SOUTH, P. M., FOSTER, T., SICILIANO, A., OLDACH, E. & SCHIEL, D. R. (2016). A sixth-level habitat cascade increases biodiversity in an intertidal estuary. *Ecology and Evolution* **6**, 8291–8303.
- THOMSEN, M. S., WERNBERG, T., ALTIERI, A. H., TUYA, F., GULBRANSEN, D., MCGLATHERY, K. J., HOLMER, M. & SILLIMAN, B. R. (2010). Habitat cascades: the conceptual context and global relevance of facilitation cascades via habitat formation and modification. *Integrative and Comparative Biology* **50**, 158–175.
- TILMAN, D. (1999). The ecological consequences of changes in biodiversity: a search for general principles. *Ecology* **80**, 1455–1474.
- TILMAN, D. & DOWNING, J. A. (1994). Biodiversity and stability in grasslands. *Nature* **367**, 363–365.
- TUYA, F., SCHUBERT, N., AGUIRRE, J., BASSO, D., BASTOS, E. O., BERCHEZ, F., BERNARDINO, A. F., BOSCH, N. E., BURDETT, H. L., ESPINO, F., FERNÁNDEZ-GARCÍA, C., FRANCINI, R. B., GAGNON, P., HALL-SPENCER, J. M., HAROUN, R., ET AL. (2023). Levelling-up rhodolith-bed science to address global-scale conservation challenges. *Science of the Total Environment* **892**, 164818.
- VALE, N. F. L., BRAGA, J. C., BASTOS, A. C., MORAES, F. C., KAREZ, C. S., BAHIA, R. G., LEAO, L. A., PEREIRA, R. C., AMADO, G. M. & SALGADO, L. T. (2022). Structure and composition of rhodolith beds from the Sergipe-Alagoas Basin (NE Brazil, Southwestern Atlantic). *Diversity-Basel* **14**, 282.
- VAN DE KOPPEL, J., VAN DER HEIDE, T., ALTIERI, A. H., ERIKSSON, B. K., BOUMA, T. J., OLFF, H. & SILLIMAN, B. R. (2015). Long-distance interactions regulate the structure and resilience of coastal ecosystems. *Annual Review of Marine Science* **7**, 139–158.
- VILLAS-BÔAS, A. B., TÂMEGA, F. T. D., ANDRADE, M., COUTINHO, R. & FIGUEIREDO, M. A. D. (2014). Experimental effects of sediment burial and light attenuation on two coralline algae of a deep water rhodolith bed in Rio de Janeiro, Brazil. *Cryptogamie Algologie* **35**, 67–76.
- VOERMAN, S. E., MARSH, B. C., BAHIA, R. G., PEREIRA-FILHO, G. H., YEE, T. W., BECKER, A. C. F., AMADO-FILHO, G. M., RUSECKAS, A., TURNBULL, G. A., SAMUEL, I. D. W. & BURDETT, H. L. (2022). Ecosystem engineer morphological traits and taxon identity shape biodiversity across the euphotic-mesophotic transition. *Proceedings of the Royal Society B-Biological Sciences* **289**, 20211834.
- WITMAN, J. D. (1987). Subtidal coexistence – storms, grazing, mutualism, and the zonation of kelps and mussels. *Ecological Monographs* **57**, 167–187.
- YAKOVIS, E. & ARTEMIEVA, A. (2017). Cocksles, barnacles and ascidians compose a subtidal facilitation cascade with multiple hierarchical levels of foundation species. *Scientific Reports* **7**, 237.

(Received 7 January 2024; revised 4 September 2024; accepted 10 September 2024)