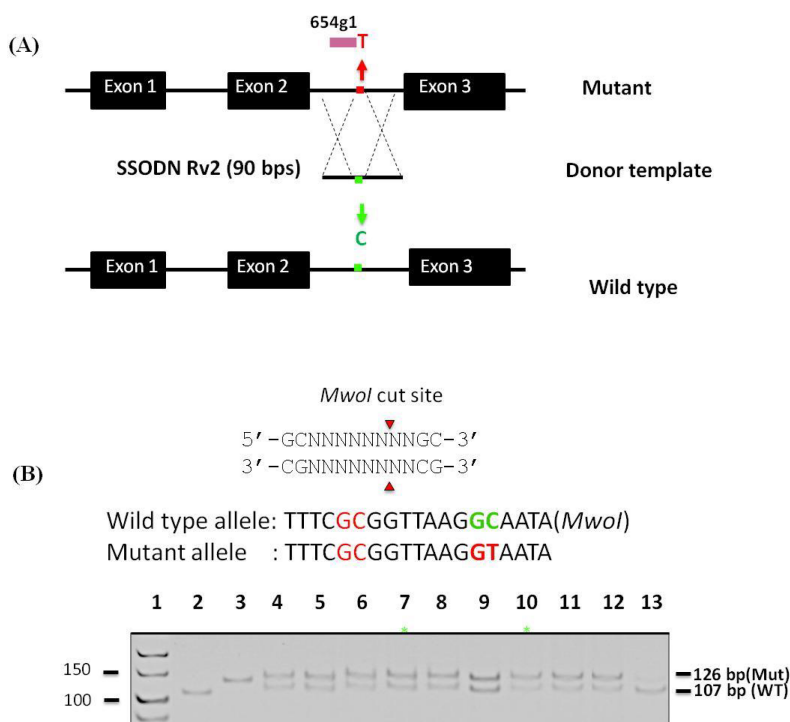


Figure 3

Human $\beta^{IVS2-654}$ -globin gene

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P0967 | Discovery, molecular modeling and biological evaluation of 1,3,5-triaril-pyrazole derivatives as novel Estrogen Receptor Modulators

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Introduction

The biological actions of estrogens are mediated by estrogen binding to one of two specific estrogen receptors (ERs), ER α and ER β . The ERs, which belong to the nuclear receptor superfamily of ligand-regulated transcription factors, are products of different genes and exhibit tissue- and cell-type specific expression. Clinically relevant, ERs are currently considered relevant drug targets for prevention and/or treatment of several diseases (from cancer, metabolic and cardiovascular, inflammation, and osteoporosis to neurodegeneration). In this work, new derivatives of 1,3,5-triaril-pyrazoles were designed, synthesized and evaluated *in vitro* as Estrogen Receptor Modulators.

Method

Virtual screening and Fluorescent Polarization assay together with real-time microphotography cell proliferation analysis and ER-based drug discovery strategies (1) were used to evaluate a chemical library enriched in ER α -based 1,3,5-triaril-pyrazoles derivatives.

Results

In this work, we found that certain substituted pyrazoles are high-affinity ligands for the hER α and hER β (from 40 to 200 nM). The results obtained in the binding assay with hER α correlate with the obtained in the *in silico* studies, confirming that the presence of hydroxyl groups on

aromatic rings B and C in the structure of 1,3,5-triaryl pyrazole derivatives is critical for a favorable interaction with ER α . They can also activate ER α -dependent transcription in adenocarcinoma breast cancer cells with high potency (EC₅₀ < 1 μ M) but with a relative efficacy lower (50-80%) than pure agonist E2. They also promote cell proliferation of ER+ breast cancer cells with EC₅₀ below 5 μ M. However, antiestrogenic activity of E2-induced breast cancer cell proliferation was observed in the presence of 1,3,5-triaryl-pyrazoles derivative concentration above 5 μ M.

Conclusions

In this study, we have identify novel 1,3,5-triaryl-pyrazoles derivatives as pure agonists of ER and, other ones, with partial agonist/antagonist activity on ER α -mediated transcription and proliferation of human ER+ breast adenocarcinoma cells.

Reference

1. Guerra-Rodríguez M et al. Discovery of Highly Functionalized 5-hydroxy-2H-pyrrol-2-ones That Exhibit Antiestrogenic Effects in Breast and Endometrial Cancer Cells and Potentiate the Antitumoral Effect of Tamoxifen. *Cancers* 2022, 14(21):5174.

P0969 | Confirmation of Proposed Xenohormetic Protection by Phytochemicals in Human Microphysiological Models of Tissue Vasculogenesis and Cancer-Induced Angiogenesis

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Introduction/Background & aims

Citrus flavonoids such as Naringenin present in grapefruit exert anti-stress effects in the plants and phytoalexins such as Glyceollin present in soybean are involved in the plant defense against infection. Understanding these effects in the context of human tissue and organ models is a necessary step in defining the health-promoting effects of diets enriched with specific micronutrients and improving the accuracy of decisions to scale up commercial production of candidate phytochemicals to accelerate preclinical research and eventual translation. We utilized microphysiological assays of 3D tissue vasculogenesis and cancer-induced angiogenesis engineered in our laboratory to test the inhibitory effects of phytochemicals from a USDA panel on multiple morphogenetic processes relevant to tumor vascularization.

Method/Summary of work

HLF and HUVEC were incorporated in hydrogels composed of collagen type I and fibrin to create stromal vascular tissues. We engineered a PDMS device for culturing an array of disc-shaped tissue constructs to test the dose-dependent effects of Naringenin (grapefruit flavonoid), its derivative Sakuranetin, and Glyceollin (soybean phytoalexin) on vasculogenesis. We performed morphometric analysis of 3D confocal image data to quantify the structural features of emergent vascular networks in the tissues. Membrane-free organ chips that enable patterning of adjacent but contiguous layers of tissues with defined compositions were loaded with the same cell-hydrogel mixture used for tissue vasculogenesis in one lane, and hydrogel containing spheroids derived from the biopsy of a patient with highly aggressive triple negative breast cancer in the adjacent lane. We quantified phytochemical effects on vascular morphometry and sprouting in the cancer-containing tissue space.

Results/Discussion

A tissue array assay allowed rapid screening of phytochemical dose responses (Fig. 1A). Naringenin (NAR) potently inhibited de novo vasculogenesis in stromal vascular tissues at and above 25 μ M (Fig. 1A). Sakuranetin (SAK) only partially inhibited vasculogenesis at 125 μ M, despite being a metabolic derivative of NAR. Glyceollin (GLY) inhibited vasculogenesis, though to a lesser degree than NAR at the same doses. We used the minimal effective dose of 25 μ M identified in vasculogenesis studies to test phytochemical inhibition of cancer-induced angiogenic processes and found that NAR completely inhibits vascularization, whereas SAK has no effect (Fig. 1C).

Conclusion(s)

Our study provides quantitative evidence of potent anti-vasculogenic, and anti-angiogenic effects elicited by USDA-specified candidate phytochemicals in multiple human cell-based MPS models. Therefore, we conclude that further investigation of phytochemicals as potential therapeutic agent in MPS models of human cancers is warranted.