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INTERACTIONSHIP CHLORIDE-NITRATE ON NUTRITION OF TOMATO PLANTS.

by

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SUMMARY.

Tomato plants vard "Marglobe" were cultivated in greenhouse by hydroponic system in order to study the effect of five treatments of Cl⁻,0, 8,16,24 and 32 me.1⁻¹ with five different concentrations of NO₂⁻: 6, 12,18,24 and 30 me.1⁻¹. We found a cooperative effect of Cl⁻ and NO₃⁻ increasing concentration of both until 24 me.1⁻¹ of Cl⁻ and 18 me.1⁻¹ of NO₃⁻ which means an increase of NO₃⁻ in the nutrient solution thus decreasing the toxic effect of Cl⁻ in plants. Org N/NO₃⁻-N in the tissues is the index that best indicates its maximum growth, its value is considered to reach 1.0.

INTRODUCTION.

The presence of Cl⁻ in the irrigation water is one of the problems that affect arid and semiarid areas where there is a high evapotranpiration which produces salination in the soil.

Dropping irrigation allow the control of soil solution due to slow and continuous fertilization.

Tomato plants permit a concentration of Cl⁻ in nutrient solution of 15 me.l⁻¹ without decreasing its growth and production (Cadahia,1968). The fertilization of tomato plants with high quantities of NO_3^- produces a decrease of Cl⁻ in the plant (Hernando et al.1964). On the other hand the excess of Cl⁻ could produce a decrease in the NO3 content in the plant running the risk of inducing nitrogen deficiencies (Torres and Bingham, 1973).

Maas et al (1977) point out, using different sources, that -1.0 is the maximum osmotic potential tolerable in tomato plants.

Our work, therefore, focused in the study of Cl⁻ and NO \bar{z} relation in nutrient solutions, which allows the utilization of water with a high concentration of Cl⁻ by means of the increase fertilization of NO \bar{z} , as well as to find out to what extend Cl⁻ could affect assimilation, transporting and reduction of NO \bar{z} in tomato plants.

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MATERIAL AND METHODS.

Tomato seeds (Lycopersicum esculentum vard Marglobe) were planted in volcanic sand. After 30 days, they were transplanted in pots containing volcanic sand with an inferior draining system. Five plants per pot were planted, two were recollected at the flowering time, two at fruiting time and the fifth on was picked two weeks later in order to determine the growth.

25 different treatments were conducted in the greenhouse, using five different concentrations of Cl⁻ (0,8,16,24,30 me.l⁻¹). Each treatment was repeated three times. Cations were kept at the same relative level (K : la : Mg = 0.35 : 0.45 : 0.20). H₂PO₄ and SO₄⁼ were kept at the same absolute level (H₂PO₄ : 0.5 me.l⁻¹; SO₄⁼ : 3.5 me.l⁻¹).

C1⁻, NO₃ and organic Nitrogen were analyzed in leaves, roots and stem juice at flowering time and fruiting time. C1⁻ was determinated by extraction with water and valoration with silver nitrate (Ulrich and Johnson, 195). NO₃ was extracted with water and determinated by Cataldo's method modificated in a higher concentration of salicilic acid in sulphuric acid (10% w/v), (Cataldo et at, 1975). Organic nitrogen was determinated by microkjeldahl method.

RESULTS AND DISCUSSION.

Fig.1 depicts as an index of growth the fresh weight of the complete plant with its fruits. We found and increase of growth with the increase of NO3 concentration in nutrient solution until 18 me.1⁻¹, and a cooperative effect of C1⁻ up to 16 me.1⁻¹ of C1⁻ in nutrient solution with 12 me.1⁻¹ NO3 and 24 me.1⁻¹ of C1⁻ with 18 me.1⁻¹ of NO3. In high concentrations of NO3 (24 and 30 me.1⁻¹) the cooperative effect of C1⁻ and NO3 disappears.



A equilibrate relation of Cl^- and NO_3^- increases the resistence of the plant to lower osmotic potential, achieving the best results at -1.8. Higher concentrations produce a significative decrease of the growth.

Cl contents in leaves, roots and stems in flowering and fruiting time (Table 1) decreases when the NO_3^- concentration in nutrient solution increases. That decrease could be produced by the interaction of Cl and NO_3^- passing through tonoplaste at the entry in the cell vacuoles (Cram, 1973). The decrease of Cl facilitates a higher resistence of the plant to the specific toxic effect of Cl.

The toxic concentration of Cl⁻ in the stem juice for tomato plants has been pointed out by Cadahia (1968) in 118 me.l⁻¹ of Cl⁻ at flowering time and 140 me.l⁻¹ of Cl⁻ at fruiting time and he formed these values with 15 me.l⁻¹ of Cl⁻ in nutrient solution. In our experiment we found these concentrations in stem juice with 16 me.l⁻¹ of Cl⁻,12 me.l⁻¹ of NO3 and 24 me.l⁻¹ of Cl -18 me.l⁻¹ of NO3. This treatment (Cl₂₄N₁₈) is the best (Fig.1).

Values of organic Nitrogen change according to the type of the part of the plant and the sampling time. In the leaves at the flowering time there is a depressive effect of $C1^-$ with 6 and 12 me.1⁻¹ of NO₃⁻¹ in nutrient solution. This effect is significant between Cl 0 and Cl 8 treatments. This could be possible by a decrease in the malate sinthesis as a response to a high interne concentration of $C1^-$, like it was pointed out by Schnabl and Raschke (1978) in stomatic cells, and that it would affect the absorption in roots and transport of NO₃⁻ to the shoots (Ben-Zioni et al, 1971).

In the leaves at fruiting time and in the roots at flowering time there is not a significative decrease of organic nitrogen values.

In the stem juice at flowering and fruiting time the depressive effect of Cl⁻ appears in higher concentrations of NO $\overline{3}$ (18, 24, and 32 me.l⁻¹).

The relationship between org.N and $NO_{3} - N$ (mg/mg) is the index that best agrees with the growth (Fig.2). We can see that this relationship increases with Cl⁻ up to 16 me.l⁻¹ of Cl⁻ in nutrient solution with 6 and 12 me.l⁻¹ of NO₃ and 24 me.l⁻¹ of Cl with 18 me.l⁻¹ of NO₃.

Ferrari et al (1973) pointed out the existence of two different pools of NO_3 in the plant cell. One of them is the vacuolar pool (metabolically inactive) and the other one is the citoplasmatic pool (metabolically active).

The competition between Cl^- and NO_3^- to enter in the vacuola may induce a decrease of total NO_3^- in the cell, but the metabolic pool can increase, in which case it would allow to an increment in the reduction of NO₃ due to the fact that NO_3^- fluxes through the cytoplasme which regulates the activity of nitrate reductase (Shanner and Boyer, 1976).

This effect could be at the root level where we did not notice significant differences in organic nitrogen (see Table 2), due to the fact that the tomato plants may have a reduction of NO_3 in roots (Suder-Moraw and Buczek, 1977). The reduced nitrogen could be translocated to the shoots.

<u>Table 1</u> Cl⁻ contens L:leaves % d.w. R=roots % d.w. S=Stemjuice me.l.⁻¹

ution	6	FL L 0.1 R 0.6 S 1.6	0 FR 0.3 0.4 10	FL 2.1 2.7 108	8 FR 1.9 3.1 130	FL 2.7 3.3 125	16 FR 3.0 3.4 147	FL 3.1 4.2 133	24 FR 3.0 4.4 185	FL 3.2 3.6 148	32 FR 3.1 3.6 181
ent soluti	12	L 0.2 R 0.6 S 1.6	0.3 0.4 17.0	2.1 2.6 100	1.8 3.2 141	3.0 3.3 118	2.9 3.5 162	3.0 3.7 122	3.3 4.3 174	2.9 3.2 118	3.2 3.7 211
nutrient	18	L 0.2 R 0.5 S 1.6	0.3 0.4 21	1.8 2.6 81	1.6 2.8 123	2.5 2.9 93.2	2.1 3.0 127	2.6 3.6 116	2.5 3.3 139	2.4 3.0 101	2.6 4.0 161
NO ₃ me.1 ⁻¹ in	24	L 0.2 R 0.5 S 1.6	0.3 0.3 19	1.4 2.3 66	1.5 2.1 77	1.7 2.5 76	1.7 2.6 126	1.8 2.5 89	2.3 4.3 171	2.1 3.0 91	2.7 3.8 178
	30	L 0.2 R 0.5 S 1.6	0.3 0.4 17	1.0 1.7 55	1.4 2.2 96	1.4 1.8 74	1.5 2.1 92	1.7 2.7 87	2.6 4.0 171	2.0 3.1 90.0	2.6 3.9 197

<u>Cl me.l. 1 in nutrient solution</u>

FL= Flowering time FR= Fruiting time

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				Table	2				-
Org N. L=leaves	%	in	d.w.	R=roots	%	in	d.w.	S=Stemjuice	g.1 ⁻¹

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<u>Cl⁻ me.l.⁻¹ in nutrient solution</u>

		ì		0			8			16			24			32	
(me.1 ⁻¹) in nutrient solution	6		FL 5.5 1.7 0.4		FR 4.0 1.8 0.7	FL 3.5 1.6 0.3		FR 3.4 1.5 0.3	FL 3.3 1.7 0.3		FR 3.4 1.5 0.3	FL 3.4 1.8 0.3		FR 3.5 1.5 0.3	FL 3.2 1.8 0.3		FR 3.3 1.6 0.3
	12	R	5.5 1.7 0.5		4.3 2.2 0.8	4.4 1.9 0.4		4.2 1.8 0.5	3.7 1.7 0.5		4.3 1.8 0.4	3.9 2.0 0.4		4.1 1.9 0.5	4.2 2.3 0.4		4.2 1.8 0.4
	18	R	5.1 2.3 0.8		4.6 2.3 0.8	4.5 2.1 0.5		4.4 1.9 0.5	4.1 2.4 0.5		4.4 1.9 0.4	4.5 2.1 0.6		4.4 2.0 0.8	4.3 2.3 0.4		3.8 1.9 0.4
	24	R	4.5 2.4 0.7		4.5 2.2 0.8	4.3 2.0 0.5		4.7 2.0 0.5	4.4 1.9 0.6		5.2 2.2 0.4	4.8 2.1 0.6		4.6 2.2 0.4	5.0 2.3 0.6		4.6 2.4 0.6
	3 0	R	5.2 2.3 1.0	<u>, </u>	4.7 2.7 0.8	4.1 2.3 0.5		3.8 2.6 0.3	4.5 2.4 0.7		4.7 2.3 0.4	5.0 2.6 0.6		3.9 2.1 0.5	4.6 2.2 0.7		4.1 2.3 0.7
NO ₃		F	L= F1	owerin	g tim	e FR	= Frui	ting	time								

FL= Flowering time FR= Fruiting time



CONCLUSIONS.

The increment of Cl^- in the culture medium could be parcially mitigated by the increase of NO_3^- fertilization in the tomato culture.

In addition this enhances a better resistence to the effect of total salinity.

The cooperative effect of C1⁻ and NO_3^- could be explained by an increase of reduction in the roots and the translocation to the shoots of the reducted nitrogen.

The relation Org N/NO \bar{z} -N in stem juice at flowering time is a good index to study the effect of Cl⁻ in the NO \bar{z} metabolism and its value should be maintained next to 1.0. However, we believe that these results ought to be further studied in order to yield applicable results.

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RESUME : INTERACTION DU CLORURE ET NITRATE EN TOMATES.

Des tomates de la varieté Marglobe ont été cultivées avec une solution noutritive dans une serre pour l'étude de l'effect du Cl⁻. Cinq traitements avec du Cl⁻: 0,8,16,24 et 32 me.1⁻¹ ont été combinés avec cinq concentrations differentes de NO3: 6,12,18,24 et 30 me.1⁻¹. On trouve un effet coopératif du Cl⁻ et du NO3 en augmentant les deux concentrations, celle du Cl⁻ jusqu'à 24 me.1⁻¹ et celle du NO3 jusqu'à 18 me.1⁻¹. Ce qui indique qu'en augmentant le NO3 dans la solution l'effet toxique du Cl⁻ diminu. L'index qui s'ajuste le mieux aux rendements c'est celui de la relation N.org: N-NO3 (mg:mg) dont les valeurs doivent être proches à 1.

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