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Trophic indices for micronektonic fishes reveal their dependence on the microbial system in the North Atlantic

Antonio Bode¹², M. Pilar Olivar² & Santiago Hernández-León³

The importance of microbes for the functioning of oceanic food webs is well established, but their relevance for top consumers is still poorly appreciated. Large differences in individual size, and consequently in growth rates and the relevant spatial and temporal scales involved, make the integration of microorganisms and large metazoans in a common food web framework difficult. Using stable isotopes, this study estimated the trophic position of 13 species of micronektonic fishes to examine the microbial and metazoan contribution to mid trophic level consumers. Vertically migrant species displayed higher trophic positions than non-migrant species in all depth layers. The estimated trophic positions agreed well with those from the literature, but all species displayed mean increases between 0.5 and 0.8 trophic positions when taking into account microbial trophic steps. Trophic position, but not the relative importance of the microbial food web, increased with individual size, suggesting that current estimates of the trophic position of top consumers and of the length of oceanic food webs are too low because they are based only on metazoan trophic steps. This finding calls for a review of trophic position estimates and of the efficiency of trophic transfers along oceanic food webs.

Since the discovery of major trophic pathways in the microbial system of the ocean four decades ago¹ their full integration in the whole food web remained elusive. The current paradigm considers oceanic food webs organized in a continuum of cases between two main types of trophic systems: microbial and metazoan food webs^{2,3}. The former is mainly related to remineralisation and respiration³ but also to the sequestration of carbon in the form of recalcitrant dissolved organic compounds^{4,5}. In contrast, the efficient transfer of organic matter to upper trophic levels (e.g. fisheries⁶), the sequestration of carbon in the deep ocean by sedimenting particles⁷ and the active flux by zooplankton^{8,9} are mainly attributed to the metazoan food web^{10,11}. Both trophic systems, however, are connected by the protozoan grazers allowing large-sized metazoans to consume the otherwise inaccessible dissolved organic matter concentrated by bacteria.

The study of the links between both trophic systems in the ocean has been limited by the large difference in the time and space scales relevant for microbes and large animals¹². Diet studies are unable to capture all the complexity of the food web because of the difficulties in identifying all prey items¹³. Integrated approaches using tracers (e.g. stable isotopes) are limited by the 'invisibility' of microbial pathways to traditional models for estimating trophic positions¹⁴. Consequently, most food web models consider mainly metazoan links and therefore underestimate true trophic positions of top consumers, as evidenced by meta-analysis studies^{15,16}. However, new models are able to track the differential processing of some organic compounds (e.g. amino acids) through the microbial vs. metazoan food webs. Trophic amino acids (as alanine or glutamic acid) are enriched in heavy isotopes with each trophic step, while source amino acids (e.g. phenylalanine), remain essentially unaltered through the food web. Therefore, the comparison of their isotopic composition allows for the estimation of the contribution of heterotrophic protists to the trophic position of metazoan consumers^{17,18}. In this way, the pathways of microbial reworking of the dissolved organic matter can be included because protists represent the main link between microbial and metazoan food webs¹⁻³. Considering microbial steps will thus produce higher

¹Centro Oceanográfico de A Coruña, Instituto Español de Oceanografía, IEO, 15080 A Coruña, Spain. ²Institut de Ciències del Mar, CSIC, 08003 Barcelona, Spain. ³Unidad Asociada ULPGC-CSIC, Instituto de Oceanografía y Cambio Global, IOCAG, Universidad de Las Palmas de Gran Canaria, Campus de Taliarte, 35214 Telde, Gran Canaria, Spain. [⊠]email: antonio.bode@ieo.es

estimations of trophic position, but how this increase in the estimations would affect different consumers along the food web is not known.

Micronektonic fishes are key components of oceanic food webs. They dominate the biomass of consumers¹⁹ and the transfer of energy from zooplankton to higher trophic levels^{20,21}. While their abundance is higher in the mesopelagic domain (200–1000 m depth), the distribution of the different species, including those performing diel vertical migrations also covers the epipelagic (0–200 m), bathypelagic (1000–4000 m) and even abyssopelagic (>4000 m) layers^{22–24}. Therefore, they provide a privileged opportunity to study trophic connections across the ocean. Studies on these organisms focused on changes in their diet and trophic position in relation to their migration habits and vertical distribution. Their trophic position varies within relatively narrow limits as some species are mostly planktivores^{20,25–28} or piscivores^{20,25,29,30}, while increasing significantly with individual size^{20,25,27}. In addition, there is evidence of different feeding strategies for active migrants vs. opportunistic, non-migrant species^{24,30,31}.

As the microbial system is able to recycle several times the produced organic matter, it can be hypothesized that the consideration of microbial steps will have a differential impact on the estimations of trophic position estimates of small and large fishes. The latter are generally predators requiring protein-rich food sources¹⁵ while the former feed mainly on prey near the base of the food web, hence being more affected by the reprocessing of organic matter by microbial trophic steps^{14,18}. The study of the trophic characteristics of micronektonic fishes with different migration habits and vertical distribution may shed some light on the increasing importance of the microbial system with trophic position and individual-size. To examine this hypothesis, we analysed isotope-based trophic indicators of micronektonic fishes in the central region of the N Atlantic to determine the overlap in their trophic niches, their trophic position, and their relative dependence of microbial trophic steps according to their migration habits and vertical distribution.

Methods

Sampling. Micronekton fishes were collected in the central N Atlantic during the BATHYPELAGIC cruise (Supplementary Fig. S1). Fish sampling was allowed by the Spanish authorities as part of a research project in international waters. No commercial species were collected and no experiments with living animals were made. Samples were obtained by using a midwater trawl of ca. 30-m² pelagic net equipped with a graded-mesh netting starting with 30 mm and ending with 4 mm²³ deployed at various depth layers (between the surface and ca. 1900 m) during day and night periods. In this study, samples were combined from different stations and layers to ensure a minimum of three specimens per species for isotopic analysis. Thirteen species of micronektonic fishes were analysed (Supplementary Table S1), including migrants reaching the surface (Neonesthes capensis, Benthosema glaciale, Lobianchia dofleini and Chauliodus danae), partial migrants reaching the subsurface layer up to 100 m (Argyropelecus hemigymnus and Photostomias guernei), and non-migrants always distributed below 250 m (Bathylagus euryops, Cyclothone alba, C. braueri, C. microdon, Sigmops bathyphilus, Scopelogadus beanii, and Taaningichthys bathyphilus). These species provided a range of individual sizes between 5 mg and 14 g of dry weight (17 to 275 mm in length). Information on migration habits, depth distribution, depth, and previous estimates of TP was obtained from the literature (see references in Table S1), our own data, and FishBase³². For subsequent analysis, the species were grouped according to three layers of vertical habitat: 0-1000 m, 1000-2000 m and > 2000 m (Supplementary Table S1).

Specimens were sorted, identified on board, and kept frozen (-20 °C). In the laboratory, each fish was first measured as standard length (SL, mm), eviscerated, freeze dried, and weighed (dry weight, DW, mg).

Stable isotope analysis. Portions of dorsal muscle (where possible) or whole individuals without gut and gonads (for small-sized specimens) were ground and homogenised, and subsequently analysed for $\delta^{15}N$ and $\delta^{13}C$ in bulk tissues. Separate determinations were made for each element, using untreated aliquots for $\delta^{15}N$, and lipid-extracted portions for $\delta^{13}C$. The latter were obtained by the removal of polar and neutral lipid fractions with a mixture of trichloromethane, methanol, and water³³. Determinations were made using an elemental analyser coupled to an isotope-ratio mass spectrometer and $\delta^{15}N$ and $\delta^{13}C$ were computed using international standards³⁴. For each analytical run, isotope standards from the International Atomic Energy Agency (IAEA-600, urea, and L-alanine), and internal standards (acetanilide and cyanobacteria of known isotope composition) were employed. Samples were analysed in triplicate. Precision (± s.e.) of replicate determinations of standards and samples for both isotopes was < 0.1 and <0.4‰, respectively.

Freeze-dried, 10 mg aliquots were also used for determinations of δ^{15} N in individual amino acids³⁵. Preparation of samples included hydrolysis with 6 N HCl (20 h, 110 °C) and the formation of trifluoroacetyl/isopropyl ester derivatives^{36,37}. An internal amino acid standard (L-norleucine) of known isotopic composition was added to each sample. Hydrolysed extracts were filtered by 0.20 µm hydrophilic filters and evaporated to dryness under N₂ at 60 °C. Esterification was made with 2.5 ml of 1:5 acetyl chloride:2-propanol, flushed with N₂ and heated to 110 °C for 60 min. For acylation, samples were first dried at ambient temperature under a stream of N₂, then heated to 110 °C for 15 min with 0.9 ml of 3:1 diclomethane:trifluoracetic anhydride (DCM:TFAA). Derivatized amino acids were purified by solvent extraction in 3 ml of 1:2 chloroform:phosphate buffer (Na₂HPO₄ + NaH₂PO₄ in Milli-Q water, pH 7.4). The chloroform and the acyl-derivatives were collected in a clean vial after centrifugation (10 min, 17,000×g) and evaporated at room temperature under N₂. Samples were stored at – 20 °C in 3:1 DCM:TFAA for up to 6 months until isotope analysis.

Derivatized products were injected into a mass spectrometer coupled to a gas chromatograph by a continuous flow interface and a combustion module. The individual amino acids were separated using a TraceGOLD TG-5MS chromatographic column (60 m, 0.32 mm ID, 1.0 μ m film). The δ^{15} N of each amino acid in the sample was calibrated with the values obtained for isolated standards analysed by combustion as described for bulk

analysis. Precision (± s.e.) of triplicate samples (two injections per sample) was < 1.5‰ per individual amino acid. The molar fraction of each amino acid was also determined along with δ^{15} N by calibration of the spectrometric signals with mixtures of amino acids using both certified and cyanobacterial standards, as for the bulk analysis³⁶. Values of δ^{15} N were obtained for alanine (Ala), glycine (Gly), threonine (Thr), serine (Ser), valine (Val), leucine (Leu), isoleucine (Ile), proline (Pro), methionine (Met), phenylalanine (Phe), lysine (Lys), and the mixtures of glutamine (Gln) and glutamic acid (Glu), and of aspartamine (Asn) and aspartic acid (Asp). The latter mixtures were caused by the acid hydrolysis and were termed as Glx and Asx, respectively. Following previous studies^{35,38,39} these amino acids were categorized as trophic (Glx, Asx, Ala, Ile, Leu, Pro and Val) and source (Gly, Ser, Lys, Phe, Thr and Met). The representativity of these amino acids for the bulk protein was determined by correlation of their weighted average δ^{15} N (δ^{15} N_{THAA}) with δ^{15} N of bulk samples, by taking into account the molar fraction of each individual amino acid³⁶. Further comparisons were made also using mean δ^{15} N values of trophic and source amino acids using both arithmetic mean and weighted mean values, but only the former were finally reported.

The size and overlap of the isotopic niche defined by bulk $\delta^{15}N$ and $\delta^{13}C$ of individual samples was computed with Bayesian statistics using the package NicheROVER⁴⁰. Because of the low number of samples in some of the combinations of migratory habits and layers, separate analyses for migrant vs. non-migrant and for depth layers were made using 10⁵ simulations. This analysis allowed the estimation of asymmetric probabilities of overlap between niche categories.

Trophic positions (TP) were estimated from CSIA results by taking into account the variability of trophic discrimination factors (TDF) accounting for the enrichment in δ^{15} N with each trophic step. The general multi-TDF model of McMahon and McCarthy³⁹ was applied to different trophic amino acids (Glx or Ala) and Phe as the source amino acid:

$$\Gamma P_{\rm Tr} = 2 + \left(\delta^{15} N_{\rm Tr} - \delta^{15} N_{\rm Phe} - \text{TDFp}_{\rm Tr} \beta_{\rm Tr}\right) / \text{TDFf}_{\rm Tr},\tag{1}$$

where subindex Tr refers to Glx or Ala, β is the difference between Tr (Glx or Ala) and Phe in primary producers, and TDFp_{Tr} and TDFf_{Tr} are the trophic discrimination factors for plankton and fish, respectively. TP_{Glx} is an estimate of the trophic position by taking into account only metazoan steps, while TP_{Ala} considers microbial + metazoan steps^{17,18}. Different β and TDF values (mean ± s.d.) were applied for plankton^{18,35}, and for teleost fish⁴¹ (β_{Glx} = 3.6 ± 0.5%; β_{Ala} = 3.2 ± 1.2%; TDFp_{Glx} = 7.6 ± 1.2%; TDFf_{Glx} = 5.7 ± 0.3%; TDFp_{Ala} = 4.5 ± 2.1%; TDFf_{Ala} = 6.1 ± 0.3‰).

The propagated error (s.d.) in the estimation of TP was calculated by taking into account the analytical errors in the individual trophic (Glx or Ala) and source amino acids (Phe), as well as the variability in the β and TDF values employed⁴¹. The fractional contribution of microbial steps to TP_{Ala} was computed as:

% Microbial =
$$100 (TP_{Ala} - TP_{Glx})/TP_{Ala}$$
. (2)

Alternatively, TP could have been estimated using a single TDF²⁹ or mean values for trophic and source amino acids^{16,41} but the multi-TDF model improved the correspondence between isotope-derived and gut-content derived TP³⁹. Moreover, the estimation of the microbial contribution would not have been possible when averaging trophic amino acids.

Differences between species grouped by migration habits and depth layers were assessed using PERMANOVA and non-parametric ANOVA (Kruskal–Wallis). Paired post-hoc tests (Bonferroni, Tukey) were applied to identify significant differences between groups. Regressions between variables were computed using product-moment regression (when there were large differences in their errors) or reduced major axis regression (when the errors in the variables were of similar magnitude). Statistical analyses were made using SPSS 17.0 (SPSS Inc.) and Past 4.0⁴².

Results

Niche overlap. Values of stable isotopes in bulk samples varied significantly between depth distribution layers, but not according to the migration habits (see Supplementary Tables S1, S2). Mean bulk δ^{15} N and δ^{13} C values for species distributed in the intermediate and deep layers were higher than those in the upper layer, with similar values for migrant and non-migrant species (Supplementary Fig. S2). Consequently, there was a large coincidence in the isotopic niche of migrants (and partial migrants) and non-migrants (Fig. 1a), and a much clearer separation of niches by depth layers (Fig. 1b). The estimated mean probability of overlap between the non-migrants niche with that of migrants exceeded 90%, and the probability of a migrant species to be found in the niche of non-migrants was ca. 80% (Supplementary Fig. S3a). In contrast, the niche of the species in the upper layer had mean overlap probabilities of <40% and <20% with the intermediate and deep layers, respectively (Supplementary Fig. S3b). Even lower values were obtained for the converse overlap. However, large overlap (>60%) was observed between the niche of the species living in the intermediate and deep layers.

Trophic and source amino acids. The values of $\delta^{15}N$ of total hydrolysable amino acids, as well as mean values for trophic and source amino acids, were significantly correlated with $\delta^{15}N$ in bulk samples, and these relationships were not affected by migration habits or layer distribution (Supplementary Fig. S4). The higher correlation values for $\delta^{15}N$ in trophic vs. source amino acids indicate that bulk $\delta^{15}N$ mainly reflected trophic positions.

As for bulk $\delta^{15}N$, there were no significant differences in mean values of $\delta^{15}N$ in trophic amino acids when species were grouped by migration habits (Supplementary Table S2), but non-migrant species had significantly higher values in source amino acids Gly and Phe (Fig. 2a, Supplementary Table S2). In contrast, $\delta^{15}N$ values for almost all trophic amino acids were higher in species from the intermediate and the deeper layers than in those from the upper layer (Fig. 2b). Source amino acids Gly and Phe also showed high $\delta^{15}N$ values in the deeper layers,



Figure 1. Plot of bulk δ^{15} N and δ^{13} C grouped by migratory (**a**) or depth layer distribution characteristics (**b**). The ellipses encompassing 95% of data for each group are indicated.





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while those for Ser and Thr were only significant for the deep or intermediate layer, respectively. In spite of these differences in source amino acids, there was no significant variation across the latitude gradient of samples, as exemplified by $\delta^{15}N_{Phe}$ (Supplementary Fig. S5).

Trophic positions. The studied species showed mean trophic positions between 2.56 and 4.36, when considering the full food web (TP_{Ala} , Fig. 3a,c). The lowest values were for the non-migrant, mesopelagic- dwelling, zooplanktivore *Cyclothone alba*, and the highest for the migrant, micronektivore *Neonesthes capensis*. Taking into account the microbial food web (TP_{Ala}) increased the mean estimations between 0.5 and 0.8 TP. These differences were higher than the errors of the estimations (mean values of 0.1 TP for both TP_{Ala} and TP_{Glx}) and imply contributions of microbial steps to the total TP between 14 and 21% (Fig. 3b,d). Migrant species and species distributed below 1000 m had significantly higher TP values than those of non-migrants and living in the upper layer, but the differences between TP_{Ala} and TP_{Glx} did not vary significantly with migration or between layers (Supplementary Table S3).

Both TP estimations increased significantly with the log of individual weight (Fig. 4a,b), but the slope of the regression was higher for TP_{Ala} than for TP_{Glx} , thus implying that the difference between both estimations



Figure 3. Mean (\pm s.d.) trophic positions (**a**,**c**) and microbial contribution (**b**,**d**) including microbial + metazoan (TP_{Ala}) or only metazoan (TP_{Glx}) food webs for migrant (**a**,**b**) or non-migrant (**c**,**d**) species coded by habitat depth. The values of integer TP are indicated by dashed lines.

also increased with body weight (Fig. 4c). However, when scaled to the TP_{Ala} values, the microbial contribution was neither related to body weight (Fig. 4d) or to other factors, such as species, taxonomic order, or diet (Supplementary Table S4). No significant effects of migration habits or depth layers were found to affect these relationships, either.

Discussion

This study shows that niche separation in micronektonic fishes is due to the vertical habitat range rather than to migration. The large niche overlap of migrant and non-migrant species can be expected because the feeding strategies of the species take advantage of different resources in each depth layer^{24,30,31}. In turn, differences in stable isotopes between depth layers can be attributed to the vertical separation of resources, as $\delta^{15}N$ and $\delta^{13}C$ increase with depth consistently with a major reworking of the settling organic matter⁴³. Previous studies also reported a vertical change in the source amino acids, and particularly in $\delta^{15}N_{Phe}$, for zooplankton⁴⁴ and micronektonic fishes⁴⁵, thus supporting the exploitation of resources in deep layers^{26,28,30,45}. In this context, vertical migration alleviates trophic competition by allowing the access to different food resources while the niche overlap was most likely between adjacent layers. The largest niche separation was found between the species distributed above and below 1000 m depth. The estimated trophic positions were also higher for species distributed in the deep layers irrespective of their migratory habits. These results suggest the existence of a major vertical limit for the influence of epipelagic vs. bathypelagic food webs. It can be argued that the high energetic costs and predation risks of migration from deep layers⁴⁶ must be compensated by the access to the abundant prey in the epipelagic layer. However, this requirement may be not always met in central areas of the ocean where the abundance of zooplankton is lower than near the coast^{9,47}. Indeed, there are indications of the existence of an active food web in bathypelagic waters supported by settling organic matter^{29,45,47,48} but also by in situ production by microorganisms⁴⁹⁻⁵¹. The latter implies an increase of the links between metazoan and microbial food webs that is partly supported by the observed isotopic enrichment in particles and organisms^{26,44,45}.

Our estimates of species-specific trophic position using amino acid δ^{15} N, but particularly TP_{Ala}, agree with those available in the literature and based on gut contents (see Supplementary Fig. S6). Previous studies suggested the use of multi-trophic isotopic models to improve the correspondence between both types of estimates to take into account the quality of the consumed food and the excretion products^{52,53}. In our study, we showed that the consideration of the microbial trophic steps produced TP values almost equivalent to those derived from stomach contents, while TP based only on the metazoan food web were systematically lower. In addition, three species (*N. capensis, Chauliodus danae*, and *Photostomias guernei*) considered as micronektivores because of the presence of small fish and crustaceans in their stomachs did not always have the highest TP when estimated from amino acid δ^{15} N. These results suggest that the diet of these species is far more complex and variable than indicated by gut content analysis, missing most of the components of the microbial food web. This calls for a review of the current assignation of trophic position values to prey items based on an established convention of TP = 1 for primary producers and detritus and TP = 2 for zooplankton^{54,55}. Indeed, it is possible that part of the



Figure 4. Relationships between trophic positions including (a) microbial + metazoan (TP_{Ala}) or (b) only metazoan (TP_{Glx}) food webs and (c) their difference ($TP_{Ala} - TP_{Glx}$) or (d) the fractional contribution of microbial food web (%microbial) with log-transformed individual dry weight (DW, mg). As the slopes for migrant (black dots) and non-migrant (red dots) were no significantly different (ANCOVA, P > 0.05), the regression lines for the whole data are indicated (P < 0.001) along with the corresponding determination coefficients (r^2).

apparent correspondence between TP estimates in FishBase and TP_{Ala} values would be a spurious result caused by the uncertainties in the assignation of TP to the identified prey in the former.

The calculated TP values are also within the range of estimates obtained using stable isotopes, even taking into account the uncertainties associated with the selection of appropriate baselines and trophic enrichment factors in case of bulk determinations. For instance, TP values for *Benthosema glaciale*, *Lobianchia dofleini*, *C. alba* and *Argyropelecus hemigymnus* were within the range estimated from bulk δ^{15} N for these species in the Atlantic^{27,56}, and those for *B. glaciale*, *A. hemigymnus* and *Cyclothone braueri* agreed with TP estimates in the Mediterranean²⁵. The main difference was for *L. dofleini*, that in the present study was ca. 1 TP lower, probably due to the smaller sizes of specimens analysed (32.0 mm in the Mediterranean vs. 17.7 mm in this study). Larger coincidence was found for TP estimations using also amino acid δ^{15} N values, as in the case of *A. hemigymnus* in the Gulf of Mexico²⁸. These results suggest that, despite regional differences in resource availability, most species show consistent trophic positions varying within relatively narrow limits. This is supported by the growing number of TP estimations based on δ^{15} N in amino acids, accounting for local and regional differences in the isotopic signature of the sources of nitrogen for primary producers^{26,28–30,45}.

In our study, migrant fishes had higher trophic position values than non-migrants, but both migrant and non-migrant species distributed in the bathypelagic layer had also higher TP than those in the upper layer. Other studies also noted this increase in TP with migration and depth distribution which was attributed to the access to a larger variety of prey and the predominance of piscivory in deep layers^{24,25,28,30}. Correspondingly, our metazoan TP estimates for species distributed in the bathypelagic layer were always above 2.5, indicating they were second-ary consumers, although the values for the gonostomatid *Cyclothone microdon* (mean \pm s.d. = 2.45 \pm 0.11) suggested a mixed diet including zooplankton, as observed in direct dietary analyses (see Supplementary Table S1). Among the stomiiforms studied we observed lower TP in the mesopelagic species, *C. alba* and *C. braueri*, than in the bathypelagic *C. microdon* and *Sigmops bathyphilus*. Our results also show that the reported higher TP in species of the family Stomiidae compared to those in Myctophidae²⁹ may not be a universal feature, and may depend on the species, sizes and depth range. For instance, we found TP > 3.5 for the three Stomiidae examined (*N. capensis, C. danae* and *P. guernei*), while the bathypelagic myctophid *Taaningichthys bathyphilus* (for which there is no published data on stomach content) had a mean \pm s.d. TP of 4.07 \pm 0.09. The comparison between

our TP estimates with those derived from diet analysis also revealed that those species ranked as piscivores (see Supplementary Table S1) were not always those with the highest TP (e.g. *Chauliodus danae*), thus reflecting a large plasticity in the diets of these fishes. As the composition of the micronekton community is more complex than suggested by the simplification to a few groups^{23,57}, we also include representatives of orders Stephanoberyciformes (*Scopelogadus beanii*) and Argentiniformes (*Bathylagus euryops*) among the selected species. These species are consumers of gelatinous plankton (see Supplementary Table S1) and ther estimated TP, ca. 3, is similar to those for other species consuming prey richer in protein than gelatinous plankton. Thus, these relatively high values may in fact reflect the deep habitat of the species, which may be the case of *S. beanii*, reported to feed on salps⁵⁸ or they could also be explained by the consumption of predatory prey, since species of *Bathylagus* showed a substantial fraction of siphonophore DNA in their stomachs⁵⁹.

Nevertheless, the estimations of trophic position based only on metazoan steps were lower than those considering also microbial steps with an average microbial contribution to overall TP of 17%. Some species (as those of genus Cyclothone) had relatively large variation in the microbial contributions that could be attributed to their feeding on detrital aggregates (see Supplementary Table S1). These results are quite similar to the only published estimations of microbial contributions to pelagic food webs, which use the same methodology, but applied to epipelagic zooplankton¹⁸. These, indicate microbial contributions up to 35% in omnivorous species and between 0 and 11% for herbivores. Fatty acid markers also revealed a significant dietary contribution of bacterial organic matter to some subtropical copepods⁴⁸. The prevalence of microbial contributions in planktivores and piscivores found in this study highlights the importance of detritus degradation and microbial reworking particularly in deep layers of the ocean. In the absence of phytoplankton, a better use of the organic matter in the bathypelagic domain can be made through trophic steps involving chemoautolithotrophy^{45,50}, uptake of dissolved organic matter by bacteria⁵¹, and protozoan grazing on bacteria^{8,60}, among those within the microbial food web. Further transfer of the organic matter concentrated by the microbial food web to the metazoan consumers is made by zooplankton^{61,62} and, as shown in our study, by micronekton fishes. Even when the main contribution to the top consumers of the food web (e.g. sharks, tunas, birds, cetaceans) requires a substantial contribution of direct transfers of primary production through the metazoan food web^{2,54}, the estimations made in our study suggest that, at least in the oligotrophic regions of the ocean, the contribution of the microbial food web cannot be neglected.

The increase in amino acid based TP with the logarithm of individual size confirms that pelagic food webs are size-structured⁶³, as already found in studies using bulk $\delta^{15}N^{12,64-66}$. This structure implies that the trophic efficiency decreases with size as the ratio between the sizes of predator and prey increases⁶⁷ thus placing an upper limit to the number of trophic levels that can be sustained in a food web^{68,69}. Meta-analysis of aquatic food web studies concluded that marine pelagic ecosystems, even having a higher number of trophic levels than freshwater ecosystems, will not have more than 5 trophic levels^{68,70}. The required energy conversions and losses in models based on biomass propagation may lead to the underrepresentation of multiple or alternative trophic pathways. However, other analyses revealed that top predators TP were underestimated, notably by conventional models based on bulk $\delta^{15}N^{15}$. In addition, estimations using the biomass spectrum suggested that the total number of trophic levels could increase because of the omnivorous and opportunistic feeding of zooplankton⁷¹. Zooplankton, indeed, represents the link between the microbial and metazoan food webs supporting higher trophic levels and vertical export⁶². The addition of more trophic levels by taking into account the microbial steps supports the increase of TP estimates, at least for zooplankton¹⁸ and micronekton (this study).

Taking into account microbial steps, TP of the micronekton fishes studied increased linearly with the logarithm of body weight. However, we did not find significant effects for size, taxonomical categories, migration habits or vertical distribution on the relative importance of the microbial food web when scaled to TP values. Our estimates varied between relatively narrow limits (9–25%) comparable to those reported for zooplankton¹⁸ and can be attributed to the widespread omnivory of these organisms. In the case of crustacean zooplankton there was a clear increase in the microbial contribution in omnivores, but the absence of a clear pattern in planktivorous and piscivorous fishes suggests the existence of an upper limit to the contribution of the microbial steps to the trophic position. In fact, food webs may exhibit a high number of trophic links with low energetical importance while only a small number of links influence the transfer of energy and biomass to upper trophic levels^{69,72}. Besides, the growth efficiency of pelagic bacteria is generally low in the deep ocean because of large respiratory losses^{4,73}. In this way, we could expect that the contribution of metazoan components to the overall TP should increase for the upper trophic levels. However, further studies including other organisms from the food web will be required to confirm this hypothesis. Our study showed that TP estimations of mid-trophic level consumers can be improved by taking into account the contribution of microbial trophic links, suggesting that current estimations of TP and food chain length need to be revised.

Data availability

The original data on sample location, individual fish characteristics and stable isotope composition, including amino acids, can be accessed through the PANGAEA repository (https://doi.org/10.1594/PANGAEA.930111).

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References

- 1. Azam, F. et al. The ecological role of water-column microbes in the sea. Mar. Ecol. Prog. Ser. 10, 257-263 (1983).
- 2. Legendre, L. & Le Fèvre, J. Microbial food webs and the export of biogenic carbon in oceans. Aquat. Microb. Ecol. 9, 69–77 (1995).
- 3. Legendre, L. & Rivkin, R. B. Planktonic food webs: Microbial hub approach. Mar. Ecol. Prog. Ser. 365, 289–309 (2008).
- Arfstegui, J., Gasol, J. M., Duarte, C. M. & Herndl, G. J. Microbial oceanography of the dark ocean's pelagic realm. *Limnol. Oceanogr.* 54, 1501–1529 (2009).

- Roshan, S. & DeVries, T. Efficient dissolved organic carbon production and export in the oligotrophic ocean. Nat. Commun. 8, 2036. https://doi.org/10.1038/s41467-017-02227-3 (2017).
- 6. Pauly, D. & Christensen, V. Primary production required to sustain global fisheries. Nature 374, 255-257 (1995).
- 7. Sarmiento, J. L. & Gruber, N. Ocean Biogeochemical Dynamics (Princeton University Press, 2006).
- Armengol, L., Calbet, A., Franchy, G., Rodríguez-Santos, A. & Hernández-León, S. Planktonic food web structure and trophic transfer efficiency along a productivity gradient in the tropical and subtropical Atlantic Ocean. *Sci. Rep.* 9, 2044. https://doi.org/ 10.1038/s41598-019-38507-9 (2019).
- Hernández-León, S. et al. Large deep-sea zooplankton biomass mirrors primary production in the global ocean. Nat. Commun. 11, 6048. https://doi.org/10.1038/s41467-020-19875-7 (2020).
- 10. Wilson, R. et al. Contribution of fish to the marine inorganic carbon cycle. Science 323, 359-362 (2009).
- Jennings, S. & van der Molen, J. Trophic levels of marine consumers from nitrogen stable isotope analysis: Estimation and uncertainty. *ICES J. Mar. Sci.* 72, 2289–2300 (2015).
- Jennings, S., Maxwell, T. A. D., Schratzberger, M. & Milligan, S. P. Body-size dependent temporal variations in nitrogen stable isotope ratios in food webs. *Mar. Ecol. Prog. Ser.* 370, 199–206 (2008).
- Bernal, A., Olivar, M. P., Maynou, F. & de Puelles, M. L. F. Diet and feeding strategies of mesopelagic fishes in the western Mediterranean. Prog. Oceanogr. 135, 1–17 (2015).
- 14. Gutiérrez-Rodríguez, A., Décima, M., Popp, B. N. & Landry, M. R. Isotopic invisibility of protozoan trophic steps in marine food webs. *Limnol. Oceanogr.* **59**, 1590–1598 (2014).
- 15. Hussey, N. E. et al. Rescaling the trophic structure of marine food webs. Ecol. Lett. 17, 239-250 (2014).
- Nielsen, J. M., Popp, B. N. & Winder, M. Meta-analysis of amino acid stable nitrogen isotope ratios for estimating trophic position in marine organisms. *Oecologia* 178, 631–642 (2015).
- Décima, M., Landry, M. R., Bradley, C. J. & Fogel, M. L. Alanine δ¹⁵N trophic fractionation in heterotrophic protists. *Limnol. Oceanogr.* 62, 2308–2322 (2017).
- Décima, M. & Landry, M. Resilience of plankton trophic structure to an eddy-stimulated diatom bloom in the North Pacific Subtropical Gyre. *Mar. Ecol. Prog. Ser.* 643, 33–48 (2020).
- Irigoien, X. et al. Large mesopelagic fish biomass and trophic efficiency in the Open Ocean. Nat. Commun. 5, 3271. https://doi. org/10.1038/ncomms4271 (2014).
- Cherel, Y., Fontaine, C., Richard, P. & Labat, J.-P. Isotopic niches and trophic levels of myctophid fishes and their predators in the Southern Ocean. *Limnol. Oceanogr.* 55, 324–332 (2010).
- Young, J. W. et al. Setting the stage for a global-scale trophic analysis of marine top predators: A multi-workshop review. Rev. Fish Biol. Fish. 25, 261–272 (2015).
- Klevjer, T. A. et al. Large scale patterns in vertical distribution and behaviour of mesopelagic scattering layers. Sci. Rep. 6, 19873. https://doi.org/10.1038/srep19873 (2016).
- Olivar, M. P. et al. Mesopelagic fishes across the tropical and equatorial Atlantic: Biogeographical and vertical patterns. Prog. Oceanogr. 151, 116–137 (2017).
- Eduardo, L. N. *et al.* Trophic ecology, habitat, and migratory behaviour of the viperfish *Chauliodus sloani* reveal a key mesopelagic player. *Sci. Rep.* 10, 20996. https://doi.org/10.1038/s41598-020-77222-8 (2020).
- Valls, M. et al. Trophic structure of mesopelagic fishes in the western Mediterranean based on stable isotopes of carbon and nitrogen. J. Mar. Syst. 138, 160–170 (2014).
- Choy, C. A., Popp, B. N., Hannides, C. C. S. & Drazen, J. C. Trophic structure and food resources of epipelagic and mesopelagic fishes in the North Pacific Subtropical Gyre ecosystem inferred from nitrogen isotopic compositions. *Limnol. Oceanogr.* 60, 1156–1171 (2015).
- Olivar, M. P., Bode, A., López-Pérez, C., Hulley, P. A. & Hernández-León, S. Trophic position of lanternfishes (Pisces: Myctophidae) of the tropical and equatorial Atlantic estimated using stable isotopes. *ICES J. Mar. Sci.* 76, 649–661 (2019).
- Richards, T. M., Sutton, T. T. & Wells, R. J. D. Trophic structure and sources of variation influencing the stable isotope signatures of meso- and bathypelagic micronekton fishes. *Front. Mar. Sci.* 7, 507992. https://doi.org/10.3389/fmars.2020.507992 (2020).
- Choy, C. A. et al. Global trophic position comparison of two dominant mesopelagic fish families (Myctophidae, Stomiidae) using amino acid nitrogen isotopic analyses. PLoS ONE 7, e50133. https://doi.org/10.1371/journal.pone.0050133 (2012).
- Wang, F. et al. Trophic interactions of mesopelagic fishes in the South China Sea illustrated by stable isotopes and fatty acids. Front. Mar. Sci. 5, 522. https://doi.org/10.3389/fmars.2018.00522 (2019).
- Czudaj, S. *et al.* Spatial variation in the trophic structure of micronekton assemblages from the eastern tropical North Atlantic in two regions of differing productivity and oxygen environments. *Deep Sea Res.* 163, 103275. https://doi.org/10.1016/j.dsr.2020. 103275 (2020).
- 32. FishBase. A Global Information System on Fishes (University of California, 2021).
- Bligh, E. G. & Dyer, W. J. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.* 37, 911–917 (1959).
 Coplen, T. B. Guidelines and recommended terms for expression of stable isotope-ratio and gas-ratio measurement results. *Rapid*
- Commun. Mass Spectrom. 25, 2538–2560 (2011).
- Chikaraishi, Y. et al. Determination of aquatic food-web structure based on compound-specific nitrogen isotopic composition of amino acids. Limnol. Oceanogr. Methods 7, 740–750 (2009).
- McCarthy, M. D., Lehman, J. & Kudela, R. Compound-specific amino acid δ15N patterns in marine algae: Tracer potential for cyanobacterial vs. eukaryotic organic nitrogen sources in the ocean. *Geochim. Cosmochim. Acta* 103, 104–120 (2013).
- Mompeán, C., Bode, A., Gier, E. & McCarthy, M. D. Bulk vs. aminoacid stable N isotope estimations of metabolic status and contributions of nitrogen fixation to size-fractionated zooplankton biomass in the subtropical N Atlantic. *Deep Sea Res.* 114, 137–148 (2016).
- McClelland, J. W. & Montoya, J. P. Trophic relationships and the nitrogen isotopic composition of amino acids in plankton. *Ecology* 83, 2173–2180 (2002).
- McMahon, K. W. & McCarthy, M. D. Embracing variability in amino acid d15N fractionation: Mechanisms, implications, and applications for trophic ecology. *Ecosphere* 7, e01511. https://doi.org/10.1002/ecs2.1511 (2016).
- 40. Swanson, H. K. *et al.* A new probabilistic method for quantifying n-dimensional ecological niches and niche overlap. *Ecology* **96**, 318–324 (2015).
- Bradley, C. J. et al. Trophic position estimates of marine teleosts using amino acid compound specific isotopic analysis. Limnol. Oceanogr. Methods 13, 476–493 (2015).
- Hammer, Ø., Harper, D. A. T. & Ryan, P. D. PAST: Paleontological statistics software package for education and data analysis. *Palaeontol. Electron.* 4, 1–9 (2001).
- McCarthy, M. D., Benner, R., Lee, C. & Fogel, M. L. Amino acid nitrogen isotopic fractionation patterns as indicators of heterotrophy in plankton, particulate, and dissolved organic matter. *Geochim. Cosmochim. Acta* 71, 4727–4744 (2007).
- Hannides, C. C. S., Popp, B. N., Choy, C. A. & Drazen, J. C. Midwater zooplankton and suspended particle dynamics in the North Pacific Subtropical Gyre: A stable isotope perspective. *Limnol. Oceanogr.* 58, 1931–1946 (2013).
- 45. Gloeckler, K. *et al.* Stable isotope analysis of micronekton around Hawaii reveals suspended particles are an important nutritional source in the lower mesopelagic and upper bathypelagic zones. *Limnol. Oceanogr.* **63**, 1168–1180 (2018).
- 46. Robison, B. H. Conservation of deep pelagic biodiversity. Conserv. Biol. 23, 847-858 (2009).

⁸

- 47. Brun, P. et al. Climate change has altered zooplankton-fuelled carbon export in the North Atlantic. Nat. Ecol. Evol. 3, 416–423 (2019).
- Bode, M. et al. Feeding strategies of tropical and subtropical calanoid copepods throughout the eastern Atlantic Ocean: Latitudinal and bathymetric aspects. Prog. Oceanogr. 138, 268–282 (2015).
- Herndl, G. J. *et al.* Contribution of Archaea to total prokarytic production in the deep Atlantic Ocean. *Appl. Environ. Microbiol.* 71, 2303–2309 (2005).
- Varela, M. M., van Aken, H. M., Sintes, E., Reinthaler, T. & Herndl, G. J. Contribution of Crenarchaeota and bacteria to autotrophy in the North Atlantic interior. *Environ. Microbiol.* 13, 1524–1533 (2011).
- Clifford, E. L. *et al.* Taurine is a major carbon and energy source for marine prokaryotes in the North Atlantic Ocean off the Iberian Peninsula. *Microb. Ecol.* 78, 299–312 (2019).
- Hoen, D. K. et al. Amino acid ¹⁵N trophic enrichment factors of four large carnivorous fishes. J. Exp. Mar. Biol. Ecol. 453, 76–83 (2014).
- 53. McMahon, K. W. & McCarthy, M. D. Embracing variability in amino acid δ^{15} N fractionation: Mechanisms, implications, and applications for trophic ecology. *Ecosphere* 7, e01511. https://doi.org/10.1002/ecs2.1511 (2016).
- Pauly, D., Christensen, V. & Walters, C. Ecopath, ecosim, and ecospace as tools for evaluating ecosystem impact of fisheries. ICES J. Mar. Sci. 57, 697–706 (2000).
- 55. Christensen, V. & Walters, C. Ecopath with ecosim: Methods, capabilities and limitations. Ecol. Model. 172, 109–139 (2004).
- McClain-Counts, J. P., Demopoulos, A. W. J. & Ross, S. W. Trophic structure of mesopelagic fishes in the Gulf of Mexico revealed by gut content and stable isotope analyses. *Mar. Ecol.* 38, e12449. https://doi.org/10.1111/maec.12449 (2017).
- 57. Olivar, M. P. et al. The contribution of migratory mesopelagic fishes to neuston fish assemblages across the Atlantic, Indian and Pacific Oceans. Mar. Freshw. Res. 67, 1114–1127 (2016).
- Gartner, J. V. & Musick, J. A. Feeding habits of the deep-sea fish, *Scopelogadus beanii* (Pisces: Melamphaide), in the western North Atlantic. *Deep Sea Res. A Oceanogr. Res. Pap.* 36(10), 1457–1469. https://doi.org/10.1016/0198-0149(89)90051-4 (1989).
- Clarke, L. J., Trebilco, R., Walters, A., Polanowski, A. M. & Deagle, B. E. DNA-based diet analysis of mesopelagic fish from the southern Kerguelen axis. Deep-Sea Res. II Top. Stud. Oceanogr. https://doi.org/10.1016/j.dsr2.2018.09.001 (2020).
- Schmoker, C., Hernández-León, S. & Calbet, A. Microzooplankton grazing in the oceans: Impacts, data variability, knowledge gaps and future directions. J. Plankton Res. 35, 691–706 (2013).
- 61. Calbet, A. & Saiz, E. The ciliate-copepod link in marine ecosystems. Aquat. Microb. Ecol. 38, 157-167 (2005).
- 62. Zeldis, J. R. & Décima, M. Mesozooplankton connect the microbial food web to higher trophic levels and vertical export in the New Zealand subtropical convergence zone. *Deep Sea Res.* 155, 103146 (2020).
- Jennings, S., Pinnegar, J. K., Polunin, N. V. C. & Boon, T. W. Weak cross-species relationships between body size and trophic level belie powerful size-based trophic structuring in fish communities. J. Anim. Ecol. 70, 934–944 (2001).
- Bode, A., Carrera, P. & Lens, S. The pelagic foodweb in the upwelling ecosystem of Galicia (NW Spain) during spring: Natural abundance of stable carbon and nitrogen isotopes. *ICES J. Mar. Sci.* 60, 11–22 (2003).
- 65. Hunt, B. P. V. et al. A coupled stable isotope-size spectrum approach to understanding pelagic food-web dynamics: A case study from the southwest sub-tropical Pacific. Deep Sea Res. 113, 208-224 (2015).
- Romero-Romero, S., Molina-Ramírez, A., Höfer, J. & Acuña, J. L. Body size-based trophic structure of a deep marine ecosystem. Ecology 97, 171–181 (2016).
- Barnes, C., Maxwell, D., Reuman, D. C. & Jennings, S. Global patterns in predator-prey size relationships reveal size dependency of trophic transfer efficiency. *Ecology* 91, 222–232 (2010).
- 68. Schoener, T. W. Food webs from the small to the large. Ecology 70, 1559-1589 (1989).
- 69. Zhou, M. What determines the slope of a plankton biomass spectrum?. J. Plankton Res. 28, 437-448 (2006).
- 70. Van der Zanden, M. J. & Fetzer, W. Global patterns of aquatic food chain length. Oikos 116, 1378-1388 (2007).
- Basedow, S. L., de Silva, N. A. L., Bode, A. & van Beusekorn, J. Trophic positions of mesozooplankton across the North Atlantic: estimates derived from biovolume spectrum theories and stable isotope analyses. J. Plankton Res. 38, 1364–1378 (2016).
- Williams, R. J. & Martinez, N. D. Limits to trophic levels and omnivory in complex food webs: Theory and data. Am. Nat. 163, 458–468 (2004).
- 73. Nagata, T. *et al.* Emerging concepts on microbial processes in the bathypelagic ocean ecology, biogeochemistry, and genomics. *Deep Sea Res. II* 57, 1519–1536 (2010).

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Author contributions

S.H.L. designed the project. A.B. and M.P.O. conceived this specific research, obtained the samples, and analysed the data. A.B. wrote the manuscript with contributions from S.H.L. and M.P.O.

Competing interests

The authors declare no competing interests.

Additional information

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Correspondence and requests for materials should be addressed to A.B.

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